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[Continued on next page]

(54) Title: TRANSPORTERS AND ION CHANNELS

(57) Abstract: The invention provides human transporters and ion channels (TRICH) and polynucleotides which identify and encode TRICH. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of TRICH.



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TRANSPORTERS AND ION CHANNELS

TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of transporters and ion channels and to the use of these sequences in the diagnosis, treatment, and prevention of transport, neurological, muscle, immunological, and cell proliferative disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

BACKGROUND OF THE INVENTION

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Eukaryotic cells are surrounded and subdivided into functionally distinct organelles by hydrophobic lipid bilayer membranes which are highly impermeable to most polar molecules. Cells and organelles require transport proteins to import and export essential nutrients and metal ions including K⁺, NH₄⁺, P_i, SO₄², sugars, and vitamins, as well as various metabolic waste products. Transport proteins also play roles in antibiotic resistance, toxin secretion, ion balance, synaptic neurotransmission, kidney function, intestinal absorption, tumor growth, and other diverse cell functions (Griffith, J. and C. Sansom (1998) The Transporter Facts Book, Academic Press, San Diego CA, pp. 3-29). Transport can occur by a passive concentration-dependent mechanism, or can be linked to an energy source such as ATP hydrolysis or an ion gradient. Proteins that function in transport include carrier proteins, which bind to a specific solute and undergo a conformational change that translocates the bound solute across the membrane, and channel proteins, which form hydrophilic pores that allow specific solutes to diffuse through the membrane down an electrochemical solute gradient.

Carrier proteins which transport a single solute from one side of the membrane to the other are called uniporters: In contrast, coupled transporters link the transfer of one solute with simultaneous or sequential transfer of a second solute, either in the same direction (symport) or in the opposite direction (antiport). For example, intestinal and kidney epithelium contains a variety of symporter systems driven by the sodium gradient that exists across the plasma membrane. Sodium moves into the cell down its electrochemical gradient and brings the solute into the cell with it. The sodium gradient that provides the driving force for solute uptake is maintained by the ubiquitous Na⁺/K⁺ ATPase system. Sodium-coupled transporters include the mammalian glucose transporter (SGLT1), iodide transporter (NIS), and multivitamin transporter (SMVT). All three transporters have twelve putative transmembrane segments, extracellular glycosylation sites, and cytoplasmically-oriented N- and C-termini. NIS plays a crucial role in the evaluation, diagnosis, and treatment of various thyroid pathologies because it is the molecular basis for radioiodide thyroid-imaging

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techniques and for specific targeting of radioisotopes to the thyroid gland (Levy, O. et al. (1997) Proc. Natl. Acad. Sci. USA 94:5568-5573). SMVT is expressed in the intestinal mucosa, kidney, and placenta, and is implicated in the transport of the water-soluble vitamins, e.g., biotin and pantothenate (Prasad, P.D. et al. (1998) J. Biol. Chem. 273:7501-7506).

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One of the largest families of transporters is the major facilitator superfamily (MFS), also called the uniporter-symporter-antiporter family. MFS transporters are single polypeptide carriers that transport small solutes in response to ion gradients. Members of the MFS are found in all classes of living organisms, and include transporters for sugars, oligosaccharides, phosphates, nitrates, nucleosides, monocarboxylates, and drugs. MFS transporters found in eukaryotes all have a structure comprising 12 transmembrane segments (Pao, S.S. et al. (1998) Microbiol. Molec. Biol. Rev. 62:1-34). The largest family of MFS transporters is the sugar transporter family, which includes the seven glucose transporters (GLUT1-GLUT7) found in humans that are required for the transport of glucose and other hexose sugars. These glucose transport proteins have unique tissue distributions and physiological functions. GLUT1 provides many cell types with their basal glucose requirements and transports glucose across epithelial and endothelial barrier tissues; GLUT2 facilitates glucose uptake or efflux from the liver; GLUT3 regulates glucose supply to neurons; GLUT4 is responsible for insulin-regulated glucose disposal; and GLUT5 regulates fructose uptake into skeletal muscle. Defects in glucose transporters are involved in a recently identified neurological syndrome causing infantile seizures and developmental delay, as well as glycogen storage disease, Fanconi-Bickel syndrome, and non-insulin-dependent diabetes mellitus (Mueckler, M. (1994) Eur. J. Biochem. 219:713-725; Longo, N. and L.J. Elsas (1998) Adv. Pediatr. 45:293-313).

Monocarboxylate anion transporters are proton-coupled symporters with a broad substrate specificity that includes L-lactate, pyruvate, and the ketone bodies acetate, acetoacetate, and beta-hydroxybutyrate. At least seven isoforms have been identified to date. The isoforms are predicted to have twelve transmembrane (TM) helical domains with a large intracellular loop between TM6 and TM7, and play a critical role in maintaining intracellular pH by removing the protons that are produced stoichiometrically with lactate during glycolysis. The best characterized H*-monocarboxylate transporter is that of the erythrocyte membrane, which transports L-lactate and a wide range of other aliphatic monocarboxylates. Other cells possess H*-linked monocarboxylate transporters with differing substrate and inhibitor selectivities. In particular, cardiac muscle and tumor cells have transporters that differ in their K_m values for certain substrates, including stereoselectivity for L- over D-lactate, and in their sensitivity to inhibitors. There are Na*-monocarboxylate cotransporters on the luminal surface of intestinal and kidney epithelia, which allow the uptake of lactate, pyruvate, and ketone bodies in these tissues. In addition, there are specific and selective transporters for organic cations and organic anions in organs including the

kidney, intestine and liver. Organic anion transporters are selective for hydrophobic, charged molecules with electron-attracting side groups. Organic cation transporters, such as the ammonium transporter, mediate the secretion of a variety of drugs and endogenous metabolites, and contribute to the maintenance of intercellular pH (Poole, R.C. and A.P. Halestrap (1993) Am. J. Physiol. 264:C761-C782; Price, N.T. et al. (1998) Biochem. J. 329:321-328; and Martinelle, K. and L. Haggstrom (1993) J. Biotechnol. 30:339-350).

ATP-binding cassette (ABC) transporters are members of a superfamily of membrane proteins that transport substances ranging from small molecules such as ions, sugars, amino acids, peptides, and phospholipids, to lipopeptides, large proteins, and complex hydrophobic drugs. ABC transporters consist of four modules: two nucleotide-binding domains (NBD), which hydrolyze ATP to supply the energy required for transport, and two membrane-spanning domains (MSD), each containing six putative transmembrane segments. These four modules may be encoded by a single gene, as is the case for the cystic fibrosis transmembrane regulator (CFTR), or by separate genes. When encoded by separate genes, each gene product contains a single NBD and MSD. These "halfmolecules" form homo- and heterodimers, such as Tap1 and Tap2, the endoplasmic reticulum-based major histocompatibility (MHC) peptide transport system. Several genetic diseases are attributed to defects in ABC transporters, such as the following diseases and their corresponding proteins: cystic fibrosis (CFTR, an ion channel), adrenoleukodystrophy (adrenoleukodystrophy protein, ALDP), Zellweger syndrome (peroxisomal membrane protein-70, PMP70), and hyperinsulinemic hypoglycemia (sulfonylurea receptor, SUR). Overexpression of the multidrug resistance (MDR) protein, another ABC transporter, in human cancer cells makes the cells resistant to a variety of cytotoxic drugs used in chemotherapy (Taglicht, D. and S. Michaelis (1998) Meth. Enzymol. 292:130-162).

A number of metal ions such as iron, zinc, copper, cobalt, manganese, molybdenum, selenium, nickel, and chromium are important as cofactors for a number of enzymes. For example, copper is involved in hemoglobin synthesis, connective tissue metabolism, and bone development, by acting as a cofactor in oxidoreductases such as superoxide dismutase, ferroxidase (ceruloplasmin), and lysyl oxidase. Copper and other metal ions must be provided in the diet, and are absorbed by transporters in the gastrointestinal tract. Plasma proteins transport the metal ions to the liver and other target organs, where specific transporters move the ions into cells and cellular organelles as needed. Imbalances in metal ion metabolism have been associated with a number of disease states (Danks, D.M. (1986) J. Med. Genet. 23:99-106).

Transport of fatty acids across the plasma membrane can occur by diffusion, a high capacity, low affinity process. However, under normal physiological conditions a significant fraction of fatty acid transport appears to occur via a high affinity, low capacity protein-mediated transport process.

Fatty acid transport protein (FATP), an integral membrane protein with four transmembrane segments, is expressed in tissues exhibiting high levels of plasma membrane fatty acid flux, such as muscle, heart, and adipose. Expression of FATP is upregulated in 3T3-L1 cells during adipose conversion, and expression in COS7 fibroblasts elevates uptake of long-chain fatty acids (Hui, T.Y. et al. (1998) J. Biol. Chem. 273:27420-27429).

Mitochondrial carrier proteins are transmembrane-spanning proteins which transport ions and charged metabolites between the cytosol and the mitochondrial matrix. Examples include the ADP, ATP carrier protein; the 2-oxoglutarate/malate carrier; the phosphate carrier protein; the pyruvate carrier; the dicarboxylate carrier which transports malate, succinate, fumarate, and phosphate; the tricarboxylate carrier which transports citrate and malate; and the Grave's disease carrier protein, a protein recognized by IgG in patients with active Grave's disease, an autoimmune disorder resulting in hyperthyroidism. Proteins in this family consist of three tandem repeats of an approximately 100 amino acid domain, each of which contains two transmembrane regions (Stryer, L. (1995)

Biochemistry, W.H. Freeman and Company, New York NY, p. 551; PROSITE PDOC00189

Mitochondrial energy transfer proteins signature; Online Mendelian Inheritance in Man (OMIM)

*275000 Graves Disease).

This class of transporters also includes the mitochondrial uncoupling proteins, which create proton leaks across the inner mitochondrial membrane, thus uncoupling oxidative phosphorylation from ATP synthesis. The result is energy dissipation in the form of heat. Mitochondrial uncoupling proteins have been implicated as modulators of thermoregulation and metabolic rate, and have been proposed as potential targets for drugs against metabolic diseases such as obesity (Ricquier, D. et al. (1999) J. Int. Med. 245:637-642).

Ion Channels

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The electrical potential of a cell is generated and maintained by controlling the movement of ions across the plasma membrane. The movement of ions requires ion channels, which form ion-selective pores within the membrane. There are two basic types of ion channels, ion transporters and gated ion channels. Ion transporters utilize the energy obtained from ATP hydrolysis to actively transport an ion against the ion's concentration gradient. Gated ion channels allow passive flow of an ion down the ion's electrochemical gradient under restricted conditions. Together, these types of ion channels generate, maintain, and utilize an electrochemical gradient that is used in 1) electrical impulse conduction down the axon of a nerve cell, 2) transport of molecules into cells against concentration gradients, 3) initiation of muscle contraction, and 4) endocrine cell secretion.

Ion Transporters

Ion transporters generate and maintain the resting electrical potential of a cell. Utilizing the energy derived from ATP hydrolysis, they transport ions against the ion's concentration gradient.

These transmembrane ATPases are divided into three families. The phosphorylated (P) class ion transporters, including Na⁺-K⁺ ATPase, Ca²⁺-ATPase, and H⁺-ATPase, are activated by a phosphorylation event. P-class ion transporters are responsible for maintaining resting potential distributions such that cytosolic concentrations of Na⁺ and Ca²⁺ are low and cytosolic concentration of K⁺ is high. The vacuolar (V) class of ion transporters includes H⁺ pumps on intracellular organelles, such as lysosomes and Golgi. V-class ion transporters are responsible for generating the low pH within the lumen of these organelles that is required for function. The coupling factor (F) class consists of H⁺ pumps in the mitochondria. F-class ion transporters utilize a proton gradient to generate ATP from ADP and inorganic phosphate (P_i).

The P-ATPases are hexamers of a 100 kD subunit with ten transmembrane domains and several large cytoplasmic regions that may play a role in ion binding (Scarborough, G.A. (1999) Curr. Opin. Cell Biol. 11:517-522). The V-ATPases are composed of two functional domains: the V₁ domain, a peripheral complex responsible for ATP hydrolysis; and the V₀ domain, an integral complex responsible for proton translocation across the membrane. The F-ATPases are structurally and evolutionarily related to the V-ATPases. The F-ATPase F₀ domain contains 12 copies of the c subunit, a highly hydrophobic protein composed of two transmembrane domains and containing a single buried carboxyl group in TM2 that is essential for proton transport. The V-ATPase V₀ domain contains three types of homologous c subunits with four or five transmembrane domains and the essential carboxyl group in TM4 or TM3. Both types of complex also contain a single a subunit that may be involved in regulating the pH dependence of activity (Forgac, M. (1999) J. Biol. Chem. 274:12951-12954).

The resting potential of the cell is utilized in many processes involving carrier proteins and gated ion channels. Carrier proteins utilize the resting potential to transport molecules into and out of the cell. Amino acid and glucose transport into many cells is linked to sodium ion co-transport (symport) so that the movement of Na⁺ down an electrochemical gradient drives transport of the other molecule up a concentration gradient. Similarly, cardiac muscle links transfer of Ca²⁺ out of the cell with transport of Na⁺ into the cell (antiport).

Gated Ion Channels

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Gated ion channels control ion flow by regulating the opening and closing of pores. The ability to control ion flux through various gating mechanisms allows ion channels to mediate such diverse signaling and homeostatic functions as neuronal and endocrine signaling, muscle contraction, fertilization, and regulation of ion and pH balance. Gated ion channels are categorized according to the manner of regulating the gating function. Mechanically-gated channels open their pores in response to mechanical stress; voltage-gated channels (e.g., Na⁺, K⁺, Ca²⁺, and Cl⁻ channels) open their pores in response to changes in membrane potential; and ligand-gated channels (e.g.,

acetylcholine-, serotonin-, and glutamate-gated cation channels, and GABA- and glycine-gated chloride channels) open their pores in the presence of a specific ion, nucleotide, or neurotransmitter. The gating properties of a particular ion channel (i.e., its threshold for and duration of opening and closing) are sometimes modulated by association with auxiliary channel proteins and/or post translational modifications, such as phosphorylation.

Mechanically-gated or mechanosensitive ion channels act as transducers for the senses of touch, hearing, and balance, and also play important roles in cell volume regulation, smooth muscle contraction, and cardiac rhythm generation. A stretch-inactivated channel (SIC) was recently cloned from rat kidney. The SIC channel belongs to a group of channels which are activated by pressure or stress on the cell membrane and conduct both Ca²⁺ and Na⁺ (Suzuki, M. et al. (1999) J. Biol. Chem. 274:6330-6335).

The pore-forming subunits of the voltage-gated cation channels form a superfamily of ion channel proteins. The characteristic domain of these channel proteins comprises six transmembrane domains (S1-S6), a pore-forming region (P) located between S5 and S6, and intracellular amino and carboxy termini. In the Na⁺ and Ca²⁺ subfamilies, this domain is repeated four times, while in the K⁺ channel subfamily, each channel is formed from a tetramer of either identical or dissimilar subunits. The P region contains information specifying the ion selectivity for the channel. In the case of K⁺ channels, a GYG tripeptide is involved in this selectivity (Ishii, T.M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11651-11656).

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Voltage-gated Na⁺ and K⁺ channels are necessary for the function of electrically excitable cells, such as nerve and muscle cells. Action potentials, which lead to neurotransmitter release and muscle contraction, arise from large, transient changes in the permeability of the membrane to Na⁺ and K⁺ ions. Depolarization of the membrane beyond the threshold level opens voltage-gated Na⁺ channels. Sodium ions flow into the cell, further depolarizing the membrane and opening more voltage-gated Na⁺ channels, which propagates the depolarization down the length of the cell. Depolarization also opens voltage-gated potassium channels. Consequently, potassium ions flow outward, which leads to repolarization of the membrane. Voltage-gated channels utilize charged residues in the fourth transmembrane segment (S4) to sense voltage change. The open state lasts only about 1 millisecond, at which time the channel spontaneously converts into an inactive state that cannot be opened irrespective of the membrane potential. Inactivation is mediated by the channel's N-terminus, which acts as a plug that closes the pore. The transition from an inactive to a closed state requires a return to resting potential.

Voltage-gated Na⁺ channels are heterotrimeric complexes composed of a 260 kDa poreforming α subunit that associates with two smaller auxiliary subunits, β 1 and β 2. The β 2 subunit is a integral membrane glycoprotein that contains an extracellular Ig domain, and its association with α

and β1 subunits correlates with increased functional expression of the channel, a change in its gating properties, as well as an increase in whole cell capacitance due to an increase in membrane surface area (Isom, L.L. et al. (1995) Cell 83:433-442).

Non voltage-gated Na⁺ channels include the members of the amiloride-sensitive Na⁺ channel/degenerin (NaC/DEG) family. Channel subunits of this family are thought to consist of two transmembrane domains flanking a long extracellular loop, with the amino and carboxyl termini located within the cell. The NaC/DEG family includes the epithelial Na⁺ channel (ENaC) involved in Na⁺ reabsorption in epithelia including the airway, distal colon, cortical collecting duct of the kidney, and exocrine duct glands. Mutations in ENaC result in pseudohypoaldosteronism type 1 and Liddle's syndrome (pseudohyperaldosteronism). The NaC/DEG family also includes the recently characterized H⁺-gated cation channels or acid-sensing ion channels (ASIC). ASIC subunits are expressed in the brain and form heteromultimeric Na⁺-permeable channels. These channels require acid pH fluctuations for activation. ASIC subunits show homology to the degenerins, a family of mechanically-gated channels originally isolated from C. elegans. Mutations in the degenerins cause neurodegeneration. ASIC subunits may also have a role in neuronal function, or in pain perception, since tissue acidosis causes pain (Waldmann, R. and M. Lazdunski (1998) Curr. Opin. Neurobiol. 8:418-424; Eglen, R.M. et al. (1999) Trends Pharmacol. Sci. 20:337-342).

K⁺ channels are located in all cell types, and may be regulated by voltage, ATP concentration, or second messengers such as Ca²⁺ and cAMP. In non-excitable tissue, K⁺ channels are involved in protein synthesis, control of endocrine secretions, and the maintenance of osmotic equilibrium across membranes. In neurons and other excitable cells, in addition to regulating action potentials and repolarizing membranes, K⁺ channels are responsible for setting resting membrane potential. The cytosol contains non-diffusible anions and, to balance this net negative charge, the cell contains a Na⁺-K⁺ pump and ion channels that provide the redistribution of Na⁺, K⁺, and Cl⁻. The pump actively transports Na⁺ out of the cell and K⁺ into the cell in a 3:2 ratio. Ion channels in the plasma membrane allow K⁺ and Cl⁻ to flow by passive diffusion. Because of the high negative charge within the cytosol, Cl⁻ flows out of the cell. The flow of K⁺ is balanced by an electromotive force pulling K⁺ into the cell, and a K⁺ concentration gradient pushing K⁺ out of the cell. Thus, the resting membrane potential is primarily regulated by K⁺ flow (Salkoff, L. and T. Jegla (1995) Neuron 15:489-492).

Potassium channel subunits of the <u>Shaker</u>-like superfamily all have the characteristic six transmembrane/1 pore domain structure. Four subunits combine as homo- or heterotetramers to form functional K channels. These pore-forming subunits also associate with various cytoplasmic β subunits that alter channel inactivation kinetics. The <u>Shaker</u>-like channel family includes the voltage-gated K⁺ channels as well as the delayed rectifier type channels such as the human ether-a-go-go

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related gene (HERG) associated with long QT, a cardiac dysrythmia syndrome (Curran, M.E. (1998) Curr. Opin. Biotechnol. 9:565-572; Kaczorowski, G.J. and M.L. Garcia (1999) Curr. Opin. Chem. Biol. 3:448-458).

A second superfamily of K⁺ channels is composed of the inward rectifying channels (Kir). Kir channels have the property of preferentially conducting K⁺ currents in the inward direction. These proteins consist of a single potassium selective pore domain and two transmembrane domains, which correspond to the fifth and sixth transmembrane domains of voltage-gated K⁺ channels. Kir subunits also associate as tetramers. The Kir family includes ROMK1, mutations in which lead to Bartter syndrome, a renal tubular disorder. Kir channels are also involved in regulation of cardiac pacemaker activity, seizures and epilepsy, and insulin regulation (Doupnik, C.A. et al. (1995) Curr. Opin. Neurobiol. 5:268-277; Curran, supra).

The recently recognized TWIK K⁺ channel family includes the mammalian TWIK-1, TREK-1 and TASK proteins. Members of this family possess an overall structure with four transmembrane domains and two P domains. These proteins are probably involved in controlling the resting potential in a large set of cell types (Duprat, F. et al. (1997) EMBO J 16:5464-5471).

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The voltage-gated Ca $^{2+}$ channels have been classified into several subtypes based upon their electrophysiological and pharmacological characteristics. L-type Ca $^{2+}$ channels are predominantly expressed in heart and skeletal muscle where they play an essential role in excitation-contraction coupling. T-type channels are important for cardiac pacemaker activity, while N-type and P/Q-type channels are involved in the control of neurotransmitter release in the central and peripheral nervous system. The L-type and N-type voltage-gated Ca $^{2+}$ channels have been purified and, though their functions differ dramatically, they have similar subunit compositions. The channels are composed of three subunits. The α_1 subunit forms the membrane pore and voltage sensor, while the $\alpha_2\delta$ and β subunits modulate the voltage-dependence, gating properties, and the current amplitude of the channel. These subunits are encoded by at least six α_1 , one $\alpha_2\delta$, and four β genes. A fourth subunit, γ , has been identified in skeletal muscle (Walker, D. et al. (1998) J. Biol. Chem. 273:2361-2367; McCleskey, E.W. (1994) Curr. Opin. Neurobiol. 4:304-312).

The transient receptor family (Trp) of calcium ion channels are thought to mediate capacitative calcium entry (CCE). CCE is the Ca²⁺ influx into cells to resupply Ca²⁺ stores depleted by the action of inositol triphosphate (IP3) and other agents in response to numerous hormones and growth factors. Trp and Trp-like were first cloned from Drosophila and have similarity to voltage gated Ca²⁺ channels in the S3 through S6 regions. This suggests that Trp and/or related proteins may form mammalian CCC entry channels (Zhu, X. et al. (1996) Cell 85:661-671; Boulay, G. et al. (1997) J. Biol. Chem. 272:29672-29680). Melastatin is a gene isolated in both the mouse and human, and whose expression in melanoma cells is inversely correlated with melanoma aggressiveness in vivo.

The human cDNA transcript corresponds to a 1533-amino acid protein having homology to members of the Trp family. It has been proposed that the combined use of malastatin mRNA expression status and tumor thickness might allow for the determination of subgroups of patients at both low and high risk for developing metastatic disease (Duncan, L.M. et al (2001) J. Clin. Oncol. 19:568-576).

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Chloride channels are necessary in endocrine secretion and in regulation of cytosolic and organelle pH. In secretory epithelial cells, Cl⁻ enters the cell across a basolateral membrane through an Na⁺, K⁺/Cl⁻ cotransporter, accumulating in the cell above its electrochemical equilibrium concentration. Secretion of Cl⁻ from the apical surface, in response to hormonal stimulation, leads to flow of Na⁺ and water into the secretory lumen. The cystic fibrosis transmembrane conductance regulator (CFTR) is a chloride channel encoded by the gene for cystic fibrosis, a common fatal genetic disorder in humans. CFTR is a member of the ABC transporter family, and is composed of two domains each consisting of six transmembrane domains followed by a nucleotide-binding site. Loss of CFTR function decreases transepithelial water secretion and, as a result, the layers of mucus that coat the respiratory tree, pancreatic ducts, and intestine are dehydrated and difficult to clear. The resulting blockage of these sites leads to pancreatic insufficiency, "meconium ileus", and devastating "chronic obstructive pulmonary disease" (Al-Awqati, Q. et al. (1992) J. Exp. Biol. 172:245-266).

The voltage-gated chloride channels (CLC) are characterized by 10-12 transmembrane domains, as well as two small globular domains known as CBS domains. The CLC subunits probably function as homotetramers. CLC proteins are involved in regulation of cell volume, membrane potential stabilization, signal transduction, and transepithelial transport. Mutations in CLC-1, expressed predominantly in skeletal muscle, are responsible for autosomal recessive generalized myotonia and autosomal dominant myotonia congenita, while mutations in the kidney channel CLC-5 lead to kidney stones (Jentsch, T.J. (1996) Curr. Opin. Neurobiol. 6:303-310).

Ligand-gated channels open their pores when an extracellular or intracellular mediator binds to the channel. Neurotransmitter-gated channels are channels that open when a neurotransmitter binds to their extracellular domain. These channels exist in the postsynaptic membrane of nerve or muscle cells. There are two types of neurotransmitter-gated channels. Sodium channels open in response to excitatory neurotransmitters, such as acetylcholine, glutamate, and serotonin. This opening causes an influx of Na⁺ and produces the initial localized depolarization that activates the voltage-gated channels and starts the action potential. Chloride channels open in response to inhibitory neurotransmitters, such as γ -aminobutyric acid (GABA) and glycine, leading to hyperpolarization of the membrane and the subsequent generation of an action potential. Neurotransmitter-gated ion channels have four transmembrane domains and probably function as pentamers (Jentsch, supra). Amino acids in the second transmembrane domain appear to be important in determining channel permeation and selectivity (Sather, W.A. et al. (1994) Curr. Opin. Neurobiol.

4:313-323).

Ligand-gated channels can be regulated by intracellular second messengers. For example, calcium-activated K^+ channels are gated by internal calcium ions. In nerve cells, an influx of calcium during depolarization opens K^+ channels to modulate the magnitude of the action potential (Ishi et al., supra). The large conductance (BK) channel has been purified from brain and its subunit composition determined. The α subunit of the BK channel has seven rather than six transmembrane domains in contrast to voltage-gated K^+ channels. The extra transmembrane domain is located at the subunit N-terminus. A 28-amino-acid stretch in the C-terminal region of the subunit (the "calcium bowl" region) contains many negatively charged residues and is thought to be the region responsible for calcium binding. The β subunit consists of two transmembrane domains connected by a glycosylated extracellular loop, with intracellular N- and C-termini (Kaczorowski, supra; Vergara, C. et al. (1998) Curr. Opin. Neurobiol. 8:321-329).

Cyclic nucleotide-gated (CNG) channels are gated by cytosolic cyclic nucleotides. The best examples of these are the cAMP-gated Na $^+$ channels involved in olfaction and the cGMP-gated cation channels involved in vision. Both systems involve ligand-mediated activation of a G-protein coupled receptor which then alters the level of cyclic nucleotide within the cell. CNG channels also represent a major pathway for Ca $^{2+}$ entry into neurons, and play roles in neuronal development and plasticity. CNG channels are tetramers containing at least two types of subunits, an α subunit which can form functional homomeric channels, and a β subunit, which modulates the channel properties. All CNG subunits have six transmembrane domains and a pore forming region between the fifth and sixth transmembrane domains, similar to voltage-gated K $^+$ channels. A large C-terminal domain contains a cyclic nucleotide binding domain, while the N-terminal domain confers variation among channel subtypes (Zufall, F. et al. (1997) Curr. Opin. Neurobiol. 7:404-412).

The activity of other types of ion channel proteins may also be modulated by a variety of intracellular signalling proteins. Many channels have sites for phosphorylation by one or more protein kinases including protein kinase A, protein kinase C, tyrosine kinase, and casein kinase II, all of which regulate ion channel activity in cells. Kir channels are activated by the binding of the Gβγ subunits of heterotrimeric G-proteins (Reimann, F. and F.M. Ashcroft (1999) Curr. Opin. Cell. Biol. 11:503-508). Other proteins are involved in the localization of ion channels to specific sites in the cell membrane. Such proteins include the PDZ domain proteins known as MAGUKs (membrane-associated guanylate kinases) which regulate the clustering of ion channels at neuronal synapses (Craven, S.B. and D.S. Bredt (1998) Cell 93:495-498).

Disease Correlation

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The etiology of numerous human diseases and disorders can be attributed to defects in the

transport of molecules across membranes. Defects in the trafficking of membrane-bound transporters and ion channels are associated with several disorders, e.g., cystic fibrosis, glucose-galactose malabsorption syndrome, hypercholesterolemia, von Gierke disease, and certain forms of diabetes mellitus. Single-gene defect diseases resulting in an inability to transport small molecules across membranes include, e.g., cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease (van't Hoff, W.G. (1996) Exp. Nephrol. 4:253-262; Talente, G.M. et al. (1994) Ann. Intern. Med. 120:218-226; and Chillon, M. et al. (1995) New Engl. J. Med. 332:1475-1480).

Human diseases caused by mutations in ion channel genes include disorders of skeletal muscle, cardiac muscle, and the central nervous system. Mutations in the pore-forming subunits of sodium and chloride channels cause myotonia, a muscle disorder in which relaxation after voluntary contraction is delayed. Sodium channel myotonias have been treated with channel blockers.

Mutations in muscle sodium and calcium channels cause forms of periodic paralysis, while mutations in the sarcoplasmic calcium release channel, T-tubule calcium channel, and muscle sodium channel cause malignant hyperthermia. Cardiac arrythmia disorders such as the long QT syndromes and idiopathic ventricular fibrillation are caused by mutations in potassium and sodium channels (Cooper, E.C. and L.Y. Jan (1998) Proc. Natl. Acad. Sci. USA 96:4759-4766). All four known human idiopathic epilepsy genes code for ion channel proteins (Berkovic, S.F. and I.E. Scheffer (1999) Curr. Opin. Neurology 12:177-182). Other neurological disorders such as ataxias, hemiplegic migraine and hereditary deafness can also result from mutations in ion channel genes (Jen, J. (1999) Curr. Opin. Neurobiol. 9:274-280; Cooper, supra).

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Ion channels have been the target for many drug therapies. Neurotransmitter-gated channels have been targeted in therapies for treatment of insomnia, anxiety, depression, and schizophrenia. Voltage-gated channels have been targeted in therapies for arrhythmia, ischemic stroke, head trauma, and neurodegenerative disease (Taylor, C.P. and L.S. Narasimhan (1997) Adv. Pharmacol. 39:47-98). Various classes of ion channels also play an important role in the perception of pain, and thus are potential targets for new analgesics. These include the vanilloid-gated ion channels, which are activated by the vanilloid capsaicin, as well as by noxious heat. Local anesthetics such as lidocaine and mexiletine which blockade voltage-gated Na⁺ channels have been useful in the treatment of neuropathic pain (Eglen, supra).

Ion channels in the immune system have recently been suggested as targets for immunomodulation. T-cell activation depends upon calcium signaling, and a diverse set of T-cell specific ion channels has been characterized that affect this signaling process. Channel blocking agents can inhibit secretion of lymphokines, cell proliferation, and killing of target cells. A peptide antagonist of the T-cell potassium channel Kv1.3 was found to suppress delayed-type hypersensitivity and allogenic responses in pigs, validating the idea of channel blockers as safe and efficacious

immunosuppressants (Cahalan, M.D. and K.G. Chandy (1997) Curr. Opin. Biotechnol. 8:749-756).

The discovery of new transporters and ion channels, and the polynucleotides encoding them, satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of transport, neurological, muscle, immunological, and cell proliferative disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

SUMMARY OF THE INVENTION

The invention features purified polypeptides, transporters and ion channels, referred to collectively as "TRICH" and individually as "TRICH-1," "TRICH-2," "TRICH-3," "TRICH-4," "TRICH-5," "TRICH-6," "TRICH-7," "TRICH-8," "TRICH-9," "TRICH-10," "TRICH-11," "TRICH-11," "TRICH-12," "TRICH-13," "TRICH-14," "TRICH-15," "TRICH-16," "TRICH-17," "TRICH-18," "TRICH-19," "TRICH-20," "TRICH-21," "TRICH-22," "TRICH-23," "TRICH-24," "TRICH-25," "TRICH-26," "TRICH-27," "TRICH-28," "TRICH-29," and "TRICH-30." In one aspect, the invention provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. In one alternative, the invention provides an isolated polypeptide comprising the amino acid sequence of SEQ ID NO:1-30.

The invention further provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-30. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:31-60.

Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting

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of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide. In another alternative, the invention provides a transgenic organism comprising the recombinant polynucleotide.

The invention also provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30.

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The invention further provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group

consisting of SEQ ID NO:31-60, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous nucleotides.

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The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

The invention further provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and a pharmaceutically acceptable excipient. In one embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

The invention also provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino

acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

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The invention further provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, and b) detecting altered expression of the target polynucleotide.

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The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target

polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the present invention.

Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog for polypeptides of the invention. The probability score for the match between each polypeptide and its GenBank homolog is also shown.

Table 3 shows structural features of polypeptide sequences of the invention, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide sequences of the invention, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for polynucleotides of the invention.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

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DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so

forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

10 **DEFINITIONS**

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"TRICH" refers to the amino acid sequences of substantially purified TRICH obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of TRICH. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

An "allelic variant" is an alternative form of the gene encoding TRICH. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding TRICH include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as TRICH or a polypeptide with at least one functional characteristic of TRICH. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding TRICH, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding TRICH. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent TRICH. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of TRICH is retained. For example,

negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

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The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of TRICH. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind TRICH polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA;

RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic TRICH, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that annual by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

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A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding TRICH or fragments of TRICH may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows

amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
	Ala	Gly, Ser
5	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys ·	Ala, Ser
	Gln	Asn, Glu, His
10	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
	Leu	Ile, Val
15	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
	Thr	Ser, Val
20	Тгр	Phe, Tyr
	Tyr	His, Phe, Trp
	Val	Ile, Leu, Thr

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

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A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide.

Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

"Exon shuffling" refers to the recombination of different coding regions (exons). Since an exon may represent a structural or functional domain of the encoded protein, new proteins may be assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

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A "fragment" is a unique portion of TRICH or the polynucleotide encoding TRICH which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:31-60 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:31-60, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:31-60 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:31-60 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:31-60 and the region of SEQ ID NO:31-60 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-30 is encoded by a fragment of SEQ ID NO:31-60. A fragment of SEQ ID NO:1-30 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-30. For example, a fragment of SEQ ID NO:1-30 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-30. The precise length of a fragment of SEQ ID NO:1-30 and the region of SEQ ID NO:1-30 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Reward for match: 1

30 Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 11

35 Filter: on

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Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10
Word Size: 3

Filter: on

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Percent identity may be measured over the length of an entire defined polypeptide sequence,

for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

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"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 µg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_m and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC

concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 μ g/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., C_0 t or R_0 t analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

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The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of TRICH which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of TRICH which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of TRICH. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of TRICH.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the

antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an TRICH may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of TRICH.

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"Probe" refers to nucleic acid sequences encoding TRICH, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that

purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, <u>supra</u>. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

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Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is

expressed, inducing a protective immunological response in the mammal.

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A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing TRICH, nucleic acids encoding TRICH, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free, preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" refers to the collective pattern of gene expression by a particular cell

type or tissue under given conditions at a given time.

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"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to

another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length of one of the polypeptides.

15 THE INVENTION

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The invention is based on the discovery of new human transporters and ion channels (TRICH), the polynucleotides encoding TRICH, and the use of these compositions for the diagnosis, treatment, or prevention of transport, neurological, muscle, immunological, and cell proliferative disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (Genbank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability score for the match between each polypeptide and its GenBank homolog. Column 5 shows the annotation of the GenBank homolog along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1

and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

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Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are transporters and ion channels. For example, SEQ ID NO:6 is 89% identical to rat neuronal nicotinic acetylcholine receptor subunit (GenBank ID g6746563) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.7e-188, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:6 also contains a neurotransmitter-gated ion channel domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:6 is a neurotransmitter-gated ion channel. In an alternative example, SEQ ID NO:14 is 93% identical to rat TAP-like ABC transporter (GenBank ID g6045150) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:14 also contains an ABC transporter domain and an ABC transporter transmembrane region as determined by searching for statistically significant matches in the hidden Markov model (HMM)based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:14 is an ABC transporter. In an alternative example, SEQ ID NO:16 is 98% identical to human voltagedependent anion channel (GenBank ID g340199) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.2e-130, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:16 also contains a eukaryotic porin active site domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:16 is a mitochondrial porin. In an alternative example, SEQ ID NO:20 is 28% identical to a rat voltage-gated calcium channel (GenBank ID g4586963) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The

BLAST probability score is 2.4e-27, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. Data from BLIMPS and BLAST analyses provide further corroborative evidence that SEQ ID NO:20 is a voltage-gated calcium channel. In an alternative example, SEQ ID NO:22 is 82% identical to human inhibitory glycine receptor (GenBank ID g31849) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.1e-175, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:22 also contains a neurotransmitter-gated ion channel domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:22 is a neurotransmitter-gated ion channel. In an alternative example, SEQ ID NO:30 is 36% identical to human ATP binding cassette (ABC) -C transporter (GenBank ID g1514530) as determined by the Basic Local Alignment Search Tool (BLAST, see Table 2). The BLAST probability score is 2.3e-127, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:30 also contains ABC transporter domains as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains (see Table 3). Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:30 is an ABC transporter. SEQ ID NO:1-5, SEQ ID NO:7-13, SEQ ID NO:15, SEQ ID NO:17-19, SEQ ID NO:21, and SEQ ID NO:23-29 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-30 are described in Table 7.

As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Columns 1 and 2 list the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and the corresponding Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) for each polynucleotide of the invention. Column 3 shows the length of each polynucleotide sequence in basepairs. Column 4 lists fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:31-60 or that distinguish between SEQ ID NO:31-60 and related polynucleotide sequences. Column 5 shows identification numbers corresponding to cDNA sequences, coding sequences (exons) predicted from genomic DNA, and/or sequence assemblages comprised of both cDNA and genomic DNA. These sequences were used to assemble the full length polynucleotide sequences of the invention. Columns 6 and 7 of Table 4 show the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences in column 5 relative to their respective full length sequences.

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The identification numbers in Column 5 of Table 4 may refer specifically, for example, to Incyte cDNAs along with their corresponding cDNA libraries. For example, 6340750H1 is the identification number of an Incyte cDNA sequence, and BRANDIN01 is the cDNA library from which it is derived. Incyte cDNAs for which cDNA libraries are not indicated were derived from pooled cDNA libraries (e.g., 71911330V1). Alternatively, the identification numbers in column 5 may refer to GenBank cDNAs or ESTs (e.g., g5110579) which contributed to the assembly of the full length polynucleotide sequences. In addition, the identification numbers in column 5 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the identification numbers in column 5 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the identification numbers in column 5 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, FL_XXXXXX_N₁_N₂_YYYYY_N₃_N₄ represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYYY is the number of the prediction generated by the algorithm, and $N_{123...}$, if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the identification numbers in column 5 may refer to assemblages of exons brought together by an "exon-stretching" algorithm. For example,

FLXXXXXX_gAAAAA_gBBBBB_1_N is the identification number of a "stretched" sequence, with XXXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," "NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

Prefix	Type of analysis and/or examples of programs
LITCHA	1 Type of analysis and/of examples of programs

GNN, GFG,	Exon prediction from genomic sequences using, for example,
ENST	GENSCAN (Stanford University, CA, USA) or FGENES
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).
GBI	Hand-edited analysis of genomic sequences.
FL	Stitched or stretched genomic sequences (see Example V).
INCY	Full length transcript and exon prediction from mapping of EST sequences to the genome. Genomic location and EST composition
	data are combined to predict the exons and resulting transcript.

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in column 5 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

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Table 5 shows the representative cDNA libraries for those full length polynucleotide sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

The invention also encompasses TRICH variants. A preferred TRICH variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the TRICH amino acid sequence, and which contains at least one functional or structural characteristic of TRICH.

The invention also encompasses polynucleotides which encode TRICH. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:31-60, which encodes TRICH. The polynucleotide sequences of SEQ ID NO:31-60, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding TRICH. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding TRICH. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:31-60 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting

of SEQ ID NO:31-60. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of TRICH.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding TRICH, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring TRICH, and all such variations are to be considered as being specifically disclosed.

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Although nucleotide sequences which encode TRICH and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring TRICH under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding TRICH or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding TRICH and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode TRICH and TRICH derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding TRICH or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:31-60 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or

combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

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The nucleic acid sequences encoding TRICH may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence

into 5'non-transcribed regulatory regions.

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Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode TRICH may be cloned in recombinant DNA molecules that direct expression of TRICH, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express TRICH.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter TRICH-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent Number 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of TRICH, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of

homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

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In another embodiment, sequences encoding TRICH may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.)

Alternatively, TRICH itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of TRICH, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

In order to express a biologically active TRICH, the nucleotide sequences encoding TRICH or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding TRICH. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding TRICH. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding TRICH and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression

vectors containing sequences encoding TRICH and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. (See, e.g., Sambrook, J. et al. (1989) Molecular Cloning, A

Laboratory Manual, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) Current Protocols in Molecular Biology, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding TRICH. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; and Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.) Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5(6):350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90(13):6340-6344; Buller, R.M. et al. (1985) Nature 317(6040):813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31(3):219-226; and Verma, I.M. and N. Somia (1997) Nature 389:239-242.) The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding TRICH. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding TRICH can be achieved using a multifunctional <u>E. coli</u> vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding TRICH into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of TRICH are needed, e.g. for the production of antibodies, vectors which direct high level expression of TRICH may be used. For example, vectors

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containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

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Yeast expression systems may be used for production of TRICH. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast <u>Saccharomyces cerevisiae</u> or <u>Pichia pastoris</u>. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, <u>supra</u>; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of TRICH. Transcription of sequences encoding TRICH may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding TRICH may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses TRICH in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of TRICH in cell lines is preferred. For example, sequences encoding TRICH can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in

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enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

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Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in tk and apr cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, dhfr confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., trpB and hisD, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. 15 Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), & glucuronidase and its substrate B-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding TRICH is inserted within a marker gene sequence, transformed cells containing sequences encoding TRICH can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding TRICH under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding TRICH and that express TRICH may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of TRICH using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing

monoclonal antibodies reactive to two non-interfering epitopes on TRICH is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) <u>Serological Methods</u>, a <u>Laboratory Manual</u>, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) <u>Current Protocols in Immunology</u>, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) <u>Immunochemical Protocols</u>, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding TRICH include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding TRICH, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding TRICH may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode TRICH may be designed to contain signal sequences which direct secretion of TRICH through a prokaryotic or eukaryotic cell membrane.

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In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding TRICH may be ligated to a heterologous sequence resulting in translation of a

fusion protein in any of the aforementioned host systems. For example, a chimeric TRICH protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of TRICH activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the TRICH encoding sequence and the heterologous protein sequence, so that TRICH may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, supra, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled TRICH may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

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TRICH of the present invention or fragments thereof may be used to screen for compounds that specifically bind to TRICH. At least one and up to a plurality of test compounds may be screened for specific binding to TRICH. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of TRICH, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which TRICH binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case, the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express TRICH, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, <u>Drosophila</u>, or <u>E. coli</u>. Cells expressing TRICH or cell membrane fractions which contain TRICH are then contacted with a test compound and binding, stimulation, or inhibition of activity of either TRICH or the

compound is analyzed.

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An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with TRICH, either in solution or affixed to a solid support, and detecting the binding of TRICH to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

TRICH of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of TRICH. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for TRICH activity, wherein TRICH is combined with at least one test compound, and the activity of TRICH in the presence of a test compound is compared with the activity of TRICH in the absence of the test compound. A change in the activity of TRICH in the presence of the test compound is indicative of a compound that modulates the activity of TRICH. Alternatively, a test compound is combined with an in vitro or cell-free system comprising TRICH under conditions suitable for TRICH activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of TRICH may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding TRICH or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent Number 5,175,383 and U.S. Patent Number 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential

therapeutic or toxic agents.

Polynucleotides encoding TRICH may also be manipulated <u>in vitro</u> in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding TRICH can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding TRICH is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress TRICH, e.g., by secreting TRICH in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

THERAPEUTICS

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of TRICH and transporters and ion channels. In addition, the expression of TRICH is closely associated with brain, liver, tumor, colon, thymus, small intestine, myometrium, testicular, bone marrow neuroblastoma tumor, parotid gland, lung, pituitary gland, and placental tissues, and Pompe's disease. Therefore, TRICH appears to play a role in transport, neurological, muscle, immunological, and cell proliferative disorders. In the treatment of disorders associated with increased TRICH expression or activity, it is desirable to decrease the expression or activity, it is desirable to increase the expression or activity, it is desirable to increase the expression or activity of TRICH.

Therefore, in one embodiment, TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy,

ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic 20 nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, 25 postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, and acid maltase deficiency (AMD, also known

as Pompe's disease); an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus. 20

In another embodiment, a vector capable of expressing TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified TRICH in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of TRICH may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those listed above.

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In a further embodiment, an antagonist of TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH. Examples of such disorders include, but are not limited to, those transport, neurological, muscle, immunological, and cell proliferative disorders described above. In one aspect, an antibody which specifically binds

TRICH may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express TRICH.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of TRICH may be produced using methods which are generally known in the art. In particular, purified TRICH may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind TRICH. Antibodies to TRICH may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with TRICH or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

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It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to TRICH have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of TRICH amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to TRICH may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma

technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce TRICH-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing <u>in vivo</u> production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for TRICH may also be generated. For example, such fragments include, but are not limited to, F(ab')₂ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between TRICH and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering TRICH epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for TRICH. Affinity is expressed as an association constant, K_a , which is defined as the molar concentration of TRICH-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple TRICH epitopes, represents the average affinity, or avidity, of the antibodies

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for TRICH. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular TRICH epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10° to 10¹² L/mole are preferred for use in immunoassays in which the TRICH-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10⁶ to 10⁷ L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of TRICH, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of TRICH-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

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In another embodiment of the invention, the polynucleotides encoding TRICH, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding TRICH. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding TRICH. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) Blood 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res.

25(14):2730-2736.)

In another embodiment of the invention, polynucleotides encoding TRICH may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by Xlinked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA. 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in TRICH expression or regulation causes disease, the expression of TRICH from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency. 20

In a further embodiment of the invention, diseases or disorders caused by deficiencies in TRICH are treated by constructing mammalian expression vectors encoding TRICH and introducing these vectors by mechanical means into TRICH-deficient cells. Mechanical transfer technologies for use with cells <u>in vivo</u> or <u>ex vitro</u> include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

Expression vectors that may be effective for the expression of TRICH include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). TRICH may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl.

Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and Blau, H.M. supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding TRICH from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

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In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to TRICH expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding TRICH under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent Number 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4+ Tcells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In the alternative, an adenovirus-based gene therapy delivery system is used to deliver

polynucleotides encoding TRICH to cells which have one or more genetic abnormalities with respect to the expression of TRICH. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent Number 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.

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In another alternative, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding TRICH to target cells which have one or more genetic abnormalities with respect to the expression of TRICH. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing TRICH to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent Number 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent Number 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding TRICH to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity

(e.g., protease and polymerase). Similarly, inserting the coding sequence for TRICH into the alphavirus genome in place of the capsid-coding region results in the production of a large number of TRICH-coding RNAs and the synthesis of high levels of TRICH in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of TRICH into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding TRICH.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques

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for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by <u>in vitro</u> and <u>in vivo</u> transcription of DNA sequences encoding TRICH. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothicate or 2'O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

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An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding TRICH. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased TRICH expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding TRICH may be therapeutically useful, and in the treatment of disorders associated with decreased TRICH expression or activity, a compound which specifically promotes expression of the polynucleotide encoding TRICH may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding TRICH is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an <u>in vitro</u> cell-free or reconstituted

biochemical system. Alterations in the expression of a polynucleotide encoding TRICH are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding TRICH. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat.

Biotechnol. 15:462-466.)

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Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of TRICH, antibodies to TRICH, and mimetics, agonists, antagonists, or inhibitors of TRICH.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

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Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising TRICH or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, TRICH or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example TRICH or fragments thereof, antibodies of TRICH, and agonists, antagonists or inhibitors of TRICH, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED₅₀ (the dose therapeutically effective in 50% of the population) or LD₅₀ (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD₅₀/ED₅₀ ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about $0.1~\mu g$ to $100,000~\mu g$, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

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In another embodiment, antibodies which specifically bind TRICH may be used for the diagnosis of disorders characterized by expression of TRICH, or in assays to monitor patients being treated with TRICH or agonists, antagonists, or inhibitors of TRICH. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for TRICH include methods which utilize the antibody and a label to detect TRICH in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring TRICH, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of TRICH expression. Normal or standard values for TRICH expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to TRICH under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of TRICH expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding TRICH may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of TRICH may be correlated

with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of TRICH, and to monitor regulation of TRICH levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding TRICH or closely related molecules may be used to identify nucleic acid sequences which encode TRICH. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding TRICH, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the TRICH encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:31-60 or from genomic sequences including promoters, enhancers, and introns of the TRICH gene.

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Means for producing specific hybridization probes for DNAs encoding TRICH include the cloning of polynucleotide sequences encoding TRICH or TRICH derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avaidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding TRICH may be used for the diagnosis of disorders associated with expression of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease,

cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, and acid maltase deficiency (AMD, also known as Pompe's disease); an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact

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dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus. The polynucleotide sequences encoding TRICH may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered TRICH expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding TRICH may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding TRICH may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding TRICH in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

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In order to provide a basis for the diagnosis of a disorder associated with expression of TRICH, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding TRICH, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from

normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

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Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under-or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding TRICH may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced <u>in vitro</u>. Oligomers will preferably contain a fragment of a polynucleotide encoding TRICH, or a fragment of a polynucleotide complementary to the polynucleotide encoding TRICH, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the

sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of TRICH include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

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In another embodiment, TRICH, fragments of TRICH, or antibodies specific for TRICH may be used as elements on a microarray. The microarray may be used to monitor or measure proteinprotein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent Number 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the

hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression <u>in vivo</u>, as in the case of a tissue or biopsy sample, or <u>in vitro</u>, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

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In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global

pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for TRICH to quantify the levels of TRICH expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lucking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

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Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

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Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in <u>DNA Microarrays: A Practical Approach</u>, M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding TRICH may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism

(RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding TRICH on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, TRICH, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between TRICH and the agent being tested may be measured.

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Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with TRICH, or fragments thereof, and washed. Bound TRICH is then detected by methods well known in the art. Purified TRICH can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding TRICH specifically compete with a test compound for binding TRICH. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more

antigenic determinants with TRICH.

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In additional embodiments, the nucleotide sequences which encode TRICH may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, including U.S. Ser. No. 60/223,269, U.S. Ser. No. 60/224,456, U.S. Ser. No. 60/226,410, U.S. Ser. No. 60/228,140, U.S. Ser. No. 60/230,067, and U.S. Ser. No. 60/231,434, are hereby expressly incorporated by reference.

EXAMPLES

I. Construction of cDNA Libraries

Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA) and shown in Table 4, column 5. Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the

appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), or pINCY (Incyte Genomics, Palo Alto CA), or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5α, DH10B, or ElectroMAX DH10B from Life Technologies.

10 II. Isolation of cDNA Clones

Plasmids obtained as described in Example I were recovered from host cells by <u>in vivo</u> excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

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Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading

frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, <u>supra</u>, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

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The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM, and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families. See, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide of the invention may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value,

the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:31-60. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 4.

IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative transporters and ion channels were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode transporters and ion channels, the encoded polypeptides were analyzed by querying against PFAM models for transporters and ion channels. Potential transporters and ion channels were also identified by homology to Incyte cDNA sequences that had been annotated as transporters and ion channels. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

V. Assembly of Genomic Sequence Data with cDNA Sequence Data

30 "Stitched" Sequences

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Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information,

generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

"Stretched" Sequences

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Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

VI. Chromosomal Mapping of TRICH Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:31-60 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:31-60 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences

had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's parm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

VII. Analysis of Polynucleotide Expression

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Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, <u>supra</u>, ch. 7; Ausubel (1995) <u>supra</u>, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the

entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotide sequences encoding TRICH are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding TRICH. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

VIII. Extension of TRICH Encoding Polynucleotides

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Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68 °C to about 72 °C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg²⁺, (NH₄)₂SO₄, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer

pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

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The concentration of DNA in each well was determined by dispensing 100 μ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1% agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotide sequences are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

IX. Labeling and Use of Individual Hybridization Probes

Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

Hybridization probes derived from SEQ ID NO:31-60 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μCi of [γ-³²P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech).

10 An aliquot containing 10⁷ counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases:

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

X. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, supra.), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena (1999), supra). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers.

25 Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645;

Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection.

After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

Tissue or Cell Sample Preparation

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)⁺ RNA is purified using the oligo-(dT) cellulose method. Each poly(A)⁺ RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/μl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)⁺ RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)⁺ RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories, Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

Microarray Preparation

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Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μ g. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a

110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522, incorporated herein by reference. 1 μ l of the array element DNA, at an average concentration of 100 ng/ μ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

Hybridization

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Hybridization reactions contain 9 μ l of sample mixture consisting of 0.2 μ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 μ l of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source,

although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

XI. Complementary Polynucleotides

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Sequences complementary to the TRICH-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring TRICH. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of TRICH. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the TRICH-encoding transcript.

XII. Expression of TRICH

Expression and purification of TRICH is achieved using bacterial or virus-based expression systems. For expression of TRICH in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the *trp-lac* (*tac*) hybrid

promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express TRICH upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of TRICH in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding TRICH by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, TRICH is synthesized as a fusion protein with, e.g., glutathione S-transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from TRICH at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, supra, ch. 10 and 16). Purified TRICH obtained by these methods can be used directly in the assays shown in Examples XVI, XVII, and XVIII, where applicable.

XIII. Functional Assays

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TRICH function is assessed by expressing the sequences encoding TRICH at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT (Life Technologies) and PCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10 μ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the

recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of TRICH on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding TRICH and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding TRICH and other genes of interest can be analyzed by northern analysis or microarray techniques.

XIV. Production of TRICH Specific Antibodies

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TRICH substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the TRICH amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, supra.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-TRICH activity by, for example, binding the peptide or TRICH to a substrate,

blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XV. Purification of Naturally Occurring TRICH Using Specific Antibodies

Naturally occurring or recombinant TRICH is substantially purified by immunoaffinity chromatography using antibodies specific for TRICH. An immunoaffinity column is constructed by covalently coupling anti-TRICH antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing TRICH are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of TRICH (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/TRICH binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and TRICH is collected.

XVI. Identification of Molecules Which Interact with TRICH

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Molecules which interact with TRICH may include transporter substrates, agonists or antagonists, modulatory proteins such as Gβγ proteins (Reimann, supra) or proteins involved in TRICH localization or clustering such as MAGUKs (Craven, supra). TRICH, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled TRICH, washed, and any wells with labeled TRICH complex are assayed. Data obtained using different concentrations of TRICH are used to calculate values for the number, affinity, and association of TRICH with the candidate molecules.

Alternatively, proteins that interact with TRICH are isolated using the yeast 2-hybrid system (Fields, S. and O. Song (1989) Nature 340:245-246). TRICH, or fragments thereof, are expressed as fusion proteins with the DNA binding domain of Gal4 or lexA, and potential interacting proteins are expressed as fusion proteins with an activation domain. Interactions between the TRICH fusion protein and the TRICH interacting proteins (fusion proteins with an activation domain) reconstitute a transactivation function that is observed by expression of a reporter gene. Yeast 2-hybrid systems are commercially available, and methods for use of the yeast 2-hybrid system with ion channel proteins are discussed in Niethammer, M. and M. Sheng (1998, Meth. Enzymol. 293:104-122).

TRICH may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

Potential TRICH agonists or antagonists may be tested for activation or inhibition of TRICH

ion channel activity using the assays described in section XVIII.

XVII. Demonstration of TRICH Activity

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Ion channel activity of TRICH is demonstrated using an electrophysiological assay for ion conductance. TRICH can be expressed by transforming a mammalian cell line such as COS7, HeLa or CHO with a eukaryotic expression vector encoding TRICH. Eukaryotic expression vectors are commercially available, and the techniques to introduce them into cells are well known to those skilled in the art. A second plasmid which expresses any one of a number of marker genes, such as &galactosidase, is co-transformed into the cells to allow rapid identification of those cells which have taken up and expressed the foreign DNA. The cells are incubated for 48-72 hours after 10 transformation under conditions appropriate for the cell line to allow expression and accumulation of TRICH and B-galactosidase.

Transformed cells expressing B-galactosidase are stained blue when a suitable colorimetric substrate is added to the culture media under conditions that are well known in the art. Stained cells are tested for differences in membrane conductance by electrophysiological techniques that are well 15 known in the art. Untransformed cells, and/or cells transformed with either vector sequences alone or B-galactosidase sequences alone, are used as controls and tested in parallel. Cells expressing TRICH will have higher anion or cation conductance relative to control cells. The contribution of TRICH to conductance can be confirmed by incubating the cells using antibodies specific for TRICH. The antibodies will bind to the extracellular side of TRICH, thereby blocking the pore in the ion channel, and the associated conductance.

Alternatively, ion channel activity of TRICH is measured as current flow across a TRICHcontaining Xenopus laevis oocyte membrane using the two-electrode voltage-clamp technique (Ishi et al., supra; Jegla, T. and L. Salkoff (1997) J. Neurosci. 17:32-44). TRICH is subcloned into an appropriate Xenopus oocyte expression vector, such as pBF, and 0.5-5 ng of mRNA is injected into mature stage IV oocytes. Injected oocytes are incubated at 18 °C for 1-5 days. Inside-out macropatches are excised into an intracellular solution containing 116 mM K-gluconate, 4 mM KCl, and 10 mM Hepes (pH 7.2). The intracellular solution is supplemented with varying concentrations of the TRICH mediator, such as cAMP, cGMP, or Ca⁺² (in the form of CaCl₂), where appropriate. Electrode resistance is set at 2-5 M Ω and electrodes are filled with the intracellular solution lacking mediator. Experiments are performed at room temperature from a holding potential of 0 mV. Voltage ramps (2.5 s) from -100 to 100 mV are acquired at a sampling frequency of 500 Hz. Current measured is proportional to the activity of TRICH in the assay.

In particular, the activity of TRICH-20 is measured as Ca²⁺ conductance, the activity of TRICH-22 is measured as Cl- conductance in the presence of glycine, the activity of TRICH-23 is

measured as Ca²⁺ conductance, and the activity of TRICH-24 is measured as K⁺ conductance in the presence of Ca²⁺, and the activity of TRICH-26 is measured as cation conductance in the presence of heat.

Transport activity of TRICH is assayed by measuring uptake of labeled substrates substrates (including but not limited to, maltose, glucose, or glycogen) into Xenopus laevis oocytes. Oocytes at stages V and VI are injected with TRICH mRNA (10 ng per oocyte) and incubated for 3 days at 18°C in OR2 medium (82.5mM NaCl, 2.5 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 1mM Na₂HPO₄, 5 mM Hepes, 3.8 mM NaOH, 50µg/ml gentamycin, pH 7.8) to allow expression of TRICH. Oocytes are then transferred to standard uptake medium (100mM NaCl, 2 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 10 mM Hepes/Tris pH 7.5). Uptake of various substrates (e.g., amino acids, sugars, drugs, ions, and neurotransmitters) is initiated by adding labeled substrate (e.g. radiolabeled with ³H, fluorescently labeled with rhodamine, etc.) to the oocytes. After incubating for 30 minutes, uptake is terminated by washing the oocytes three times in Na⁺-free medium, measuring the incorporated label, and comparing with controls. TRICH activity is proportional to the level of internalized labeled substrate. In particular, test substrates include sulfate for TRICH-13, tricarboxylates for TRICH-21, dicarboxylates and Na⁺ for TRICH-25, ornithine for TRICH-27, and monocarboxylates for TRICH-28.

ATPase activity associated with TRICH can be measured by hydrolysis of radiolabeled ATP- $[\gamma^{-32}P]$, separation of the hydrolysis products by chromatographic methods, and quantitation of the recovered ³²P using a scintillation counter. The reaction mixture contains ATP- $[\gamma^{-32}P]$ and varying amounts of TRICH in a suitable buffer incubated at 37°C for a suitable period of time. The reaction is terminated by acid precipitation with trichloroacetic acid and then neutralized with base, and an aliquot of the reaction mixture is subjected to membrane or filter paper-based chromatography to separate the reaction products. The amount of ³²P liberated is counted in a scintillation counter. The amount of radioactivity recovered is proportional to the ATPase activity of TRICH in the assay.

XVIII. Identification of TRICH Agonists and Antagonists

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TRICH is expressed in a eukaryotic cell line such as CHO (Chinese Hamster Ovary) or HEK (Human Embryonic Kidney) 293. Ion channel activity of the transformed cells is measured in the presence and absence of candidate agonists or antagonists. Ion channel activity is assayed using patch clamp methods well known in the art or as described in Example XVII. Alternatively, ion channel activity is assayed using fluorescent techniques that measure ion flux across the cell membrane (Velicelebi, G. et al. (1999) Meth. Enzymol. 294:20-47; West, M.R. and C.R. Molloy (1996) Anal. Biochem. 241:51-58). These assays may be adapted for high-throughput screening using microplates. Changes in internal ion concentration are measured using fluorescent dyes such as the Ca²⁺ indicator Fluo-4 AM, sodium-sensitive dyes such as SBFI and sodium green, or the Cl

indicator MQAE (all available from Molecular Probes) in combination with the FLIPR fluorimetric plate reading system (Molecular Devices). In a more generic version of this assay, changes in membrane potential caused by ionic flux across the plasma membrane are measured using oxonyl dyes such as DiBAC₄ (Molecular Probes). DiBAC₄ equilibrates between the extracellular solution and cellular sites according to the cellular membrane potential. The dye's fluorescence intensity is 20-fold greater when bound to hydrophobic intracellular sites, allowing detection of DiBAC₄ entry into the cell (Gonzalez, J.E. and P.A. Negulescu (1998) Curr. Opin. Biotechnol. 9:624-631). Candidate agonists or antagonists may be selected from known ion channel agonists or antagonists, peptide libraries, or combinatorial chemical libraries.

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Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

Table 1

Incyte	Polvpeptide	Incyte	Polynucleotide	Incvte
Project ID	SEQ ID NO:	Polypeptide ID	SEQ_ID NO:	Polynucleotide ID
2194064	-1	2194064CD1	31	2194064CB1
2744094	2	2744094CD1	32	2744094CB1
2798241	3	2798241CD1	33	2798241CB1
3105257	4	3105257CD1	34	3105257CB1
3200979	2	3200979CD1	35	3200979CB1
6754139	9	6754139CD1	. 36	6754139CB1
639669	7	6996659CD1	37	6996659CB1
7472747	œ	7472747CD1	38	7472747CB1
7474121	6	7474121CD1	39	7474121CB1
7475615	10	7475615CD1	40	7475615CB1
7475656	11	7475656CD1	41	747565CB1
7480632	12	7480632CD1	42	7480632CB1
6952742	13	6952742CD1	43	6952742CB1
7478795	14	7478795CD1	77	7478795CB1
656293	15	656293CD1	45	656293CB1
7473957	16	7473957CD1	46	7473957CB1
7474111	1.7	7474111CD1	47	7474111CB1
7480826	18	7480826CD1	87	7480826CB1
6025572	19	6025572CD1	67	6025572CB1
5686561	20	5686561CD1	20	5686561CB1
1553725	21	1553725CD1	51	1553725¢B1
1695770	22	1695770CD1	52	1695770CB1
4672222	23	4672222CD1	53	4672222CB1
6176128	24	6176128CD1	54	6176128CB1
7473418	25	7473418CD1	55	7473418CB1
7474129	26	7474129CD1	26	7474129CB1
7481414	27	7481414CD1	57	7481414CB1
7481461	28	7481461CD1	58	7481461CB1
7472541	29	7472541CD1	59	7472541CB1
6999183	30	6999183CD1	09	6999183CB1

Table 2

Polypeptide SEQ ID NO:	Incyte Polypeptide	GenBank ID NO:	Probability score	GenBank Homolog
	2194064CD1	02463634	1.608-41	Monocarboxvlate transporter [Homo sapiens]
ı				
2	2744094CD1	g13346481	0	ATP-binding cassette transporter MRP8 [Homo sapiens]
3	2798241CD1	g1699038	2.90E-142	ABC3 [Homo sapiens] (Connors. T. D. et al. (1997) Genomics 39:231-234)
4	3105257CD1	q8650412	0	
)		(Zhang, F. et al. (2000) Characterization of ABCB9, an
				ATP binding cassette protein associated with lysosomes
				J. Biol. Chem. 275:23287-23294)
S	3200979CD1	g1514530	3.10压-119	ABC-C transporter [Homo sapiens]
				(Klugbauer, N. and F. Hofmann (1996)
				FEBS Lett. 391:61-65)
9	6754139CD1	g6746563	1.70E-188	neuronal nicotinic acetylcholine receptor subunit
				[Rattus norvegicus]
				(Elgoyhen, A. B. et al. (2001) alpha 10: A determinant
٠.				of nicotinic cholinergic receptor function in mammalian
				vestibular and cochlear mechanosensory hair cells
				Proc. Natl. Acad. Sci. U.S.A. 98:3501-3506)
7	6996659CD1	g1050330	0	Ionotropic glutamate receptor [Rattus norvegicus]
				(Ciabarra, A.M. et al. (1995) J. Neurosci. 15:6498-
				6508)
8	7472747CD1	g13926108	1.00E-157	2P domain potassium channel Talk-1 [Homo sapiens]
				(Girard, C. et al. (2001) Genomic and functional
				characteristics of novel human pancreatic 2P domain
				K(+) channels. Biochem Biophys Res Commun. 282:249-256)
6	7474121CD1	g2465542	7.00E-20	TWIK-related acid-sensitive K+ channel [Homo sapiens] (Duprat, F. et al. (1997) EMBO J. 16:5464-5471)
10	7475615CD1	g2654005	5.70E-114	Pendrin [Homo sapiens]
				(EVERECT, L.A. et al. (1997) Nature Genet. 1/:411-422)

Table 2 (cont.)

Polypeptide	Incvte	GenBank ID	Probability	GenBank Homolog
SEQ ID NO:	Polypeptide ID	NO:	score	
11	7475656CD1	g3168874	0	<pre>Ion channel BCNG-1 [Homo sapiens] (Santoro, B. et al. (1997) Proc. Natl. Acad. Sci. USA 94:14815-14820)</pre>
12	7480632CD1	g1514530	9.80E-123	ABC-C transporter [Homo sapiens] (Klugbauer, M. and F. Hofmann (1996) FEBS Lett. 391:61- 65)
133	6952742CD1	g10719650	0	sulfate/anion transporter SAT-1 protein [Homo sapiens] (Lohi, H. et al. (2000) Mapping of Five New Putative Anion Transporter Genes in Human and Characterization of SLC26A6, A Candidate Gene for Pancreatic Anion Exchanger. Genomics 70:102-112)
		9431453	3.10E-276	Sulfate anion transporter [Rattus norvegicus] (Bissig,M. et al. (1994) Functional expression cloning of the canalicular sulfate transport system of rat hepatocytes. J. Biol. Chem. 269:3017-3021)
14	7478795CD1	g6045150	0	TAP-like ABC transporter [Rattus norvegicus] (Yamaguchi, Y. et al. (1999) An ABC transporter homologous to TAP proteins. FEBS Lett. 457:231-236)
15	656293CD1	g6746563	1.30E-220	neuronal nicotinic acetylcholine receptor [Rattus norvegicus]
16	7473957CD1	g340199	1.20E-130	voltage-dependent anion channel [Homo sapiens] (Blachly-Dyson, E. et al. (1993) J. Biol. Chem. 268:1835-1841)
1.7	7474111CD1	g6006493	1.50E-75	Cardiac potassium channel subunit (Kv6.2) [Homo sapiens] (Zhu, X., et al. (1999) Receptors Channels 6:337-350)
18	7480826CD1	g8248427	1.50E-235	amino acid transporter system A [Rattus norvegicus] (Sugawara, M. et al. (2000) J. Biol. Chem. 275:16473-16477)
19	6025572CD1	g402628	4.20E-114	adenine nucleotide carrier [Mus musculus]

Table 2 (cont.)

Polypeptide	Incyte	GenBank ID	Probability	Probability GenBank Homolog
SEQ ID NO:		NO:	score	
20	5686561CD1	g4586963	2.40E-27	voltage-gated ca channel [Rattus norvegicus] (Ishibashi, K. et al. (2000) Molecular cloning of a novel form (Two-repeat) protein related to voltage-gated sodium and calcium channels. Biochem. Biophys. Res. Commun. 270:376)
21	1553725CD1	g545998	1.60E-89	tricarboxylate carrier [Rattus sp.] (Azzi, A. et al. (1993) The mitochondrial tricarboxylate carrier. J. Bioenerg. Biomembr. 25:515-524)
22	1695770CD1	g31849	1.10E-175	inhibitory glycine receptor [Homo sapiens] (Grenningloh, G. et al. (1990) Alpha subunit variants of the human glycine receptor: primary structures, functional expression and chromosomal localization of the corresponding genes. EMBO J. 9:771-776)
23	4672222CD1	g13562153	0	channel-kinase 1 [Homo sapiens] (Ryazanov, A. G. et al. (1999) Alpha-kinases: a new class of protein kinases with a novel catalytic domain Curr. Biol. 9:R43-R45)
24	6176128CD1	g3978472	0	potassium channel subunit [Rattus norvegicus] (Joiner, W.J. et al. (1998) Formation of intermediate- conductance calcium-activated potassium channels by interaction of Slack and Slo subunits. Nat Neurosci. 1:462-469)
25	7473418CD1	g2811122	2.90E-177	NaDC-2 [Xenopus laevis]
26	7474129CD1	g2570933	1.20E-134	vanilloid receptor subtype 1 [Rattus norvegicus] (Caterina, M.J. et al. (1997) The capsaicin receptor: a heat-activated ion channel in the pain pathway. Nature 389:816-824)

Table 2 (cont.)

Polypeptide	Incyte	GenBank ID	Probability	GenBank Homolog
SEQ ID NO:		NO:	score	
27	7481414CD1	g13445630	1.00E-151	mutant ornithine transporter 2 [Mus musculus]
				(Wu, Q. and Maniatis, T. (1999) A striking
				organization of a large family of human neural
				cadherin-like cell adhesion genes. Cell 97:779-790)
28	7481461CD1	9458247	1.40E-136	X-linked PEST-containing transporter [Homo sapiens]
				(Lafreniere, R.G. et al. (1994) A novel transmembrane
				transporter encoded by the XPCT gene in Xq13.2.
				Mol. Genet. 3:1133-1139)
29	7472541CD1	g6457270	0	Putative E1-E2 ATPase [Mus musculus]
				(Halleck, M.S. et al. (1999) Differential expression of
				putative transbilayer amphipath transporters.
				Physiol. Genomics (Online) 1:139-150)
30	6999183CD1	g1514530	2.30E-127	ABC-C transporter [Homo sapiens]
				(Klugbauer N. and Hofmann F. (1996) Primary structure of
				a novel ABC transporter with a chromosomal localization
-				on the band encoding the multidrug resistance-
				associated protein, FEBS Lett. 391:61-65)

Table 3

			_					_		T			F.	_		 		1		-
Analytical Methods and Databases	SPScan	HMMER	BLAST-DOMO			HMMER	HMMER-PFAM	HMMER-PFAM	ProfileScan	MOTITED	27102	MOTIFS	BLIMPS-PRODOM		BLAST-DOMO			BLAST-PRODOM		
U. H.	Signal peptide: M1-A17	Transmembrane domains: W197-V224, Y248-G270		DM05037 P53988 1-465:M1-L109, L126-K289 DM05037 Q03064 1-475:M1-L109, V110-K289	DM05037 P36021 155-612:G3-G288	86 Transmembrane domains: P25-W49, Q82-I107, L166-L187, P184-M203	ABC transporter: H392-G575	ABC transporter transmembrane region: S30-A319	ABC transporters family signature:	DRC transporter.	F502-V516	ATP/GTP binding site: G399-S406	er:	PD00131:G141-D150, S403-I456, G550-R587	family:	DM00008 QT0183 1739-1448: 1363-63/3	\$55105 1272-1462: \$64757 1302-1528;	port protei	T61-G292	PD002040: G434-P488
Potential Glycosyla- tion Sites						N216 N386 N62 N68														
Potential Phosphorylation Sites	S287 S51 T132					S13 S40	S604 S63 T112 T253 T318 T330	T388 T455 T543 T70						***						
Amino Acid Residues	308					909														
Incyte Polypeptide ID	2194064CD1					2744094CD1														
SEQ NO:						7										 				

Table 3 (cont.)

д <u>2</u>			FORESTRAT	131313334	to the second of	15010 21511
	Polypeptide Acid	Acid	Phosphorylation	1 Glycosyla-	osphorylation Glycosyla-Domains and Motifs	Methods and
	TD OI	Residues	Sites	tion Sites		Databases
3	2798241CD1	1642	S199 S32 T1366	N190 N388	Transmembrane domains:	HMMER
	-		S431 S443 S1367	S443 S1367 N458 N499	Q34-M52, S272-P292, S295-F313, V327-	
			9	S546 T1390 N576 N86	I346,	
				S616 S1405 N943 N973	I401-L427, V865-H883, P1075-Y1098,	
				S761 T1454 N996 N1245		
				L N1556	ABC transporter:	HMMER-PFAM
			S88	3 N1627	G507-G689, G1326-G1509	
			S962 T28 T1635		ABC transporters family signature:	ProfileScan
			T486 T518 T1099	•	V595D646, I1413-D1464	
			T572 T606 T112	10	ABC transporter:	MOTIFS
			T779 T780 S1190		L615-V629	
			T854 Y168 S1236		ATP/GTP binding sites:	MOTIFS
			S1247 S1308		G514-S521, G1333-S1340	
			S1372 T1429		ABC transporters family:	BLAST-DOMO
			X1552		DM00008 P41233 839-1045:1478-S687,	
					K1313-M1506	
					ABC transporters family:	BLAST-DOMO
					DM00008 P34358 611-816:1478-S687,	
					I1319-M1506	
					DM00008 P26050 8-212:K1313-S1508,	
					1478-1686	
					DM00008 P41233 1851-2058:R1309-S1508, 1478-1686	

Table 3 (cont.)

SEC.		Amino	Potential	Potential 6	Potential Signature Semences	Analytical
) F	Dolymentide Arid	2,04	Phosphory at ion	L Accession	Dhocaphory ation (2) woody a lower man work for	Methods and
 0 N	QI QI	lues		tion Sites		Databases
4	3105257CD1	629	S206 S26 S300	N131 N210	N131 N210 ABC transporter: G441-G628	HMMER-PFAM
			S452 S504 S583		ABC transporter transmembrane region:	HMMER-PFAM
	-		S62 T261 T284		L92-I366	
			T293 T348 T520	7	ABC transporters family signature:	ProfileScan
			T615 Y121 Y298		A535-D586	
_					ABC transporter:	MOTIFS
					L555-L569	
					ATP/GTP binding site:	MOTIFS
					G448-S455	
					ABC transporters family:	BLIMPS-BLOCKS
					BL00211: L446-V457, L555-D586	
				<u>, · N</u>	ATP-binding transporter: PD00131:	BLIMPS-PRODOM
					G190-D199, S452-I505, G603-L640	
					ABC transporters family:	BLAST-DOMO
					DM00008 A42150 367-576: L413-L625	
					DM00008 P34712 1076-1290: F415-G628	
_					ATP-binding transport protein:	BLAST-PRODOM
					PD000130: L135-Y358	
		0			Multidrug resistance ATP-binding	BLAST-PRODOM
					transport protein:	
					PD167072: W486-G552	

Table 3 (cont.)

	Charles A section 1			The second secon		
SEQ	Incyte	0	Potential	Potential	Potential Signature Sequences,	Analytical
A	Polypeptide Acid		Phosphorylation	on Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
SON SON	ជ	dues	Sites	tion Sites		Databases
2	3200979CD1	1592	S125 S187 T1117 N185 N62		Transmembrane domains:	HMMER
-			S207 S386 T1135 N75 N870	35 N75 N870	I265-V285, L296-I315, M319-L340,	
			S453 Y906 T12:	Y906 T1214 N871 N899	I390-F410, L815-M834, L1063-M1082,	
-			14 S733	T1346 N949 N1164	W1099-T1117, L1126-L1145	
			S745 S770 T1388 N1273	38 N1273	ABC transporter:	HMMER-PFAM
			S778 S874 T1417	17	G500-G642, G1281-G1465	
			S882 S994 S1454	54	ABC transporters family signature:	ProfileScan
			T368 T439 T1494	94	L1372-D1420	
				30	ATP/GTP binding sites:	MOTIFS
			T565 T673 S1116	91	G507-S514, G1288-S1295	
			T691 T706 S1206	96	ABC transporters family:	BLIMPS-BLOCKS
			T766 T1257 T782	82	BL00211: I505-L516, L1389-D1420	
			T801 T1264 T927	27	ABC transporters family:	BLAST-DOMO
			_	92	DM00008 P41233 839-1045: K1268-M1462,	
			S7 S1297 S1320		I471-P600, E587-N641	
			T77 S1328 T1434	34	DM00008 P34358 611-816: F1262-M1462,	
			T1466		I471-D592, E585-N641	
					DM00008 P41233 1851-2058: K1266-S1464,	
	_				I471-V584, V588-N641	
					DM00008 P23703 41-246:K1268-G1465,	
					V476-L609, E585-G642	

Table 3 (cont.)

SEO	Incyte	Amino	Potential	Potential	Signature Sequences,	Analytical
ü	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	ID	Residues	Sites	tion Sites		Databases
ဖ	6754139CD1	382	S124 S260 S340	,	Transmembrane domains:	HMMER
			S85 T337		A168-H191, V200-L217, Y233-N253, F361-L378	
				_	Neurotransmitter-dated ion channel:	HMMER-PFAM
				·	D2-L378	
					mitter-gated ion-channels	ProfileScan
					signature:	
_					V66-G120	
					Neurotransmitter-gated ion channel:	MOTIFS
					C86-C100	
					Neurotransmitter-gated ion channel:	BLIMPS-BLOCKS
					BL00236:M1-D26, Y155-S196, V43-N52,	
					D71-H109	
					Neurotransmitter-gated ion channel:	BLIMPS-PRINTS
					PR00252:T9-W25, L42-K53, C86-C100,	
		_			L162-N174	
					Nicotinic acetylcholine channel:	BLIMPS-PRINTS
***					PR00254:M1-L12, Y30-W44, I48-G60, V66-	
_					584	
	_				Neurotransmitter-gated ion channel:	BLAST-DOMO
					DM00195 P43144 5-478:M1-E296, R323-A381	
					DM00195 JH0173 14-503:M1-P314,	
-					L327-A381	
					DM00195 P09478 5-538:R4-L297, E296-A381	
					DM00195 P54131 3-491:M1-A312, L327-A381	
					Postsynaptic ion channel:	BLAST-PRODOM
					PD000153: M1-R262, S298-V377	

Table 3 (cont.)

Analytical	Methods and	Databases	HMMER	SPScan	HMMER	HMMER-PFAM	MOTIFS	BLIMPS-PRINTS	BLAST-DOMO			BLAST-PRODOM				SPScan		HMMER		HMMER	
Signature Sequences,	Phosphorylation Glycosyla-Domains and Motifs		Signal peptide: M1-V24	Signal peptide: M1-S33	Transmembrane domains: M677-T693, F931-I946	اا	ATP/GTP binding site: G373-T380	NMDA receptor signature: PR00177:M677-G702, F744-E771,F931-V955,	Glutamate receptor:	DM00247 P35436 615-886: T731-0993	DM00393 Q01097 377-614: G482-F728 DM00247 001097 616-887: T731-Y956		S170~X577	PD139812: M1-P169	PD124284: S986-S1115 PD00500: M670-E952		M1-A41	Transmembrane domains:	F95-L114, V167-F187	Transmembrane domains:	
Potential	Glycosyla-	tion Sites	N264 N285	N426 N549	N565 N709 N886 N965	N1015	N1069		. .			-1			_	N57 N86				96N 04N	
Potential	Phosphorylation	Sites	S110 S202 S1030 N145 S248 S303 S1080 N275	S317 S334 T1101 N296 S383 S448 S1098 N439			T693 T704 T741 T796 T85 T949	T997 Y1106								S193 S199 S91	T59			S205 S252 S267	74 1300 1323
٥		dues	1115		***************************************											295				384	
Incyte	Polypeptide Acid	ID	6996659CD1													7472747CD1				7474121CD1	
SEQ	日	NO:	7	=-							 					ω				თ	

Table 3 (cont.)

Analytical	Methods and	Databases	HWMER	F491-V510	ily: HMMER-PFAM		ofile: BLIMPS-BLOCKS	217-1268	BLAST-DOMO	R49-V456	: E67-P495	: K78-S485	S87-I481	tein: BLAST-PRODOM		L521-D579
Potential Signature Sequences,	hosphorylation Glycosyla-Domains and Motifs	33	N195 N198 Transmembrane domains:	F245-I265, N294-V311, F491-V510	Sulfate transporter family:	L229-T513	Sulfate transporters profile:	BL01130: G119-V172, T217-L268	Sulfate transporter:	DM01229 P40879 5-462: R49-V456	DM01229 P50443 49-505: E67-P495	DM01229 P45380 10-468: K78-S485	DM01229 Q02920 1-447: S87-I481	Sulfate transporter protein:	PD001121: V93-T197	PD001755: H641-R720, L521-D579
Potential	Glycosyla	tion Sites	B6TN S6TN	N596												
Potential	Phosphorylation	Sites	S200 S3 S407	S461 S475 S572	S651 S707 S738	S742 S748 S87	T15 T282 T60	X470 Y57								
Amino	Acid	Residues	694													
Incyte	Polypeptide Acid	ID	7475615CD1							:						
SEQ	A	NO:	10	-								***				

Table 3 (cont.)

SEO	SEQ Incyte	Amino	Potential	Potential	Potential Signature Sequences,	Analytical
A	Polypeptide Acid	Acid	Phosphorylation [1]	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	El I	Residues		tion Sites		Databases
11	7475656CD1	882	S102 S108 S13	N330 N640	Transmembrane domains:	HMMER
			S324 S360 S394	N770 N8	L139-F159, T242-L258, I366-L392	
			S395 S518 S544		Transmembrane region cyclic nucleotide	HMMER-PFAM
			S591 T190 T242		domain:	
			T649 T754 T799		Y209-I453	
			T869 Y240 Y529		Cyclic nucleotide-binding domain:	HMMER-PFAM
					K482-M570	
					Cyclic nucleotide-binding domain:	MOTIFS
					I494-I515	
					Cyclic nucleotide-binding site:	BLIMPS-BLOCKS
					BL00888: G491-V514, G527-L536	
					Cyclic nucleotide-binding domain:	BLAST-DOMO
					DM01165 A55251 333-706: H302-E576	
					DM01165 P29973 311-684: H302-E576	
					DM01165 Q03041 286-658: H302-E576	
-					DM01165 S52072 262-635: H302-R572	
					Cyclic nucleotide gated hyperpolarization BLAST-PRODOM	BLAST-PRODOM
					activated cation channel:	
					PD079330: P747-L882	
					PD089437: A627-M722	
					PD108745: M1-D62	
					PD151315: T577-Q626	

Table 3 (cont.)

SEO	Incyte	Amino	Potential	Potential	Signature Sequences,	Analytical
î	Polypeptide Acid		Phosphorylation	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
No:	CI.	dues	Sites	tion Sites		Databases
12	7480632CD1	1547	S134 S196 S1102 N194 N71		Transmembrane domains:	HMMER
			S216 S395 T1301 N84 N879	N84 N879	I274-V294, L305-I324, M328-L349,	
			S7 T1343 T1389 N880 N908	806N 088N	I399-F419, L824-M843, M946-1963,	
				N958 N1100	L1021-F1040, L1046-L1064, D1105-F1123	
_			S754 S779 S1405 N1228		ABC transporter:	HMMER-PFAM
			S787 S883 T1449		G509-G651, G1236-G1420	
			T107		ABC transporters family signature:	ProfileScan
			T377 T448 S1158		L1327-D1375	
			T493 T551 T1212		ATP/GTP binding sites:	MOTIFS
					G516-S523, G1243-S1250	
			00		ABC transporters family:	BLIMPS-BLOCKS
			75		BL00211: I514-L525, L1344-D1375	
			10 T86 S		ABC transporters family:	BLAST-DOMO
			T936 T975 S1283		DM00008 P41233 839-1045: K1223-M1417,	
			Y915 S462 T1421		I480-P609, E596-N650	
			Y1144		DM00008 P41233 1851-2058:R1220-S1419,	
					I480-V593, V597-N650	
_			_		DM00008 P34358 611-816:F1217-M1417,	
					I480-D601, E594-N650	
					DM00008 P23703 41-246:K1223-G1420,	
					V485-L618, E594-G651	

Table 3 (cont.)

		_	Γ	7	-		-			-	7		_		7	_	_		U)		7			7		7
Analytical	Methods and	BLAST-DOMO	BLAST-DOMO	BLAST-PRODOM				BLAST-PRODOM				BLAST-PRODOM				BLAST-PRODOM			SYNOTE SENT TE	YOOTT - C JUITED	HMMER		HMMER-PFAM			MOTIFS	
Signature Sequences,	Phosphorylation Glycosyla-Domains and Motifs	SULFATE TRANSPORTERS: DM01229 P45380 10-468: V15-R462	do TRANSPORTER; SULFATE;	PROTEIN TRANSPORT SULFATE TRANSPORTER	TRANSMEMBRANE PERMEASE INTERGENIC REGION	AFFINITY GLYCOPROTEIN	PD001255: L285-L498	SULFATE TRANSPORTER TRANSPORT PROTEIN	TRANSMEMBRANE GLYCOPROTEIN AFFINITY	SULPHATE HIGH PERMEASE	PD001121: L49-R136	SULFATE TRANSPORTER PROTEIN TRANSPORT	TRANSMEMBRANE AFFINITY GLYCOPROTEIN	SULPHATE HIGH DISEASE	PD001755: H607-R689, A508-F551	SULFATE ANION TRANSPORTER 1 CANALICULAR	SULFATE/CARBONATE ANTIPORTER TRANSPORT	TRANSMEMBRANE GLYCOPROTEIN	PD085148: D155-D191	BL01130: A180-V231, D72-L125	1	E67-Y87, L411-A428	Sulfate transporter family	Sulfate_transp:	M192-T502	Sulfate_Transporter:	F33-KITO
Potential	Glycosyla-	N155 N160																									
Potential	Phosphorylation	S278 S355 S367 S446 S464 S594	T114	83 Y519																							
o																											
Incyte	Polypeptide Acid	952742CD1									•				_												
SEQ.	<u>a</u> §	13					_								·-												

Table 3 (cont.)

SEQ	Incyte	Amino	Potential	Potential	Potential Signature Sequences,	Analytical
A	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	ΠD	Residues	Sites	tion Sites		Databases
14	7478795CD1	992	S161 S275 S28	N280 N508	MALK PROTEIN:	BLAST-DOMO
			S33 S354 S46	N524 N599	DM00130 S13426 168-477: L195-G502	
			S543 S571 S595	N761	ATP-BINDING TRANSPORT PROTEIN	BLAST-PRODOM
			S671 S702 S763		TRANSMEMBRANE GLYCOPROTEIN TRANSPORTER	
	·		T139 T153 T181		MULTIDRUG RESISTANCE ABC PGLYCOPROTEIN	
			T209 T311 T367		PD000130: V229-L455	
•			T377 T512 Y602		ATP-BINDING TRANSPORT TRANSMEMBRANE	BLIMPS-PRODOM
					REGION	
-	-				PD00131:G283-D292, S543-I596, K691-L728	
					Transmembrane domain:	HMMER
					V85-F104, V185-F204, L328-G347,	
					Y411-G431	
					ABC transporter transmembrane region.	HMMER-PFAM
	•				ABC_membrane: L188-M459	
					ABC transporter	HMMER-PFAM
					ABC_tran: G532-G716	
					Abc_Transporter:	MOTIFS
					L643-L657	
					ATP/GTP-binding site motif A (P-loop)	MOTIFS
		_			Atp_Gtp_A: G539-S546	
					ABC transporters family signature	PROFILESCAN
					atp_bind_transport.prf: 1625-D674	

Table 3 (cont.)

SEO	Incyte	Amino	Potential	Potential	Signature Sequences,	Analytical
ពួ	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-1	Phosphorylation Glycosyla- Domains and Motifs	Methods and
NO:	ID II	dues		tion Sites		Databases
15	656293CD1	450	S153 S192 S328 S408 T405	N40 N56	NEUROTRANSMITTER-GAIED ION-CHANNELS DM00195 P43144 5-478: A25-E364, R391-A449	BLAST_DOMO
	_				CHANNEL IONIC TRANSMEMBRANE GLYCOPROTEIN	BLAST_PRODOM
					POSTSYNAPTIC MEMBRANE RECEPTOR PRECURSOR SIGNAL PROTEIN	
					PD000153:S131-R361	
				<u> </u>	Neurotransmitter-gated ion channel	BLIMPS_BLOCKS
					BL00236:D139-H177, Y223-S264, V57-D94, V111-N120	
					NEUROTRANSMITTER-GATED Ion Channel	BLIMPS_PRINTS
					PR00252:T77-W93, L110-K121, C154-C168,	
				_1.5		השואדמת המאד זמ
				7	MICOLINIC ACELINCHOLINE RECEPTOR	CINTUA CAMTOR
					ENOCES#:VIS#=SISK, SO4=DOC, 190=WILE, III16-G128	
					signal peptide:	HMMER
					M1-G24	
					transmembrane domain:	HMMER
					A236-H259, V268-L285, Y301-N321,	
				1	F429-L446	
_					Neurotransmitter-gated ion-channel	HIMMER_PFAM
	-				neur_chan:A30-L446	
				-	Weurotr_Ion_Channel	MOTIFS
_					C154-C168	
					Neurotransmitter-gated ion-channels	PROFILESCAN
					signature	
			-		neurotr_ion_channel.prf:V134-G188	
,			9		signal_cleavage: M1-G24	SPSCAN
				4		

Table 3 (cont.)

SEO	Incvte	Amino	Potential	Potential	3,	Analytical
βĤ	Polypeptide Acid		Phosphorylation	Glycosyla-		Methods and
NO:	ID	Residues	Sites	tion Sites		Databases
16	7473957CD1	260	S114 S12 S211 T136 T227 T28	N215 N216	S114 S12 S211 N215 N216 EUKARYOTIC MITOCHONDRIAL PORIN T136 T227 T28 DM01893 P45879 1-282:S12-A260	BLAST_DOMO
			T47 T49 T63 T84		GEDEPENDENT OUTER	BLAST_PRODOM
					MEMBRANE PROTEIN MITOCHONDRION	
					ANIONSELECTIVE MITOCHONDRIAL VDAC	
			•	-	PD003211:A15-Q259	
					Eukaryotic mitochondrial porin	BLIMPS_BLOCKS
					BL00558:G33-L46, T57-S81	
						BLIMPS_PRINTS
					PR00185:G45-T60, E124-E135, Y224-D241	
				-		HMMER_PFAM
					Euk_porin:A5-A260	
					Eukaryotic_Porin	MOTIFS
					Y202-Y224	
				•	signature	PROFILESCAN
					eukaryotic_porin.prf:M16-S81	
17	7474111CD1	206	S194	N284		BLAST_DOMO
					DM00436 JH0595 144-307:P230-I366	
			S493 S57 T241		CHANNEL IONIC PROTEIN POTASSIUM SUBUNIT	BLAST_PRODOM
			T273 T357 T385	•	VOLTAGEGATED TRANSMEMBRANE CALCIUM	
					TRANSPORT ION	
					PD000141:F319-Y486	
					GNATURE	BLIMPS_PRINTS
					PR00169:F319-V339, M363-C389, E392-	
					E415, F427-M449, G456-F482, E211-P230,	
					P245-T273, I293-K316	
			-		transmembrane domain:	HMMER
					I253-C270, V356-A373, V394-L413	
					Ion transport protein	HIMMER_PFAM
					ion_trans:1263-1478	

Table 3 (cont.)

SEO	Incyte	Amino	Potential	Potential	Potential Signature Sequences,	Analytical
i E	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	a a	Residues Si	Sites	tion Sites		Databases
18	7480826CD1	506	S12 S22 S280	N254 N258	N254 N258 TRANSPORTER PROTEIN	BLAST_PRODOM
			S320 T125 T181	N27 N274	PD138374:H360-H506	
			T276 T349 T433	N278 N326	N278 N326 ACID AMINO PROTEIN TRANSPORTER PERMEASE	BLAST_PRODOM
				64N	TRANSMEMBRANE INTERGENIC REGION PUTATIVE	
					PROLINE	
					PD001875:S76-I394	
					transmembrane domain:	HIMMER
					A97-L116, L224-V243, L192-S210,	
					I330-T349, V375-F392, I416-I441,	
					I473-I493	
		-		_	Transmembrane amino acid transporter	HMMER_PFAM
					protein	
					Aa_trans:A95-S489	

Table 3 (cont.)

SEQ	Incyte	Amino.	Potential	Potential :	Potential Signature Sequences,	Analytical
A	Polypeptide Acid			Glycosyla-1	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	A	dues		tion Sites		Databases
19	6025572CD1	315	S53 T209 T245	1	MITOCHONDRIAL ENERGY TRANSFER PROTEINS DM00026 S31935 110-208:0120-K218	BLAST_DOMO
					EINS	BLAST_DOMO
				1	AT	BLAST_PRODOM
			-		MITOCHONDRION CARRIER MEMBRANE INNER MITOCHONDRIAL ADP/ATP	
					PD000117:518-V210	
				, ~	Mitochondrial energy transfer proteins	BLIMPS_BLOCKS
					BL00215:L25-Q49, I271-G283	
						BLIMPS_PRINTS
					PR00926:A229-M251, D23-T36, T36-V50,	
					G85-D105, T138-D156, Y186-F204	
				. •	ADENINE NUCLEOTIDE TRANSLOCATOR	BLIMPS_PRINTS
					PR00927:F20-A32, Y63-R84, T96-K108,	
					R123-G136, R164-L185, S225-	
					Y241,	
					E275-R290	
					Mitochondrial carrier proteins	HMMER_PFAM
					mito_carr:S19-F308	
					Mitoch Carrier:	MOTIFS
					P40-L48, P145-L153, P242-M250	
					Mitochondrial energy transfer proteins	PROFILESCAN
_					signature	
					mitoch_carrier.prf:F20-I73, F125-I176,	
					F222-I271	

Table 3 (cont.)

SEQ	_	Amino	tential	Potential	Potential Signature Sequences,	Analytical
A	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	nosphorylation Glycosyla- Domains and Motifs	Methods and
NO:	A	Residues	Sites	tion Sites		Databases
50	5686561CD1	540		904N 66EN	N399 N406 Transmembrane domains:	HIMMER
			S29 S327 S349		A77-Y100, Y220-L243, I259-L285,	
			S454 T527		V291-Y311, A369-F389	
					Sodium channel signature:	BLIMPS-PRINTS
_					PR00170:G362-F389, Y76-G105, L361-F389,	
					K109-G134	
					Calcium channel:	BLAST-DOMO
					DM00043 A55645 1137-1259: A250-V298	
					(P-value = 2.7e-5)	
					Voltage gated calcium channel	BLAST-PRODOM
					PD000032:Y221-G391, I460-F486, N423-	
					W443	
					(P-value = 1.1e-6)	
21	1553725CD1	322	S142 S217 S295	N123 N131	142 S217 S295 N123 N131 PROTEIN TRANSMEMBRANE CHROMOSOME PUTATIVE BLAST_PRODOM	BLAST_PRODOM
			39 T133 T168	N29	TRANSPORTER C17G6.15C TRANSPORT XV	
			T304 T62 Y315		READING FRAME	
_					PD006986:F8-L253	

		_							_				_		_		_			-		_				_	-		_
Analytical	Methods and	Databases	HMMER	HMMER		HMMER_PFAM			BLIMPS_BLOCKS				PROFILESCAN			BLIMPS_PRINTS			BLIMPS_PRINTS			BLAST_PRODOM				BLAST_DOMO		MOTIFS	
Potential Signature Sequences,	Glycosyla-Domains and Motifs	tion Sites	N72 Signal peptide: M1-A28	Transmembrane domains:	M255-I279, I320-I339	Neurotransmitter-gated ion-channel	domain:	P44-F341	Neurotransmitter-gated ion channels	signature BL00236:	V73-R110, I127-N136, N157-Y195, F242-	A283	Neurotransmitter-gated ion-channels	signature:	L152-E206	Neurotransmitter-gated ion-channel family BLIMPS_PRINTS	PR00252:R93-Y109, S126-E137, C172-C186,	F249-Q261	Gamma-aminobutyric acid A (GABAA)	receptor signature	PR00253:Y258-W278, A284-S305, I318-I339	CHANNEL IONIC TRANSMEMBRANE GLYCOPROTEIN	POSTSYNAPTIC MEMBRANE RECEPTOR PRECURSOR	SIGNAL PROTEIN	PD000153: R99-K347	NEUROTRANSMITTER-GATED ION-CHANNELS	DM00560 S18836 18-453: R24-D417	Neurotransmitter-gated ion channel motif:	C172-C186
Potential	Phosphorylation Glycosyla-	Sites	S108 S122 S163	S43 S56 T196	T239 T243 T410	T411 T88			-																				
Amino		dues	417		-	, -																							
Incyte	Polypeptide Acid	A	1695770CD1																										
SEQ	H	NO:	22		•				-							-										_			

Table 3 (cont.)

SEQ	Incyte	Amino	Potential	Potential	Potential Signature Sequences,	Analytical
日	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	TI OI	Residues	Sites	tion Sites		Databases
23	4672222CD1	1864	S103 S195 S196		Transmembrane domains:	HIMMER
			S2 S22 S406 S5	N715 N718	F858-M878, N999-L1022,	
			S547 S697 S727	N805 N925	V1079-Q1102	
			S757 S836 S87	N1058	PROTEIN MELASTATIN CHROMOSOME	BLAST_PRODOM
			S883 T115 T12	N1465	TRANSMEMBRANE C05C12.3 T01H8.5 I F54D1.5	
			T299 T318 T349	N1466	IV	
			T367 T508 T523	N1595	PD018035: Y108-L439	
			T529 T593 T603	N1773	PD039592:E597-N801	
			T615 T675 T778	N1849	PD151509:V974-P1063, W1030-K1253	
			T795 T842 Y327		PD022180:W434-R545	
			S1476 S1503			
			T1163 S1191			
::			S1361 S1413			
			T1430 S1493			
			S1526 S1555			
		•	S1614 T1631			
			S1633 T1742			
			T1758 S1850			
			T1245 S1410			
			S1456 T1471			
-			S1499 S1698			
			S1859 Y1220			
			Y1552		And the second s	

Table 3 (cont.)

Signature Sequences,	d Motifs Methods and Databases	ins: F264, L310-L330	CHANNEL POTASSIUM IONIC CALCIUMACTIVATED BLAST_PRODOM	TON SOBONTI ACITARIED FACIETIN	PD003090:R337-F629, I784-M889, L926-	X1003-E1033, Q1176-S1215	; POTASSIUM; MSLO; ACTIVATED; BLAST_DOMO	DM05442 A48206 351-1123: R337-F618,	F744-F785, QII/0-51220				Transmembrane domains:	V15-C38, C50-F67, F264-F282, A323-R341	Sodium: sulfate symporter signature: BLIMPS_BLOCKS	BL01271:S451-1505, T132-1151,	П	PROTEIN TRANSMEMBRANE TRANSPORT MEMBRANE BLAST_PRODOM	SECRETARION SINEONI OF	PD000549:V15-V173, M216-W518	do RENAL; BOUND; PRO-SER-ALA; NA; BLAST_DOMO	
otential Signatur	lycosyla-Domains	N100 N133 Transmer N137 N279 M155-Y		N1153	PD0030	1000	do CHANNEL;	DM0544	• • • • • • • • • • • • • • • • • • • •				N533 Transmen	V15-C3	Sodium: £	BL0127		PROTEIN	AETROCKNERTON	PD000	do RENAI	OH THE FOR OC PARCE VENCONCE
Potential Po	Phosphorylation Glycosyla-Domains and Motifs Sites	S102 S135 S139 NJ S168 S179 S361 NJ	S438 S439	S720 S726	2808	S954 T156 T302	T351 T391 T446	T517 T609 T718	177 1394 S1090 S1098 S1219	S1013 S1030	T1190 T1231	S1125 S1215 S1221	S299 S321 T535 N									
Amino	Acid Residues												539									
Incyte	ptide	6176128CD1											7473418CD1									_
SEO	日 日 日 日	24		····							_		25									

Table 3 (cont.)

SEQ	Incyte	Amino	Potential	Potential	Signature Sequences,	Analytical
A	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	ID	Residues	Sites	tion Sites		Databases
26	7474129CD1	755	S339 S353 S367	N417 N648	Transmembrane domains:	HMMER
			S463 S53 S572	N735	V490-F507, L556-L573, P616-M642	
			S589 S653 S732		Ank repeat:	HMMER PFAM
			T128 T132 T255		E179-K211, F226-S259, D305-K333	
 -			T270 T277 T300		VANILLOID RECEPTOR SUBTYPE 1 PD101189:	BLAST_PRODOM
_			T343 T358 T362		Q52-L291	
			T37 T376 T441		PROTEIN OLFACTORY CHANNEL B0212.5	BLAST_PRODOM
			T664 Y225 Y347		T09A12.3 T10B10.7 VANILLOID RECEPTOR	
			X587		SUBTYPE F28H7.10	
`					PD011151:N303-E430	
27	7481414CD1	301	S143 S203 S290		Transmembrane domain:	HMMER
			T136 T32		L212-V230	
					Mitochondrial carrier proteins domain:	HMMER_PFAM
					Q8-M294	
					Mitochondrial energy transfer proteins	BLIMPS_BLOCKS
					signature:	
					BL00215:L214-Q238, V256-G268	
					Mitochondrial energy transfer proteins	PROFILESCAN
					signature:	
	<u></u>				A10-G59, L107-I160, K204-A276,	
		•			K213-N259	
					PROTEIN TRANSPORT TRANSMEMBRANE REPEAT	BLAST_PRODOM
_		<i>'</i>			MITOCHONDRION CARRIER MEMBRANE INNER	
					MITOCHONDRIAL ADP/ATP PD000117:	
					Y44-S241	
					Mitochondrial carrier protein motifs:	MOTIFS
					P126-L134 P229-I237	

Table 3 (cont.)

SEQ	Incyte	Amino	Potential	Potential	Potential Signature Sequences,	Analytical
ព	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	日	Residues	Sites	tion Sites		Databases
28	7481461CD1	515	S10 S104 S163	N81	Transmembrane domains:	HMMER
			S257 S272 S277		V117-F135, Y169-L191, I190-I215,	
			S4 S474 S511		G229-F245, I376-F395	
			S97 T233 T250		Monocarboxylate transporter domain:	HMMER_PFAM
			T484		A77-A455	
					XLINKED PESTCONTAINING TRANSPORTER SOLUTE BLAST_PRODOM	BLAST_PRODOM
					CARRIER FAMILY MONOCARBOXYLIC ACID	
					TRANSPORTERS MEMBER	
:					PD030892: P33-V111	
					do PEST; TRANSPORTER; LINKED;	BLAST_DOMO
					DM05037 P36021 155-612: E63-M489	

Table 3 (cont.)

7472541CD1 1519 S223 S307 S432 N148 N298 S456 S472 S486 N339 N354 S498 S510 S538 N41 N51 S579 S628 S63 N69 N991 S648 S68 S701 N1249 S756 S779 S826 S832 S903 S912 S986 T275 T341 T437 T449 T466 T495 T563 T597 T64 T716 T73 T755 T805 T880 T945 T961 S1509 S1110 S1131 T1198 S1256 S1278 T1431 S1480 S1406 T1439
T4437 T44 T495 T56 T664 T67 T73 T755 T880 T94 S1509 S1 S1131 T1 S1256 S1

Table 3 (cont.)

SEQ	Incyte	Amino	Potential	Potential	Potential Signature Sequences,	Analytical
a	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	Phosphorylation Glycosyla-Domains and Motifs	Methods and
NO:	ID	Residues	Sites	tion Sites		Databases
30	6999183CD1	1585	S2, S7, T30,	N72, N121,	N72, N121, ABC TRANSPORTERS FAMILY:	BLAST-DOMO
			S61, S70, T86,	N196,	DM00008 P41233 839-1045: I1268-M1455,	
			S114, S198,	N245,	I482-P611, E598-N652	
			T267, T459,	N457,	ABC transporters family:	BLIMPS-BLOCKS
			T576,	N546,	BL00211: L516-L527, L1382-D1413	
				N557,	Transmembrane domain (transmem_domain):	HMMER
				N881,	I1058-L1082, I1099-L1117, G1124-I1147,	
			T757, S779,	N910,	L1167-M1193, T30-F48, T224-V242,	
			S789, T793,	N960,	W271-I289, T306-I326, P329-L346,	
			8885, 8893,	N1272,	F358-M375, Y398-Y420, V1034-F1053	
			Y917 S924,	N1337	ABC transporter (ABC_tran):	HMMER-PFAM
			T938, T967,		G511-G653, G1280-G1458	
			T971, S1005,		ATP/GTP-binding site motif A (P-loop)	MOTIFS
			.054,		(Atp_Gtp_A): G518-T525, G1287-S1294	
			158,		ABC transporters family signature	PROFILESCAN
			S1262, T1267,		(atp bind transport.prf): I1362-D1413	
			T1296, T1339,			
			T1381, T1410,			
_			T1427, T1431,			
			S1457, Y1544			
			S1574, S1549,			

Table 4

Polynucleotide	Incyte	Sequence	Selected	Sequence Fragments	5,	3,
SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)		Position	Position
	2194064CB1	1129	1071-1129,	g5110579	1	485
			833-898	FL2194064_g7770598_000019_g7 670446	203	1129
				6542780F9 (LNODNON02)	32	481
32	2744094CB1	2699	1-2196,	FL097646_00001	431	2542
			2541-2587	55058921H1	1	793
				70317743D1	2347	2699
				70317681D1	2209	2639
33	2798241CB1	6369	1-1210,	71911330V1	5832	6369
			1759-5012	70300809D1	5128	5690
					5650	6322
				7601441J1 (ESOGTME01)	4623	5186
				6314138H1 (NERDTDN03)	5235	5750
					4145	4636
					5764	6357
				4013186F9 (MUSCNOT10)	3758	4391
				7606344H1 (COLRIUE01)	1764	2219
				6913644H1 (PITUDIR01)	4608	5181
				55052455J1	1981	2827
				7400061H1 (SINIDME01)	1	502
				2798241T6 (NPOLNOT01)	1325	1955
				4	2548	3298
-				7100413F7 (BRAWIDR02)	483	1185
				6744456H1 (BRAFNOT02)	568	1274
				5505364771	3011	3823
				6586921H1 (TLYMUNTO3)	1157	1724
34	3105257CB1	2558	1-587,	70864718V1	1864	2353
			2435-2558	70549000V1	1608	2310
				O	1	1843
				6451207H1 (BRAINOC01)	1868	2558

Table 4 (cont.)

	_	-			_	_	_	_	-	-					_	_	_	_	-	~	_		_	_		-	_	_	=		
3,	Position	4779	5065	800	875	531	1677	883	874	1915	3006		2942	3707	2776	3714	3466	309	1209	1174	1994	1009		450	1155	1756		1221		1849	
5,	Position	1	4463	532	789	1	386	684	684	1180	1303		2752	3116	2605	3322	2967	1	510	219	1768	122		1	н	1580		986		1687	
Seguence Fragments		FL3200979_g3810670_g4240130	71698878V1	656293H1 (EOSINOTO3)	55062573H1	GBI:edit	GNN: 98017750_000028_004	g5678193	6754139J1 (SINTFER02)	6996659F8 (BRAXTDR17)	GBI.99211864_01_04_05_12.edi	t	55098348H2	1596150T6 (BRAINOT14)	7124651F6 (COLNDIY01)	g4622477	1596150F6 (BRAINOT14)	55063531J1	7291716R6 (BRAIFER06)	7291716F6 (BRAIFER06)	55063924J1	FL7472747_g6983242_000026_g3	925427	7616162H1 (COLNTUNO3)	GNN.97259672_000014_002	FL7475615_98980204_000002_g2	654005_1_11-12	FL7475615_g8980204_000002_g2	654005_1_6-7	FL7475615_g8980204_000002_g2	654005_1_12-13
Selected	Fragment(s)	5030-5065,	1-3313	1-686						1-1916,	3071~3091,	2092-2619										1-388, 571-	704, 778-	1009	1-1155	1852-2185,	1484-1579,	665-1340,	1-249,	2334-2733,	454-495
Sequence	Length	5065		1677						3714												6001			1155	2733					
Incyte	Polynucleotide ID	3200979CB1		6754139CB1						6996659CB1												7472747CB1			7474121CB1	7475615CB1					
Polynucleotide	SEQ ID NO:	35		36	•					37												38			39	40					

Table 4 (cont.)

Polynucleotide	Incyte	Sequence	Selected	Sequence Fragments	5,	3,
SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)		Position	Position
				FL7475615_g8980204_000002_g2	1139	1369
				654005_1_7~8		
==				55092029J1	341	1088
				55083049H1	1	470
				1509180F6 (LUNGNOT14)	1744	2228
				FL7475615_98980204_000002_g2	1222	1483
				654005_1_8-9		
				GNN.97342135_000012_002	821	1579
				6806177J1 (SKIRNOR01)	1995	2733
				FL7475615_g8980204_000002_g2	1484	1686
				654005_1_10-11		
41	747565CB1	3457	3284-3346,	55073909H2	1	110
			1169-1646,	g3168873_CD	382	2628
-			1-290,	7946572H1 (BRABNOE02)	228	536
			2835-2868,	5373417T9 (BRAINOT22)	2245	2901
			2018-2292,	GNN.g6532090_000006_000019.e	43	867
			3030-3174,	dit		
			780-844,	2428507R6 (SCORNON02)	2911	3457
			456-653	5373417F8 (BRAINOT22)	1396	1620
					2879	3160
				4787380H1 (BRATNOT03)	1461	1717
.42	7480632CB1.	5622	1-3676,	1450339F1 (PENITUT01)	4134	4646
			5557-5622	7270152H1 (OVARDIJ01)	167	646
				71697049V1	4737	5460
				3488927H1 (EPIGNOT01)	2908	3106
				6774619J1 (OVARDIR01)	2052	2720
				5063703F6 (ARTFTDT01)	4886	5622
				55072886J1	3675	4262
				GBI.g3810670_000001.edit	266	4674
				6488228F9 (MIXDUNB01)	1	632
				7670233H2 (BONRNOC01)	4317	4878

Table 4 (cont.)

Polynucleotide	Incyte	Seguence	Selected	Sequence Fragments	5,	3,
SEQ ID NO:	Polymucleotide ID	Length	Fragment(s)		Position	Position
43	6952742CB1	2600	2329-2600,	5884027F8 (LIVRNONO8)	2088	2600
			1-224,	55063579H1	779	1276
			1190~1560,	GBI:g7232144_000013.edit.3	1626	2351
			1957~2046,	6816048H1 (ADRETURO1)	1	316
			1006-1030	6952742H1 (BRAITDR02)	1140	1824
				GNN.96970605_000013_002	342	1355
				GBI:g7232144_000013.fasta.ed	255	506
				it		
44	7478795CB1	2917	2698-2917,	72016954V1	2193	2917
			1808~2065,	71989431V1	2165	2912
			398-714,	72017820V1	1369	2155
			923-976	72017055V1	1160	2053
				72017371V1	570	1212
				72017076V1	1958	2859
		110		72017430V1	476	1146
				55076285H1	.7	995
45	656293CB1	1474	1-362	GBI.g8017750_edit	T	1353
				FL656293_g8017750_000028_g67 46563_2_2-3	130	568
				FL656293_g8017750_000028_g67	363	1353
				46563_2_3~4		
				7675576H1 (NOSETUE01)	907	741 4
46	7473957CB1	1742	1-367,	4648731F9 (PROSTUT20)	610	1274
			1680-1742	71166638V1	1	019
				71165785V1	1010	1742
				6830443J1 (SINTNOR01)	592	1266
47	7474111CB1	2312	1-639,		9	632
			1686-1712,		1692	2312
			2004-2312,	7761487J1 (THYMNOE02)	1	905
			1860-1908	GNN.g7243948_CDS_1	183	1845

Table 4 (cont.)

Polymucleotide	Theyte	Segnence	Selected	Sequence Fragments	5,	3,
SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)		Position	Position
48	7480826CB1	2320	161-224,	7752763J1 (HEAONOE01)	1668	2320
			2044-2320	60143671D1	467	917
				6052064J1 (BRABDIR03)	1080	1658
				6484950H1 (MIXDUNB01)	1276	1723
				2944045H1 (BRAITUT23)	827	1118
				7469461H1 (LUNGNOE02)	1	498
49	6025572CB1	1781	1-170	FL6025572_g7382154_000015_g1 197164	347	1063
				4923834H1 (TESTNOT11)	1	291
				g3838735	1313	1781
				g3734777	252	472
				TATT904614	1285	1780
				6025572F6 (TESTNOT11)	883	1627
50	5686561CB1	2433	1-1078,	H	1088	1702
			1197-1275	6060785H1 (BRAENOTO4)	551	1100
				7695065J1 (LNODTUE01)	387	1052
					н	483
				- [2148	2433
					1765	2304
				5564984F6 (TLYMNOT08)	860	1528
-				70730430V1	1525	2108
51	1553725CB1	1772	1571-1772	60211064U1	344	823
				72050509V1	1176	1772
				70300327D1	984	1428
				7030706D <u>1</u>	1	262
				1553725X15C1 (BLADTUT04)	54	694
				70300332D1	729	1286
52	1695770CB1	1874	1-479,	55117454H1	1155	1874
			1298-1874,	55110123H1	286	1179
			1131-1216, 886-984	55072985J1	T	542

Table 4 (cont.)

3,	sition	2815	6211	5344	4598	3162	5239	3886	1499	3318	4537	8	1915	3903	2436	5927	5866	1143	2953	3714	3711	3324	1503	2374	3088	2006	1053	5	0;	3115
3	Po	28	62	53	45	31	52	38	14	33	45	878	13	39	24	55	58	11	29	37	37	33	15	23	30	20	10	645	740	31
5,	Position	1925	5663	4613	3913	2738	4522	3346	818	2942	3811	1	1086	2957	1797	5374	5325	7	2265	3612	3115	2991	899	1741	2586	1383	543	383	369	2658
Sequence Fragments		55047368J1	71007436V1	71998604V1	71995592V1	3462433F7 (293TF2T01)	71997753V1	71995863V1	55073038H1)	5514117701	71998657V1	6141577F6 (BMARTXT03)	55140386J1	GBI:g8189326.edit	1		2505959F6 (CONUTUTO1)	GBI.g979669_000005_000004.ed	6859776H1 (BRAIFEN08)	GBI.9979669_000002.edit	GBI.g7739135_000005.edit	6772216J1 (BRAUNOR01)	6887873J1 (BRAITDR03)	8039114H1 (SPLNNOE01)	-	- 1	6891702F6 (BRAITDR03)	7065904R6 (BRATNOR01)	FL7473418_g3176728_g5531902_ 1 4-5	7056016H1 (BRALNON02)
Selected	Fragment(s)	3238-3683,	4625-4798,	2313-2462,	1-1636													1-197, 329- 2513, 3301-	3336										1-1411	
Sequence	Length	6211																3714											3115	
Incyte	Polynucleotide ID	4672222CB1																6176128CB1											7473418CB1	
Polynucleotide	SEQ ID NO:	53																54											55	

Table 4 (cont.)

Polynucleotide	Incyte	Sequence	Selected	Sequence Fragments	5,	3,
SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)		Position	Position
				FL7473418_g3176728_g5531902_ 1_7-8	864	1188
				1324158F6 (LPARNOT02)	2114	2695
				6899347H1 (LIVRIMR01)	1074	1568
				FL7473418_g3176728_g5531902_	548	863
				70075691U1	2291	2782
				FL7473418_g3176728_g5531902_	739	1069
				1_6-7		
				FL7473418_g3176728_g5531902_ 1_1-2	Ħ	231
				FL7473418_g3176728_g5531902_	1351	1620
				1_10-11		
				FL7473418_g3176728_g5531902_ 1_2-3	103	368
				7114876H1 (BRAENOK01)	1549	1954
				FL7473418_g3176728_g5531902_	232	547
				1 3 - ¢		
				4895008F6 (LIVRTUT12)	1752	2240
56	7474129CB1	2846	1-1696,	55109928H1	2480	2846
			2073-2846,	55109306J1	1837	2660
	*		1777-2012	55124533H1	.1	832
				55124525H1	1073	1893
				55073088J1	796	1208
57	7481414CB1	906	441-541,	GBI.g9454493_000005_000056.e	1	906
			262-348	dit		

Table 4 (cont.)

Polynucleotide		Sequence	Selected	Seguence Fragments	5,	3,
	Polynucleotide ID	Length	Fragment(s)		Position	Position
	7481461CB1	1840	1-91	70481006V1	551	1137
				70465445V1	673	1250
				1748722F6 (STOMTUT02)	1423	1840
				60266587D1	1225	1801
				7637372J1 (SINTDIE01)	221	642
				g5933739	1	407
ட்	7472541CB1	5348	1384-1560,	6772907J1 (BRAUNOR01)	1641	2204
			1-1188,	2182261F6 (SININOT01)	4301	4634
			4239-4906,	5459667H1 (SINITUTO3)	3276	3550
			2145-2970,	GNN.g7710567_000006_002.edit	1057	2001
			4944-5348	7362215H1 (BRAIFEE05)	7	526
_				GNN.g7708823_000019_002	4306	4862
				7032970H1 (BRAXTDR12)	2692	3426
				7313608H1 (BRABDIE02)	731	1260
				71462931V1	4732	5348
				5767060H1 (STOMFET02)	3887	4428
		-		8069315J1 (BRAIFEE05)	91	792
				7582660H1 (BRAIFEC01)	3510	4131
				GNN.g7454125_000004_002.edit	1382	3918
_				6772907H1 (BRAUNOR01)	2476	2994
	6999183CB1	5149	1-1797,	GBI.g3873182_000001.edit5p	1	3167
			4753-4852,	72017145V1	4087	4903
			3028-3711,	6999183R8 (HEALDIRO1)	384	1128
			2471-2667	55051672H1	606	1573
		-		72017349V1	3836	4767
				72293922V1	2616	3434
				55144835H1	2154	2939
				55144834J1	3263	3947
_				55076606J1	1126	1592
				72017610V1	4219	5149

Table 5

Polynucleotide	Incyte	Representative Library
SEQ ID NO:	Project ID	
31.	2194064CB1	THYRTUT03
32	2744094CB1	BRSTTUT15
33	2798241CB1	PROSTME06
34	3105257CB1	BLADNOT01
35	3200979CB1	PENITUT01
36	6754139CB1	BRSTNOR01
37	6996659CB1	BRAIFER06
38	7472747CB1	COLNTUNO3
40	7475615CB1	LUNGNON07
41	7475656CB1	BRAINOT22
42	7480632CB1	PENITUT01
43	6952742CB1	LIVRNON08
77	7478795CB1	BRAENOT02
45	656293CB1	COLNNOT22
46	7473957CB1	BRAHTDR03
47	7474111CB1	THYMNOE02
48	7480826CB1	MIXDUNB01
49	6025572CB1	TESTNOT11
50	5686561CB1	BRAENOT04
51	1553725CB1	THYMNON04
52	1695770CB1	COLINIOT23
53	4672222CB1	PITUDIR01
54	6176128CB1	BRAITDR03
55	7473418CB1	LPARNOT02
56	7474129CB1	PLACNOT05
58	7481461CB1	OVARTUTO5
59	7472541CB1	BRAIFEE05
60	6999183CB1	HEALDIR01

Table 6

Library	Vector	Library Description
BLADNOT01	PBLUESCRIPT	Library was constructed using RNA isolated from the bladder tissue of a 78-year-old Caucasian female, who died from an intracranial bleed. Patient history included basal cell carcinoma, arthritis, and chronic hypertension.
BRAENOT02	pincy	Library was constructed using RNA isolated from posterior parietal cortex tissue removed from the brain of a 35-year-old Caucasian male who died from cardiac failure.
BRAENOT04	pINCY	Library was constructed using RNA isolated from inferior parietal cortex tissue removed from the brain of a 35-year-old Caucasian male who died from cardiac failure. Pathology indicated moderate leptomeningeal fibrosis and multiple microinfarctions of the cerebral neocortex. Patient history included dilated cardiomyopathy, congestive heart failure, cardiomegaly and an enlarged spleen and liver.
BRAHTDR03	PCDNA2.1	This random primed library was constructed using RNA isolated from archaecortex, anterior hippocampus tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorhinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydorthorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and resection of 85% of the liver.
BRAIFEE05	PCDNA2.1	This 5' biased random primed library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation.
BRAIFER06	PCDNA2.1	This random primed library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation. Serologies were negative.

Library	Vector	Library Description
BRAINOT22	pincy	Library was constructed using RNA isolated from right temporal lobe tissue removed from a 45-year-old Black male during a brain lobectomy. Pathology for the associated tumor tissue indicated dysembryoplastic neuroepithelial tumor of the right temporal region dura was consistent with calcifying pseudotumor of the neuraxis. Family history included obesity, benign hypertension, cirrhosis of the liver, obesity, hyperlipidemia, cerebrovascular disease, and type II diabetes.
BRAITDR03	PCDNA2.1	This random primed library was constructed using RNA isolated from allocortex, cingulate posterior tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorhinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydorthorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and resection of 85% of the liver.
BRSTNOR01	pINCY	Library was constructed using RNA isolated from diseased breast tissue removed from a 59-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology for the associated tumor tissue indicated an invasive lobular carcinoma with extension into ducts. Patient history included cirrhosis, esophageal ulcer, hyperlipidemia, and neuropathy.
BRSTTUT15	DINCK .	Library was constructed using RNA isolated from breast tumor tissue removed from a 46-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology indicated invasive grade 3, nuclear grade 2 adenocarcinoma, ductal type. An intraductal carcinoma component, non-comedo, comprised approximately 50% of the neoplasm, including the lactiferous ducts. Angiolymphatic involvement was present. Metastatic adenocarcinoma was present in 7 of 10 axillary lymph nodes. The largest nodal metastasis measured 3 cm, and focal extracapsular extension was identified. Family history included atherosclerotic coronary artery disease, type II diabetes, cerebrovascular disease, and depressive disorder.

Libramy	Vertor	Tibrary Description
COLINIOT22	pINCY	Library was constructed using RNA isolated from colon tissue removed from a 56- Library was constructed using RNA isolated from colon tissue removed from a 56- year-old Caucasian female with Crohn's disease during a partial resection of the small intestine. Pathology indicated Crohn's disease of the ileum and ileal- colonic anastomosis, causing a fistula at the anastomotic site that extended into pericolonic fat. The ileal mucosa showed linear and puncture ulcers with intervening normal tissue. Previous surgeries included a partial ileal resection and permanent ileostomy. Family history included irritable bowel syndrome in the mother and the siblings.
COLMOT23	pincy	Library was constructed using RNA isolated from diseased colon tissue removed from a 16-year-old Caucasian male during a total colectomy with abdominal/perineal resection. Pathology indicated gastritis and pancolonitis consistent with the acute phase of ulcerative colitis. Inflammation was more severe in the transverse colon, with inflammation confined to the mucosa. There was only mild involvement of the ascending and sigmoid colon, and no significant involvement of the cecum, rectum, or terminal ileum. Family history included irritable bowel syndrome.
COLNTUNO3	pincy	This normalized pooled colon tumor tissue library was constructed from 1.16 million independent clones from a pooled colon tumor library. Starting library was constructed using pooled cDNA from 6 donors. cDNA was generated using mRNA isolated from colon tumor tissue removed from a 55-year-old Caucasian male (A) during hemicolectomy; from a 60-year-old Caucasian male (B) during hemicolectomy; from a 64-year-old Caucasian female (E) during hemicolectomy; from a 64-year-old Caucasian female (E) during hemicolectomy; and from a 70-year-old Caucasian female (E) during hemicolectomy. Pathology indicated invasive grade 3 adenocarcinoma (A); invasive grade 2 adenocarcinoma (B); invasive grade 2 adenocarcinoma (F). Donors B, C, D, E, and F had positive lymph nodes. Patient medications included Ativan (A); Seldane (B), Tri-Levlen (D); Synthroid (E); Tamoxifen, prednisone, Synthroid, and Glipizide (F). The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.

Library	Vector	Library Description
HEALDIR01	PCDNA2.1	This random primed library was constructed using RNA isolated from diseased left
		ventricle tissue removed from a 7-month-old Caucasian male who died from
		cardiopulmonary arrest due to Pompe's disease. Patient history included Pompe's
		disease, left ventricular hypertrophy, pyrexia, right complete cleft lip, cleft
		palate, chronic serous otitis media, hypertrophic cardiomyopathy, congestive heart
		failure, and developmental delays. Family history included acute myocardial
		infarction, diabetes, cystic fibrosis and Down's syndrome.
LIVRNON08	DINCY	This normalized library was constructed from 5.7 million independent clones from a
		pooled liver tissue library. Starting RNA was made from pooled liver tissue
		removed from a 4-year-old Hispanic male who died from anoxia and a 16 week female
		fetus who died after 16-weeks gestation from anencephaly. Serologies were positive
		for cytolomegalovirus in the 4-year-old. Patient history included asthma in the 4-
		year-old. Family history included taking daily prenatal vitamins and mitral valve
		prolapse in the mother of the fetus. The library was normalized in 2 rounds using
		conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al.,
		Genome Research 6 (1996):791, except that a significantly longer (48 hours/round)
		reannealing hybridization was used.
LPARNOT02	PINCY	Library was constructed using RNA isolated from tissue obtained from the left
		parotid (salivary) gland of a 70-year-old male with parotid cancer.
LUNGNON07	DINCY	This normalized lung tissue library was constructed from 5.1 million independent
		clones from a lung tissue library. Starting RNA was made from RNA isolated from
		lung tissue. The library was normalized in two rounds using conditions adapted
		from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research
		(1996) 6:791, except that a significantly longer (48 hours/round) reannealing
		hybridization was used.

	17004000	
MIXDUNBOL	PINCY	Library Description Library was constructed using RNA isolated from myometrium removed from a 41-year- old Caucasian female (A) during vaginal hysterectomy with a dilatation and curettage and untreated smooth muscle cells removed from the renal vein of a 57- year-old Caucasian male. Pathology for donor A indicated the myometrium and cervix were unremarkable. The endometrium was secretory and contained fragments of endometrial polyps. Benign endo- and ectocervical mucosa were identified in the endocervix. Pathology for the associated tumor tissue indicated uterine leiomyoma. Medical history included an unspecified menstrual disorder, ventral herhia, normal delivery, a benign ovarian neoplasm, and tobacco abuse in donor A. Previous surgeries included a bilateral destruction of fallopian tubes, removal of a solitary ovary, and an exploratory laparotomy in donor A. Medications included ferrous sulfate in donor A.
OVARIUTOS	pincy	Library was constructed using RNA isolated from ovarian tumor tissue removed from a 62-year-old Caucasian female during a total abdominal hysterectomy, removal of the fallopian tubes and ovaries, exploratory laparotomy, regional lymph node excision, and dilation and curettage. Pathology indicated a grade 4 endometrioid carcinoma with extensive squamous differentiation, forming a solid mass in the right ovary. The uterine endometrium was inactive, the cervix showed mild chronic cervicitis, and focal endometriums was observed in the posterior uterine serosa. Curettings indicated weakly proliferative endometrium with excessive stromal breakdown in the uterus, and a prior cervical biopsy indicated mild chronic cervicitis with a prominent nabothian cyst in the cervix. Patient history included longitudinal deficiency of the radioulna, osteoarthritis, thrombophlebitis, and abnormal blood chemistries. Family history included atherosclerotic coronary artery disease, pulmonary embolism, and cerebrovascular disease.
PENITUT01	pincy	Library was constructed using RNA isolated from tumor tissue removed from the penis of a 64-year-old Caucasian male during penile amputation. Pathology indicated a fungating invasive grade 4 squamous cell carcinoma involving the inner wall of the foreskin and extending onto the glans penis. Patient history included benign neoplasm of the large bowel, atherosclerotic coronary artery disease, angina pectoris, gout, and obesity. Family history included malignant pharyngeal neoplasm, chronic lymphocytic leukemia, and chronic liver disease.

		Library Description
_	PCDNA2.1	This random primed library was constructed using RNA isolated from pituitary gland
L		tissue removed from a 70-year-old female who died from metastatic adenocarcinoma.
PLACNOTOS DI	DINCY	Library was constructed using RNA isolated from placental tissue removed from a
		Caucasian male fetus, who died after 18 weeks' gestation from fetal demise.
PROSTME06 PC	PCDNA2.1	This 5' biased random primed library was constructed using RNA isolated from
		diseased prostate tissue removed from a 57-year-old Caucasian male during closed
		prostatic biopsy, radical prostatectomy, and regional lymph node excision.
	-	Pathology indicated adenofibromatous hyperplasia. Pathology for the matched tumor
		tissue indicated adenocarcinoma, Gleason grade 3+3, forming a predominant mass
		involving the right side centrally. The patient presented with elevated prostate
		specific antigen and prostate cancer. Patient history included tobacco abuse in
		remission. Previous surgeries included cholecystectomy, repair of diaphragm
		hernia, and repair of vertebral fracture. Patient medications included Pepsid,
		Omnipen, and Eulexin. Family history included benign hypertension, cerebrovascular
		accident, atherosclerotic coronary artery disease, uterine cancer and type II
		diabetes in the mother; prostate cancer in the father; drug abuse, prostate
		cancer, and breast cancer in the sibling(s).
TESTNOT11 pi	pINCY	Library was constructed using RNA isolated from testicular tissue removed from a
		16-year-old Caucasian male who died from hanging. Patient history included drug
		use (tobacco, marijuana, and cocaine use), and medications included Lithium,
		Ritalin, and Paxil.

Library	Vector	Library Description
THYMNOE02	PCDNA2.1	This 5' biased random primed library was constructed using RNA isolated from
		thymus tissue removed from a 3-year-old Hispanic male during a thymectomy and
		closure of a patent ductus arteriosus. The patient presented with severe pulmonary
		stenosis and cyanosis. Patient history included a cardiac catheterization and
		echocardiogram. Previous surgeries included Blalock-Taussig shunt and pulmonary
		valvotomy. The patient was not taking any medications. Family history included
		benign hypertension, osteoarthritis, depressive disorder, and extrinsic asthma in
		the grandparent(s).
THYMNON04	PSPORT1	This normalized library was constructed from a thymus tissue library. Starting RNA
		was made from thymus tissue removed from a 3-year-old Caucasian male, who died
		from anoxia. Serologies were negative. The patient was not taking any
		medications. The library was normalized in two rounds using conditions adapted
		from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996)
		6:791, except that a significantly longer (48-hours/round) reannealing
		hybridization was used.
THYRTUT03 DINCY	PINCY	Library was constructed using RNA isolated from benign thyroid tumor tissue
		removed from a 17-year-old Caucasian male during a thyroidectomy. Pathology
		indicated encapsulated follicular adenoma forming a circumscribed mass.

Table 7

Parameter Threshold		Mismatch <50%		ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0E-10 or less	ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater; fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater	Probability value= 1.0E-3 or less	PFAM hits: Probability value=1.0E-3 or less Signal peptide hits: Score=0 or greater
Reference	Applied Biosystems, Foster City, CA.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Applied Biosystems, Foster City, CA.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489.	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, E.L.L. et al. (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350.
Description	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	A program that assembles nucleic acid sequences.	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, tfasta, fastx, tfastx, and ssearch.	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.
Program	ABIFACTURA	ABIPARACEL FDF	ABI AutoAssembler	BLAST	FASTA	BLIMPS	HMMER

Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality score>GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	72.
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Intl. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182.	i. cial 32.
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	;217-221; , page WI.

What is claimed is:

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- 1. An isolated polypeptide selected from the group consisting of:
- a) a polypeptide comprising an amino acid sequence selected from the group consisting of
 SEQ ID NO:1-30,
 - b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-30,
 - c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, and
 - d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30.
 - An isolated polypeptide of claim 1 selected from the group consisting of SEQ ID NO:1 30.

3. An isolated polynucleotide encoding a polypeptide of claim 1.

- 4. An isolated polynucleotide encoding a polypeptide of claim 2.
- 5. An isolated polynucleotide of claim 4 selected from the group consisting of SEQ ID NO:31-60.
 - 6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.
 - 7. A cell transformed with a recombinant polynucleotide of claim 6.
 - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
- 30 9. A method of producing a polypeptide of claim 1, the method comprising:
 - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and

- b) recovering the polypeptide so expressed.
- 10. An isolated antibody which specifically binds to a polypeptide of claim 1.
- 11. An isolated polynucleotide selected from the group consisting of:
- a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60,
- b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:31-60,
 - c) a polynucleotide complementary to a polynucleotide of a),
 - d) a polynucleotide complementary to a polynucleotide of b), and
 - e) an RNA equivalent of a)-d).

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- 12. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 11.
 - 13. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:
 - a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
 - b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.
 - 14. A method of claim 13, wherein the probe comprises at least 60 contiguous nucleotides.
 - 15. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:
- a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
 - b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

16. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.

- 17. A composition of claim 16, wherein the polypeptide has an amino acid sequence selected from the group consisting of SEQ ID NO:1-30.
 - 18. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition of claim 16.

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- 19. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting agonist activity in the sample.

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- 20. A composition comprising an agonist compound identified by a method of claim 19 and a pharmaceutically acceptable excipient.
- 21. A method for treating a disease or condition associated with decreased expression of
 20 functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 20.
 - 22. A method of screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting antagonist activity in the sample.
 - 23. A composition comprising an antagonist compound identified by a method of claim 22 and a pharmaceutically acceptable excipient.

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- 24. A method for treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 23.
 - 25. A method of screening for a compound that specifically binds to the polypeptide of claim

1, the method comprising:

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- a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
- b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a
 compound that specifically binds to the polypeptide of claim 1.
 - 26. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:
- a) combining the polypeptide of claim 1 with at least one test compound under conditions
 permissive for the activity of the polypeptide of claim 1,
 - b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
 - c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
 - 27. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
 - a) exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
 - b) detecting altered expression of the target polynucleotide, and
- c) comparing the expression of the target polynucleotide in the presence of varying amounts
 of the compound and in the absence of the compound.
 - 28. A method of assessing toxicity of a test compound, the method comprising:
 - a) treating a biological sample containing nucleic acids with the test compound,
 - b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 11 under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 11 or fragment thereof,
 - c) quantifying the amount of hybridization complex, and
 - d) comparing the amount of hybridization complex in the treated biological sample with the

amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

- 29. A diagnostic test for a condition or disease associated with the expression of TRICH in a biological sample, the method comprising:
- a) combining the biological sample with an antibody of claim 10, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex, and
- b) detecting the complex, wherein the presence of the complex correlates with the presence
 of the polypeptide in the biological sample.
 - 30. The antibody of claim 10, wherein the antibody is:
 - a) a chimeric antibody,
 - b) a single chain antibody,
- 15 c) a Fab fragment,

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- d) a F(ab')2 fragment, or
- e) a humanized antibody.
- 31. A composition comprising an antibody of claim 10 and an acceptable excipient.
- 32. A method of diagnosing a condition or disease associated with the expression of TRICH in a subject, comprising administering to said subject an effective amount of the composition of claim 31.
- 33. A composition of claim 31, wherein the antibody is labeled.
- 34. A method of diagnosing a condition or disease associated with the expression of TRICH in a subject, comprising administering to said subject an effective amount of the composition of claim 33.
- 35. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 10, the method comprising:
- a) immunizing an animal with a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, or an immunogenic fragment thereof, under conditions to

elicit an antibody response,

- b) isolating antibodies from said animal, and
- c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which binds specifically to a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30.
 - 36. An antibody produced by a method of claim 35.
 - 37. A composition comprising the antibody of claim 36 and a suitable carrier.

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- 38. A method of making a monoclonal antibody with the specificity of the antibody of claim 10, the method comprising:
- a) immunizing an animal with a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
 - b) isolating antibody producing cells from the animal,
- c) fusing the antibody producing cells with immortalized cells to form monoclonal antibodyproducing hybridoma cells,
 - d) culturing the hybridoma cells, and
- e) isolating from the culture monoclonal antibody which binds specifically to a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30.
 - 39. A monoclonal antibody produced by a method of claim 38.
- 25 40. A composition comprising the antibody of claim 39 and a suitable carrier.
 - 41. The antibody of claim 10, wherein the antibody is produced by screening a Fab expression library.
- 42. The antibody of claim 10, wherein the antibody is produced by screening a recombinant immunoglobulin library.
 - 43. A method of detecting a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30 in a sample, the method comprising:

a) incubating the antibody of claim 10 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and

- b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30 in the sample.
- 44. A method of purifying a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30 from a sample, the method comprising:
- a) incubating the antibody of claim 10 with a sample under conditions to allow specific
 binding of the antibody and the polypeptide, and
 - b) separating the antibody from the sample and obtaining the purified polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-30.
 - 45. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
 - 46. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
 - 47. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
- 20 48. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.

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- 49. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.
- 50. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
- 51. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
- 52. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
- 30 53. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
 - 54. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10.
 - 55. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11.

	56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12.
	57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13.
5	58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14.
	59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15.
10	60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16.
10	61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17.
	62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18.
15	63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19.
•	64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20.
20	65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21.
20	66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22.
	67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23.
25	68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24.
	69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25.
20	70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26.
30	71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:27.
	72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:28.

73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:29.

- 74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30.
- 5 75. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:31.
 - 76. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:32.
- 77. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:33.

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- 78. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:34.
 - 79. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:35.
- 20 80. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:36.
 - 81. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:37.
 - 82. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:38.
- 83. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID30 NO:39.
 - 84. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:40.

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- 86. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:42.
 - 87. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:43.
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 - 89. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:45.
 - 90. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:46.

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- 91. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID 20 NO:47.
 - 92. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:48.
- 93. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:49.
 - 94. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:50.
 - 95. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:51.
 - 96. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID

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97. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:53.

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- 98. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:54.
- 99. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID 10 NO:55.
 - 100. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:56.
- 15 101. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:57.
 - $102.\,$ A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:58.

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- $103.\,$ A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:59.
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      TRIBOULEY, Catherine M.
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      LO, Terence P.
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Thr Val Gln His Leu Lys Ser Lys Phe Gly Lys Gly Tyr Phe Leu
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Leu Gln Arg Glu Ile Gln Tyr Ile Phe Pro Asn Ala Ser Arg Gln
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Glu Ser Phe Ser Ser Ile Leu Ala Tyr Lys Ile Pro Lys Glu Asp
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His Ala Phe Ala Ile Glu Glu Tyr Ser Phe Ser Gln Ala Thr Leu
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Glu Gln Val Phe Val Glu Leu Thr Lys Glu Gln Glu Glu Glu Asp
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Pro	Ser	Arg	Asp	Asp 65	Gln	Arg	Leu	Arg	Pro 70	Met	Ala	Pro	Gly	Leu 75
Ser	Glu	Ala	Gly	Lys 80	Leu	Leu	Gly	Leu	Glu 85	Tyr	Pro	Glu	Arg	Gln 90
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Ser	Met	Ser	Ala	Pro 110	Phe	Phe	Leu	Gly	Lys 115	Ile	Ile	Asp	Ala	11e 120
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			Val	155					160					165
			Arg	170					175					180
			Phe	185					190					195
			Asp	200					205					210
			Gly Phe	215					220					225
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			Gly	275					280					285
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			Phe	305					310					315
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			Ile	350					355					360
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Ala	Tyr	Pro	Ala		Pro	Glu	Val	Pro		Phe	Gln	Asp	Phe	
Leu	Ser	Ile	Pro		Gly	Ser	Val	Thr		Leu	Val	Gly	Pro	
Gly	Ser	Gly	Lys		Thr	Val	Leu	Ser		Leu	Leu	Arg	Leu	
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Lys Asn Pro Lys Ile Leu Leu Leu Asp Glu Ala Thr Ser Ala Leu
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Lys Gly Phe Val Ala Phe Gln Ala Ala Ile Asn Ala Ala Ile Ile
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Glu Val Thr Thr Asn His Ser Val Met Glu Glu Leu Thr Ser Val
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Ile Met Asn Glu Trp Phe His Phe Thr Cys Leu Val Ser Phe Ser
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Phe	Trp	Leu	Ser		Gly	Leu	Thr	Tyr	Ile 265	Сув	Phe	Ile	Phe	Ile 270
Met	Ser	Ile	Phe	Met 275	Ala	Leu	Val	Ile	Thr 280	Ser	Ile	Pro	Ile	Val 285
Phe	His	Thr	Gly	Phe 290	Met	Val	Ile	Phe	Thr 295	Leu	Tyr	Ser	Leu	Tyr 300
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			Phe	395					400					405
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			His	605					610					615
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				830					835				Arg	840
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				860					865				Val	870
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				890					895				Gly	900
				905	-				910				Lys	915
		-	_	920					925				Leu	930
	_			935			_		940				Thr	945
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				965					970				Туг	975
				980					985				Ile	990
				995		-	-		1000					1005
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۷al	Leu	Val	Val		Ile	Ile	Gly	Cys		Ala	Ser	Leu	Ile	
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Asn	Gly	Phe	_	Ser L100	Phe	Gly	Phe		11e 1105	Val	Leu	Ile	Cys 1	Val 1110
Ser	Thr	Ile		Val 1115	Ser	Thr	Lys		Glu L120	ГЛS	Pro	Asn	Leu 1	Ile 1125
Leu	Сув	Met		Phe 1130	Ile	Pro	Ser		Thr 1.135	Phe	Leu	Asp	Met	Ser 1140
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Leu	Asp	Asn	Arg	Ile	Asn	Glu	Val	Asn	Lys	Thr	Ile	Leu	Leu	Thr

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				Ile Ser 1370	_			1375		_		1380
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			_	Glu Pro 1415				1420				1425
				Gln Ile 1430				1435				1440
				Leu Thr 1445			_	1450				1455
	-	_		Val Ala 1460				1465				1470
	_			Gln His 1475		-		1480				1485
				Lys Met 1490				1495				1500
				Leu Lys 1505				1510				1515
				Leu Met 1520		-		1525				1530
				Arg Ala 1535				1540				1545
				Glu Glu 1550				1555				1560
				Glu Leu 1565	_	_		1570				1575
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Trp Asp Ala Pro Ala Ile Thr Arg Ser Ser Cys Arg Val Asp Val
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Ser Trp Thr His Gly Gly His Gln Val Asp Val Arg Pro Arg Gly
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Ala Ala Ala Ser Leu Ala Asp Phe Val Glu Asn Val Glu Trp Arg
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Val Leu Gly Met Pro Ala Arg Arg Val Leu Thr Tyr Gly Cys
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Cys Ser Glu Pro Tyr Pro Asp Val Thr Phe Thr Leu Leu Leu Arg
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Arg Arg Ala Ala Ala Tyr Val Cys Asn Leu Leu Leu Pro Cys Val
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Thr Val Phe Gln Leu Leu Leu Ala Glu Ser Met Pro Pro Ala Glu
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Tyr Cys Gly Pro Ser Val Arg Pro Val Pro Ala Trp Ala Arg Ala
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Leu Leu Cly His Leu Ala Arg Gly Leu Cys Val Arg Glu Arg
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Pro Gln Ser Pro Glu Gly Gly Ala Gly Pro Pro Ala Gly Pro Cys
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His Glu Pro Arg Cys Leu Cys Arg Gln Glu Ala Leu Leu His His
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Val Ala Thr Ile Ala Asn Thr Phe Arg Ser His Arg Ala Ala Gln
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Arg Cys His Glu Asp Trp Lys Arg Leu Ala Arg Val Met Asp Arg
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415

Ile Gln Pro Glu Leu Ala Leu Ile Pro Ser Thr Met Asn Cys Met

410

405

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Phe	Leu	Ala	Asn		Thr	Phe	Arg	Gly		Ser	Gly	Ser	Ile	
Val	Lys	Gly	Ser		Ile	Val	Ser	Ser		Asn	Asn	Phe	Phe	
Trp	Asn	Leu	Gln		Asp	Pro	Met	Gly		Pro	Met	Trp	Thr	
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Ser	rys	Leu	His	Leu 515	Arg	Val	Val	Thr	Leu 520	Ile	Glu	His	Pro	Phe 525
V al	Phe	Thr	Arg	Glu 530	Val	Asp	Asp	Glu	Gly 535	Leu	Суѕ	Pro	Ala	Gly 540
Gln	Leu	Сув	Leu	Asp 545	Pro	Met	Thr	Asn	Asp 550	Ser	Ser	Thr	Leu	Asp 555
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			Lys	575					580					585
Lys	Ile	Ala	Glu	Asp 590	Met	Asn	Phe	Asp	Phe 595	Asp	Leu	Tyr	Ile	Val 600
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Asp	Thr	Ala	Ala	Pro 665	Ile	Gly	Ala	Phe	Met 670	Trp	Pro	Leu	His	Trp 675
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			Asn	710					715					720
		_	Tyr	725				_	730					735
		_	Сув	740		_	_		745				_	750
			Met	755					760					765
			Val	770					775					780
			Lys	785					790					795
		_	Glu	800				_	805					810
			His	815					820					825
		_	Val	830	_		_		835			_		840
			Met	845	_				850					855
			Сув	860					865					870
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Phe Ser Gly Leu Phe Val Leu Cys Ile Gly Phe Gly Leu Ser
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Ile Leu Thr Thr Ile Gly Glu His Ile Val Tyr Arg Leu Leu Leu
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Pro Arg Ile Lys Asn Lys Ser Lys Leu Gln Tyr Trp Leu His Thr
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Ser Gln Arg Leu His Arg Ala Ile Asn Thr Ser Phe Ile Glu Glu
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Lys Gln Gln His Phe Lys Thr Lys Arg Val Glu Lys Arg Ser Asn
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Val Gly Pro Arg Gln Leu Thr Val Trp Asn Thr Ser Asn Leu Ser
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His Asp Asn Arg Arg Lys Tyr Ile Phe Ser Asp Glu Glu Gly Gln
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Asn Gln Leu Gly Ile Arg Ile His Gln Asp Ile Pro Leu Pro Pro
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Arg Arg Arg Glu Leu Pro Ala Leu Arg Thr Thr Asn Gly Lys Ala
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Asp Ser Leu Asn Val Ser Arg Asn Ser Val Met Gln Glu Leu Ser
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Glu Leu Glu Lys Gln Ile Gln Val Ile Arg Gln Glu Leu Gln Leu
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Gln Phe Gln Leu Glu Lys Leu Arg Phe Leu Glu Asn Tyr Thr Cys
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Leu Asp Gln Trp Ala Met Glu Gln Phe Val Gln Val Ile Met Glu
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Ala Trp Val Lys Gly Val Asn Pro Lys Gly Asn Ser Thr Asn Pro
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Ser Asn Trp Asp Phe Gly Ser Ser Phe Phe Phe Ala Gly Thr Val
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Val Thr Thr Ile Gly Tyr Gly Asn Leu Ala Pro Ser Thr Glu Ala
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Glu Gly Trp Ser Phe Ser Glu Gly Phe Tyr Phe Ala Phe Ile Thr
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Leu Ser Thr Ile Gly Phe Gly Asp Tyr Val Ala Gly Thr Asp Pro
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Ser Lys His Tyr Ile Ser Val Tyr Arg Ser Leu Ala Ala Ile Trp
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Ile Leu Leu Gly Leu Ala Trp Leu Ala Leu Ile Leu Pro Leu Gly
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Pro Leu Leu His Arg Cys Cys Gln Leu Trp Leu Leu Ser Arg
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Asp Gly Gln Val Leu Val Ala Ala Asp Asp Gly Glu Phe Glu Lys
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Val Glu Asp Arg Lys Gln Asp Leu Gln Gly His Leu Gln Lys Val
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Lys Pro Gln Trp Phe Asn Arg Thr Thr His Trp Ser Phe Leu Ser
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Ser Leu Phe Phe Cys Cys Thr Val Phe Ser Thr Val Gly Tyr Gly
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Tyr Ile Tyr Pro Val Thr Arg Leu Gly Lys Tyr Leu Cys Met Leu
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Tyr Ala Leu Phe Gly Ile Pro Leu Met Phe Leu Val Leu Thr Asp
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Thr Gly Asp Ile Leu Ala Thr Ile Leu Ser Thr Ser Tyr Asn Arg
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Phe Arg Lys Phe Pro Phe Phe Thr Arg Pro Leu Leu Ser Lys Trp
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Cys Pro Lys Ser Leu Phe Lys Lys Pro Asp Pro Lys Pro Ala
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Asp Glu Ala Val Pro Gln Ile Ile Ile Ser Ala Glu Glu Leu Pro
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Gly Pro Lys Leu Gly Thr Cys Pro Ser Arg Pro Ser Cys Ser Met
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Glu Leu Phe Glu Arg Ser His Ala Leu Glu Lys Gln Asn Thr Leu
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Gln Leu Pro Pro Gln Ala Met Glu Arg Ser Asn Ser Cys Pro Glu
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Glu Val Gly Gln Gln Val Glu Arg Leu Asp Ile Pro Leu Pro Ile
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Ile Ala Leu Ile Val Phe Ala Tyr Ile Ser Cys Ala Ala Ala Ile
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                                     295
Leu Pro Phe Trp Glu Thr Gln Leu Asp Phe Glu Asn Ala Phe Tyr
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Phe Cys Phe Val Thr Leu Thr Thr Ile Gly Phe Gly Asp Thr Val
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Leu Glu His Pro Asn Phe Phe Leu Phe Phe Ser Ile Tyr Ile Ile
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Val Gly Met Glu Ile Val Phe Ile Ala Phe Lys Leu Val Gln Asn
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Pro Arg Pro Arg Tyr Val Val Asp Arg Ala Ala Tyr Ser Leu Thr
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Gly Glu Lys Leu Arg Asn Ala Phe Arg Cys Ser Ser Ala Lys Ile
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Lys Tyr Lys Ile Lys Asp Tyr Ile Ile Pro Asp Leu Leu Gly Gly
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                                     115
                                                         120
Leu Ser Gly Gly Ser Ile Gln Val Pro Gln Gly Met Ala Phe Ala
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Leu Leu Ala Asn Leu Pro Ala Val Asn Gly Leu Tyr Ser Ser Phe
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Phe Pro Leu Leu Thr Tyr Phe Phe Leu Gly Gly Val His Gln Met
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Val Pro Gly Thr Phe Ala Val Ile Ser Ile Leu Val Gly Asn Ile
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Cys Leu Gln Leu Ala Pro Glu Ser Lys Phe Gln Val Phe Asn Asn
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Ala Thr Asn Glu Ser Tyr Val Asp Thr Ala Ala Met Glu Ala Glu
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Arg Leu His Val Ser Ala Thr Leu Ala Cys Leu Thr Ala Ile Ile
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                                     220
Gln Met Gly Leu Gly Phe Met Gln Phe Gly Phe Val Ala Ile Tyr
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                                     235
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Leu Ser Glu Ser Phe Ile Arg Gly Phe Met Thr Ala Ala Gly Leu
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Gln	Ile	Leu	Ile		Val	Leu	Lys	Tyr		Phe	Gly	Leu	Thr	255 Ile 270
Pro	Ser	Tyr	Thr		Pro	Gly	Ser	Ile	265 Val 280	Phe	Thr	Phe	Ile	
Ile	Cys	Lys	Asn	275 Leu 290	Pro	His	Thr	Asn		Ala	Ser	Leu	Ile	
Ala	Leu	Ile	Ser		Ala	Phe	Leu	Val		Val	Lys	Glu	Leu	
Ala	Arg	Tyr	Met		Lys	Ile	Arg	Phe		Ile	Pro	Thr	Głu	
Ile	Val	Va1	Val		Ala	Thr	Ala	Ile	Ser 340	Gly	Gly	Сув	Lys	Met 345
Pro	Lys	Lys	Tyr	His 350	Met	Gln	Ile	Val	Gly 355	Glu	Ile	Gln	Arg	Gly 360
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				590					595				Gln	600 Ser
Glu	Pro	Pro	Ala	605 Ser	Ala	Glu	Ala	Pro		Glu	Pro	Ser	Asp	
Leu	Ala	Ser	Va1		Pro	Phe	Val	Thr		His	Thr	Leu	Ile	
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Leu	Ala	Lys	Leu		Ser	Thr	Tyr	Gly			Gly	Val	Lys	
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Gly	Gly	Val	Phe	680 Glu 695	Asp	Gly	Ser	Leu	685 Glu 700	Cys	Lys	His	Val	
Pro	Ser	Ile	His		Ala	Val	Leu	Phe		Gln	Ala	Asn	Ala	
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Gly Gly Ala Gly Ala Lys Glu His Gly Asn Ser Val Cys Phe Lys
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Ala Glu Gly Pro Arg Arg Gln Tyr Gly Phe Met Gln Arg Gln Phe
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Thr Ser Met Leu Gln Pro Gly Val Asn Lys Phe Ser Leu Arg Met
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Phe Gly Ser Gln Lys Ala Val Glu Lys Glu Gln Glu Arg Val Lys
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Thr Ala Gly Phe Trp Ile Ile His Pro Tyr Ser Asp Phe Arg Phe
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Tyr Trp Asp Leu Ile Met Leu Ile Met Met Val Gly Asn Leu Val
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Ile Ile Pro Val Gly Ile Thr Phe Phe Thr Glu Gln Thr Thr
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Pro Trp Ile Ile Phe Asn Val Ala Ser Asp Thr Val Phe Leu Leu
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Asp Leu Ile Met Asn Phe Arg Thr Gly Thr Val Asn Glu Asp Ser
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Ser Glu Ile Ile Leu Asp Pro Lys Val Ile Lys Met Asn Tyr Leu
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Lys Ser Trp Phe Val Val Asp Phe Ile Ser Ser Ile Pro Val Asp
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Tyr Ile Phe Leu Ile Val Glu Lys Gly Met Asp Ser Glu Val Tyr
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Lys Thr Ala Arg Ala Leu Arg Ile Val Arg Phe Thr Lys Ile Leu
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Ser Leu Leu Arg Leu Leu Arg Leu Ser Arg Leu Ile Arg Tyr Ile
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Ala Val Val Arg Ile Phe Asn Leu Ile Gly Met Met Leu Leu Leu
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Cys His Trp Asp Gly Cys Leu Gln Phe Leu Val Pro Leu Leu Gln
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Asp Phe Pro Pro Asp Cys Trp Val Ser Leu Asn Glu Met Val Asn
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Ser	His	Met	Leu	Cys 350	Ile	Gly	Tyr	Gly	Ala 355	Gln	Ala	Pro	Val	Ser 360
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Thr	Суѕ	Tyr	Ala		Phe	Va1	G1y	His		Thr	Ala	Leu	Ile	
Ser	Leu	Asp	Ser		Arg	Arg	Gln	Tyr		Glu	Lys	Tyr	Lys	Gln 405
Va1	Glu	Gln	Tyr		Ser	Phe	His	Lys		Pro	Ala	Asp	Met	Arg 420
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Phe	Asp	Glu	Glu	Asn 440	Ile	Leu	Asn	Glu	Leu 445	Asn	Asp	Pro	Leu	Arg 450
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Arg	Glu	Gly	Ala	Val 500	Gly	Lys	Lys	Met	Tyr 505	Phe	Ile	Gln	His	Gly 510
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			Lys	590					595					600
			Gly	605					610					615
			Lys	620	_	_			625					630
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			Ser	665					670					675
			Thr	680					685					690
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	_		Phe	710					715					720
			Gln	725					730					735
			Gln	740					745					750
_			Thr	755					760					765
			Thr	770			_		775					780
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Thr Val Pro Gln Arg Val Thr Leu Phe Arg Gln Met Ser Ser Gly
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Ala Ile Pro Pro Asn Arg Gly Val Pro Pro Ala Pro Pro Pro Pro
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Pro Lys Val Leu Gly Ser Val Asp Gln Phe Asn Asp Ser Gly Leu
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Asn Lys Met Ala Leu Ala Ser Phe Met Lys Gly Arg Thr Val Ile
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Gly Thr Pro Asp Glu Glu Thr Met Asp Ile Glu Leu Pro Lys Lys
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Tyr His Glu Met Val Gly Val Ile Phe Ser Asp Thr Phe Ser Tyr
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Arg Leu Lys Phe Asn Trp Gly Tyr Arg Ile Pro Val Ile Lys Glu
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His Ser Glu Tyr Thr Gly His Cys Trp Ala Met His Gly Glu Ile
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Phe Cys Tyr Leu Ala Lys Tyr Trp Leu Lys Gly Phe Val Ala Phe
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Gln Ala Ala Ile Asn Ala Ala Ile Ile Glu Val Thr Thr Asn His
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Ser Val Met Glu Glu Leu Thr Ser Val Ile Gly Ile Asn Met Lys
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Ile Pro Pro Phe Ile Ser Lys Gly Glu Ile Met Asn Glu Trp Phe
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His Phe Thr Cys Leu Val Ser Phe Ser Ser Phe Ile Tyr Phe Ala
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Ser Leu Asn Val Ala Arg Glu Arg Gly Lys Phe Lys Lys Leu Met
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Thr Val Met Gly Leu Arg Glu Ser Ala Phe Trp Leu Ser Trp Gly
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Leu Thr Tyr Ile Cys Phe Ile Phe Ile Met Ser Ile Phe Met Ala
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                                     280
Leu Val Ile Thr Ser Ile Pro Ile Val Phe His Thr Gly Phe Met
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                290
Val Ile Phe Thr Leu Tyr Ser Leu Tyr Gly Leu Ser Leu Val Ala
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Phe	Thr	Val	Leu		Arg	Gln	Leu	Pro		Ser	Leu	Gly	Trp	Val 360
Leu	Ser	Leu	Leu	Ser 365	Pro	Phe	Ala	Phe	Thr 370	Ala	Gly	Met	Ala	Gln 375
Ile	Thr	His	Leu		Asn	Tyr	Leu	Ser	Gly 385	Val	Ile	Phe	Pro	Asp 390
Pro	Ser	Gly	Asp	Ser	Tyr	Lys	Met	Ile	Ala 400	Thr	Phe	Phe	Ile	Leu 405
Ala	Phe	Asp	Thr	Leu 410	Phe	Tyr	Leu	Ile	Phe 415	Thr	Leu	Tyr	Phe	Glu 420
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Thr	Ile	Tyr	Asn	Thr 545	Gln	Leu	Ser	Glu	11e 550	Thr	Asp	Met	Glu	Glu 555
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Lys	Gly	Ile	Gln	Pro 590	Lys	Glu	Val	Glu	Gln 595	Glu	Val	Leu	Leu	Leu 600
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				620			Lys		625					630
				635			Ala		640					645
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Arg	Asn	Glu	Met	Cys 680	Asp	Thr	Glu	Lys	Ile 685	Thr	Ser	Leu	Ile	Lys 690
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Leu	Tyr	Ser	Asp		Asp	Lys	Сув	Ser	Asp 730		Gly	Ile	Arg	Asn 735
Tyr	Ala	Val	Ser		Thr	Ser	Leu	Asn		Val	Phe	Leu	Asn	
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Leu	Leu	Leu	Val	Leu 830	Gly	Ile	Ala	Phe		Pro	Ile	Ile	Leu	
Lys	Ile	Met	Tyr	Lys 845	Val	Thr	Arg	Gļu		His	Cys	Trp	G1u	
Ser	Pro	Ser	Met	Tyr	Phe	Leu	Ser	Leu		Gln	Ile	Pro	Lys	
Pro	Leu	Thr	Ser	860 Leu	Leu	Ile	Val	Asn		Thr	Gly	Ser	Asn	
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Ile	qaA	Asp	Phe	Arg 905	Asn	Arg	Asn	Gly		Asp	Asp	Pro	Ser	-
Asn	Gly	Ala	Ile	11e 920	Val	Ser	Gly	Asp		Lys	Asp	Tyr	Arg	
Ser	Val	Ala	Суз	Asn 935	Thr	Lys	Lys	Leu		Суз	Phe	Pro	Val	
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Trp	Pro	Ser		Тут 1010	Trp	Суз	Gly	Gln		Leu	Val	Asp	Ile	
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Val	Val	Cys		11e	Gly	Суѕ	Ala		Ser 1060	Leu	Ile	Phe		Thr 1065
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Ser	Phe	His	-	Asn 1100	Val	Ser	Arg	_	Asp 1105	Phe	Leu	Phe		Phe L110
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Суз	Phe	Ser		Arg L220	Lys	Lys	Lys		Ala 1225	Ile	Arg	Asn		Ser L230
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Tyr Ser Ser Leu Met Ala Tyr Lys Leu Pro Val Glu Asp Val His
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Pro Leu Ser Arg Ala Phe Phe Lys Leu Glu Ala Met Lys Gln Thr
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Gly Leu Val Ile Gly Ile Ile Leu Ala Ile Ala Tyr Ser Leu Leu
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Gly	Ile	Phe	Ser	Leu 125	Leu	Сув	Leu	Met	Val 130	Gly	Gln	Val	Val	Asp 135
Arg	Glu	Leu	Gln	Leu 140	Ala	Gly	Phe	qaA	Pro 145	Ser	Gln	Asp	Gly	Leu 150
Gln	Pro	Gly	Ala	Asn 155	Ser	Ser	Thr	Leu	Asn 160	Gly	Ser	Ala	Ala	Met 165
Leu	Asp	Суз	Gly	Arg 170	Ąsp	Cys	Tyr	Ala	Ile 175	Arg	Val	Ala	Thr	Ala 180
Leu	Thr	Leu	Met	Thr 185	Gly	Leu	Tyr	Gln	Val 190	Leu	Met	Gly	Val	Leu 195
Arg	Leu	Gly	Phe	Val 200	Ser	Ala	Tyr	Leu	Ser 205	Gln	Pro	Leu	Leu	Asp 210
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				Gly 230					235					240
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				Asp 260					265					270
				Lys 275				-	280					285
_				Pro 290					295					300
				Phe 305					310	_		_		315
		_	_	11e 320			_		325					330
				335					340	_				345
				Ala 350					355					360
				Gly 365 Cys					370					375
			•	380 Ala					385					390
				395 Gln					400					405
				410 Leu					415					420
				425 Ala					430			_		435
				440 Trp					445					450
	_	_		455 Val	_			_	460	-	_			465
	_			470 Ala	_		_		475			_		480
				485 Gly	_				490					495
				500 Asp					505					510
				515 Pro					520					525
				530 Tyr					535					540
OT.Y	110	A.G.U.	-1.	545	111G	2 1174.4	~ <u>,</u> 5	ւաբ	550	1116	Leu	للندن	501	555

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Tyr Ser Leu Thr Gly Leu Asp Ala Gly Cys Met Ala Ala Arg Arg
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                560
Lys Glu Gly Gly Ser Glu Thr Gly Val Gly Glu Gly Pro Ala
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                575
Gln Gly Glu Asp Leu Gly Pro Val Ser Thr Arg Ala Ala Leu Val
                590
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Pro Ala Ala Ala Gly Phe His Thr Val Val Ile Asp Cys Ala Pro
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Leu Leu Phe Leu Asp Ala Ala Gly Val Ser Thr Leu Gln Asp Leu
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Arg Arg Asp Tyr Gly Ala Leu Gly Ile Ser Leu Leu Leu Ala Cys
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Cys Ser Pro Pro Val Arg Asp Ile Leu Ser Arg Gly Gly Phe Leu
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Gly Glu Gly Pro Gly Asp Thr Ala Glu Glu Glu Gln Leu Phe Leu
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                                      40
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Ser Val Leu Asp Leu Trp Ala Ala Cys Leu Tyr Arg Ser Cys Leu
                                      55
                 50
Leu Leu Gly Ala Thr Ile Gly Val Ala Lys Asn Ser Ala Leu Gly
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                                      70
Pro Arg Arg Leu Arg Ala Ser Trp Leu Val Ile Thr Leu Val Cys
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                                      85
Leu Phe Val Gly Ile Tyr Ala Met Val Lys Leu Leu Phe Ser
                                                         105
                                    100
                 95
Glu Val Arg Arg Pro Ile Arg Asp Pro Trp Phe Trp Ala Leu Phe
                110
                                     115
                                                         120
Val Trp Thr Tyr Ile Ser Leu Gly Ala Ser Phe Leu Leu Trp Trp
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                                     130
Leu Leu Ser Thr Val Arg Pro Gly Thr Gln Ala Leu Glu Pro Gly
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                140
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Ala Ala Thr Glu Ala Glu Gly Phe Pro Gly Ser Gly Arg Pro Pro
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                                    160
                                                         165
Pro Glu Gln Ala Ser Gly Ala Thr Leu Gln Lys Leu Leu Ser Tyr
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                                    175
                                                         180
Thr Lys Pro Asp Val Ala Phe Leu Val Ala Ala Ser Phe Phe Leu
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                                    190
                                                         195
Ile Val Ala Ala Leu Gly Glu Thr Phe Leu Pro Tyr Tyr Thr Gly
                200
                                     205
                                                         210
Arg Ala Ile Asp Gly Ile Val Ile Gln Lys Ser Met Asp Gln Phe
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                                     220
Ser Thr Ala Val Val Ile Val Cys Leu Leu Ala Ile Gly Ser Ser
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Phe	Ala	Ala	Gly	Ile 245	Arg	Gly	Gly	Ile	Phe 250	Thr	Leu	Ile	Phe	Ala 255
Arg	Leu	Asn	Ile		Leu	Arg	Asn	Cys	Leu 265	Phe	Arg	Ser	Leu	Val 270
Ser	Gln	G1u	Thr	Ser 275	Phe	Phe	Asp	Glu	Asn 280	Arg	Thr	Gly	Asp	Leu 285
Ile	Ser	Arg	Leu	Thr 290	Ser	Asp	Thr	Thr	Met 295	Val	Ser	Asp	Leu	Val 300
Ser	Gln	Asn	Ile	Asn 305	Val	Phe	Leu	Arg	Asn 310	Thr	Val	Lys	Val	Thr 315
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			Met	335					340					345
			Тух	350					355					360
			Ala	365					370					375
-			Arg	380					385					390
			Lys	395					400					405
			Tyr	410					415					420
			Gln	425					430·					435
			Gln	440					445					450
			Val	455					460					465
			Leu	470					475					480
			Asp	485					490					495
			His	500					505					510
			Arg	515					520					525
			Leu	530					535					540
			Gly Glu	545					550					555
			Asp	560					565					570
				575					580					585 Ile
				590					595					600 Ala
			Ala	605					610					615
				620					625					630 Gly
			Gln	635					640					645
				650					655					660 Ala
				665					670					675 Gln
				680					685					690 Glu
				695					700					705 Gln
urs	ATG	uis	ьeu	ттв	val	val	nen	usb	пÃр	GTĀ	AL Y	val	val	GIII

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715
Gln Gly Thr His Gln Gln Leu Leu Ala Gln Gly Gly Leu Tyr Ala
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                                     730
                725
Lys Leu Val Gln Arg Gln Met Leu Gly Leu Gln Pro Ala Ala Asp
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Phe Thr Ala Gly His Asn Glu Pro Val Ala Asn Gly Ser His Lys
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Arg Pro Val Ala Asp Thr Asp Gln Thr Leu Asn Val Thr Leu Glu
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Val Thr Leu Ser Gln Ile Ile Asp Met Asp Glu Arg Asn Gln Val
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Leu Thr Leu Tyr Leu Trp Ile Arg Gln Glu Trp Thr Asp Ala Tyr
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                 80
Leu Arg Trp Asp Pro Asn Ala Tyr Gly Gly Leu Asp Ala Ile Arg
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Ile Pro Ser Ser Leu Val Trp Arg Pro Asp Ile Val Leu Tyr Asn
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                                     115
                110
Lys Ala Asp Ala Gln Pro Pro Gly Ser Ala Ser Thr Asn Val Val
                                     130
                125
Leu Arg His Asp Gly Ala Val Arg Trp Asp Ala Pro Ala Ile Thr
                                                         150
                140
                                     145
Arg Ser Ser Cys Arg Val Asp Val Ala Ala Phe Pro Phe Asp Ala
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                155
                                     160
Gln His Cys Gly Leu Thr Phe Gly Ser Trp Thr His Gly Gly His
                                     175
                 170
Gln Leu Asp Val Arg Pro Arg Gly Ala Ala Ala Ser Leu Ala Asp
                                                         195
                                     190
                185
Phe Val Glu Asn Val Glu Trp Arg Val Leu Gly Met Pro Ala Arg
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Arg Arg Val Leu Thr Tyr Gly Cys Cys Ser Glu Pro Tyr Pro Asp
                                     220
                 215
Val Thr Phe Thr Leu Leu Leu Arg Arg Arg Ala Ala Tyr Val
                                                          240
                 230
                                     235
Cys Asn Leu Leu Pro Cys Val Leu Ile Ser Leu Leu Ala Pro
                 245
                                     250
Leu Ala Phe His Leu Pro Ala Asp Ser Gly Glu Lys Val Ser Leu
                                     265
                 260
Gly Val Thr Val Leu Leu Ala Leu Thr Val Phe Gln Leu Leu
                 275
                                     280
                                                          285
Ala Glu Ser Met Pro Pro Ala Glu Ser Val Pro Leu Ile Gly Lys
                                     295
                 290
Tyr Tyr Met Ala Thr Met Thr Met Val Thr Phe Ser Thr Ala Leu
                                     310
Thr Ile Leu Ile Met Asn Leu His Tyr Cys Gly Pro Ser Val Arg
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320
Pro Val Pro Ala Trp Ala Arg Ala Leu Leu Leu Gly His Leu Ala
                                     340
                335
Arg Gly Leu Cys Val Arg Glu Arg Gly Glu Pro Cys Gly Gln Ser
                350
                                     355
Arg Pro Pro Glu Leu Ser Pro Ser Pro Gln Ser Pro Glu Gly Gly
                                                         375
                365
                                     370
Ala Gly Pro Pro Ala Gly Pro Cys His Glu Pro Arg Cys Leu Cys
                                                         390
Arg Gln Glu Ala Leu Leu His His Val Ala Thr Ile Ala Asn Thr
                                     400
                395
Phe Arg Ser His Arg Ala Ala Gln Arg Cys His Glu Asp Trp Lys
                410
                                     415
                                                         420
Arg Leu Ala Arg Val Met Asp Arg Phe Phe Leu Ala Ile Phe Phe
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Ser Met Ala Leu Val Met Ser Leu Leu Val Leu Val Gln Ala Leu
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Leu Thr Phe Thr Glu Lys Trp Asn Thr Asp Asn Thr Leu Gly Thr
                                      55
                 50
Glu Ile Thr Val Glu Asp Gln Leu Ala Arg Gly Leu Lys Leu Thr
                                      70
                 65
Phe Asp Ser Ser Phe Ser Pro Asn Thr Gly Lys Lys Asn Ala Lys
                 80
                                      85
                                                          90
Ile Lys Thr Gly Tyr Lys Arg Glu His Ile Asn Leu Gly Cys Asp
                 95
                                     100
Met Asp Phe Asp Ile Ala Gly Pro Ser Ile Arg Gly Ala Leu Val
                110
                                     115
Leu Gly Tyr Glu Gly Trp Leu Ala Gly Tyr Gln Met Asn Phe Glu
                                     130
Thr Ala Lys Ser Arg Val Thr Gln Ser Asn Phe Ala Val Gly Tyr
                                     145
                140
Lys Thr Asp Glu Phe Gln Leu His Thr Asn Val Asn Asp Gly Thr
                                     160
                155
Glu Phe Gly Gly Ser Ile Tyr Gln Lys Val Asn Lys Lys Leu Glu
                170
                                     175
Thr Ala Val Asn Leu Ala Trp Thr Ala Gly Asn Ser Asn Thr Arg
                                     190
                185
Phe Gly Ile Ala Ala Lys Tyr Gln Ile Asp Pro Asp Ala Cys Phe
                200
                                     205
                                                         210
Ser Ala Lys Val Asn Asn Ser Ser Leu Ile Gly Leu Gly Tyr Thr
                                     220
                215
Gln Thr Leu Lys Pro Gly Ile Lys Leu Thr Leu Ser Ala Leu Leu
                                     235
                230
                                                         240
Asp Gly Lys Asn Val Asn Ala Gly Gly His Lys Leu Gly Leu Gly
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250

245

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365

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Phe Ile Gly Leu Gln Thr Leu Gly Leu Thr Leu Lys Arg Cys Tyr
                380
                                     385
                                                         390
Arg Glu Met Val Met Leu Leu Val Phe Ile Cys Val Ala Met Ala
                                     400
                395
Ile Phe Ser Ala Leu Ser Gln Leu Leu Glu His Gly Leu Asp Leu
                410
                                     415
Glu Thr Ser Asn Lys Asp Phe Thr Ser Ile Pro Ala Ala Cys Trp
                                     430
                425
Trp Val Ile Ile Ser Met Thr Thr Val Gly Tyr Gly Asp Met Tyr
                440
                                     445
                                                         450
Pro Ile Thr Val Pro Gly Arg Ile Leu Gly Gly Val Cys Val Val
                                     460
                455
Ser Gly Ile Val Leu Leu Ala Leu Pro Ile Thr Phe Ile Tyr His
                470
                                    475
Ser Phe Val Gln Cys Tyr His Glu Leu Lys Phe Arg Ser Ala Arg
                485
                                     490
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Tyr Ser Arg Ser Leu Ser Thr Glu Phe Leu Asn
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Pro Thr Lys Gln Ala Ala Leu Lys Ser His Tyr Ala Asp Val Asp
                                      40
Pro Glu Asn Gln Asn Phe Leu Leu Glu Ser Asn Leu Gly Lys Lys
                 50
                                      55
Lys Tyr Glu Thr Glu Phe His Pro Gly Thr Thr Ser Phe Gly Met
                                     70
                 65
Ser Val Phe Asn Leu Ser Asn Ala Ile Val Gly Ser Gly Ile Leu
                 80
                                     85
                                                          90
Gly Leu Ser Tyr Ala Met Ala Asn Thr Gly Ile Ala Leu Phe Ile
                 95
                                     100
Ile Leu Leu Thr Phe Val Ser Ile Phe Ser Leu Tyr Ser Val His
                                     115
                110
Leu Leu Lys Thr Ala Asn Glu Gly Gly Ser Leu Leu Tyr Glu
                125
                                     130
                                                         135
Gln Leu Gly Tyr Lys Ala Phe Gly Leu Val Gly Lys Leu Ala Ala
                140
                                     145
Ser Gly Ser Ile Thr Met Gln Asn Ile Gly Ala Met Ser Ser Tyr
                                     160
                155
Leu Phe Ile Val Lys Tyr Glu Leu Pro Leu Val Ile Gln Ala Leu
                170
                                     175
Thr Asn Ile Glu Asp Lys Thr Gly Leu Trp Tyr Leu Asn Gly Asn
                185
                                     190
Tyr Leu Val Leu Leu Val Ser Leu Val Val Ile Leu Pro Leu Ser
                200
                                     205
Leu Phe Arg Asn Leu Gly Tyr Leu Gly Tyr Thr Ser Gly Leu Ser
                                     220
                215
Leu Leu Cys Met Val Phe Phe Leu Ile Val Val Ile Cys Lys Lys
                                                         240
                230
                                     235
Phe Gln Val Pro Cys Pro Val Glu Ala Ala Leu Ile Ile Asn Glu
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Thr Ile Asn Thr Thr Leu Thr Gln Pro Thr Ala Leu Val Pro Ala
                260
                                     265
Leu Ser His Asn Val Thr Glu Asn Asp Ser Cys Arg Pro His Tyr
                275
                                     280
                                                         285
Phe Ile Phe Asn Ser Gln Thr Val Tyr Ala Val Pro Ile Leu Ile
                                     295
                290
Phe Ser Phe Val Cys His Pro Ala Val Leu Pro Ile Tyr Glu Glu
                                                         315
                305
                                     310
Leu Lys Asp Arg Ser Arg Arg Arg Met Met Asn Val Ser Lys Ile
                                     325
                320
Ser Phe Phe Ala Met Phe Leu Met Tyr Leu Leu Ala Ala Leu Phe
                                     340
                335
Gly Tyr Leu Thr Phe Tyr Glu His Val Glu Ser Glu Leu Leu His
                                     355
                350
Thr Tyr Ser Ser Ile Leu Gly Thr Asp Ile Leu Leu Leu Ile Val
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                365
Arg Leu Ala Val Leu Met Ala Val Thr Leu Thr Val Pro Val Val
                                                         390
                380
                                     385
Ile Phe Pro Ile Arg Ser Ser Val Thr His Leu Leu Cys Ala Ser
                395
                                     400
Lys Asp Phe Ser Trp Trp Arg His Ser Leu Ile Thr Val Ser Ile
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                410
Leu Ala Phe Thr Asn Leu Leu Val Ile Phe Val Pro Thr Ile Arg
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Asp Ile Phe Gly Phe Ile Gly Ala Ser Ala Ala Ser Met Leu Ile
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Phe Ile Leu Pro Ser Ala Phe Tyr Ile Lys Leu Val Lys Lys Glu
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                                     460
Pro Met Lys Ser Val Gln Lys Ile Gly Ala Leu Phe Phe Leu Leu
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Asp Trp Val His Asn Ala Pro Gly Gly His
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Ala Ala Val Ser Lys Thr Ala Val Ala Pro Ile Glu Arg Val Lys
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Leu Leu Leu Gln Val Gln Ala Ser Ser Lys Gln Ile Ser Pro Glu
                                      55
                 50
Ala Arg Tyr Lys Gly Met Val Asp Cys Leu Val Arg Ile Pro Arg
                                      70
                                                          75
                 65
Glu Gln Gly Phe Phe Ser Phe Trp Arg Gly Asn Leu Ala Asn Val
                 80
                                      85
Ile Arg Tyr Phe Pro Thr Gln Ala Leu Asn Phe Ala Phe Lys Asp
                                     100
                 95
Lys Tyr Lys Gln Leu Phe Met Ser Gly Val Asn Lys Glu Lys Gln
                                     115
                                                         120
Phe Trp Arg Trp Phe Leu Ala Asn Leu Ala Ser Gly Gly Ala Ala
                125
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Gly Ala Thr Ser Leu Cys Val Val Tyr Pro Leu Asp Phe Ala Arg
                                     145
                140
Thr Arg Leu Gly Val Asp Ile Gly Lys Gly Pro Glu Glu Arg Gln
                155
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Phe Lys Gly Leu Gly Asp Cys Ile Met Lys Ile Ala Lys Ser Asp
                                     175
                170
Gly Ile Ala Gly Leu Tyr Gln Gly Phe Gly Val Ser Val Gln Gly
                                                         195
                                     190
                185
Ile Ile Val Tyr Arg Ala Ser Tyr Phe Gly Ala Tyr Asp Thr Val
                                     205
                200
Lys Gly Leu Leu Pro Lys Pro Lys Lys Thr Pro Phe Leu Val Ser
                                     220
                215
Phe Phe Ile Ala Gln Val Val Thr Thr Cys Ser Gly Ile Leu Ser
                                     235
                230
Tyr Pro Phe Asp Thr Val Arg Arg Arg Met Met Gln Ser Gly
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                245
Glu Ala Lys Arg Gln Tyr Lys Gly Thr Leu Asp Cys Phe Val Lys
                260
                                     265
                                                          270
Ile Tyr Gln His Glu Gly Ile Ser Ser Phe Phe Arg Gly Ala Phe
                275
                                     280
Ser Asn Val Leu Arg Gly Thr Gly Gly Ala Leu Val Leu Val Leu
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Tyr Asp Lys Ile Lys Glu Phe Phe His Ile Asp Ile Gly Gly Arg
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Leu Pro Glu Ser Leu Thr Ser Leu Leu Val Leu Leu Thr Thr Ala
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Asn Asn Pro Asp Val Met Ile Pro Ala Tyr Ser Lys Asn Arg Ala
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                                      70
Tyr Ala Ile Phe Phe Ile Val Phe Thr Val Ile Gly Ser Leu Phe
                  80
                                      85
Leu Met Asn Leu Leu Thr Ala Ile Ile Tyr Ser Gln Phe Arg Gly
                 95
                                     100
Tyr Leu Met Lys Ser Leu Gln Thr Ser Leu Phe Arg Arg Leu
                                     115
                 110
                                                          120
Gly Thr Arg Ala Ala Phe Glu Val Leu Ser Ser Met Val Gly Glu
                 125
                                     130
                                                          135
Gly Gly Ala Phe Pro Gln Ala Val Gly Val Lys Pro Gln Asn Leu
                                     145
                140
Leu Gln Val Leu Gln Lys Val Gln Leu Asp Ser Ser His Lys Gln
                 155
                                     160
Ala Met Met Glu Lys Val Arg Ser Tyr Asp Ser Val Leu Leu Ser
                 170
                                                          180
Ala Glu Glu Phe Gln Lys Leu Phe Asn Glu Leu Asp Arg Ser Val
                                     190
                 185
Val Lys Glu His Pro Pro Arg Pro Glu Tyr Gln Ser Pro Phe Leu
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205

220

225

Gln Ser Ala Gln Phe Leu Phe Gly His Tyr Tyr Phe Asp Tyr Leu

200

215

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Gly Asn Leu Ile Ala Leu Ala Asn Leu Val Ser Ile Cys Val Phe
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                                    235
Leu Val Leu Asp Ala Asp Val Leu Pro Ala Glu Arg Asp Asp Phe
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                                    250
Ile Leu Gly Ile Leu Asn Cys Val Phe Ile Val Tyr Tyr Leu Leu
                260
                                    265
Glu Met Leu Leu Lys Val Phe Ala Leu Gly Leu Arg Gly Tyr Leu
                                    280
                275
Ser Tyr Pro Ser Asn Val Phe Asp Gly Leu Leu Thr Val Val Leu
                                    295
                290
Leu Val Leu Glu Ile Ser Thr Leu Ala Val Tyr Arg Leu Pro His
                                    310
                305
Pro Gly Trp Arg Pro Glu Met Val Gly Leu Leu Ser Leu Trp Asp
                320
                                    325
                                                         330
Met Thr Arg Met Leu Asn Met Leu Ile Val Phe Arg Phe Leu Arg
                335
                                    340
Ile Ile Pro Ser Met Lys Pro Met Ala Val Val Ala Ser Thr Val
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                                    355
                350
Leu Gly Leu Val Gln Asn Met Arg Ala Phe Gly Gly Ile Leu Val
                                    370
Val Val Tyr Tyr Val Phe Ala Ile Ile Gly Ile Asn Leu Phe Arg
                380
                                    385
Gly Val Ile Val Ala Leu Pro Gly Asn Ser Ser Leu Ala Pro Ala
                                    400
                395
Asn Gly Ser Ala Pro Cys Gly Ser Phe Glu Gln Leu Glu Tyr Trp
                410
                                    415
                                                         420
Ala Asn Asn Phe Asp Asp Phe Ala Ala Ala Leu Val Thr Leu Trp
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Asn Leu Met Val Val Asn Asn Trp Gln Val Phe Leu Asp Ala Tyr
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                                    445
Arg Arg Tyr Ser Gly Pro Trp Ser Lys Ile Tyr Phe Val Leu Trp
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Trp Leu Val Ser Ser Val Ile Trp Val Asn Leu Phe Leu Ala Leu
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                470
Ile Leu Glu Asn Phe Leu His Lys Trp Asp Pro Arg Ser His Leu
                                    490
                485
Gln Pro Leu Ala Gly Thr Pro Glu Ala Thr Tyr Gln Met Thr Val
                500
                                    505
                                                         510
Glu Leu Leu Phe Arg Asp Ile Leu Glu Glu Pro Gly Glu Asp Glu
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Leu Thr Glu Arg Leu Ser Gln His Pro His Leu Trp Leu Cys Arg
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Ala Lys Val Met Val Glu Lys Ser Arg Met Gly Val Val Pro Pro
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Gly Thr Gln Val Glu Gln Leu Leu Tyr Ala Lys Lys Leu Tyr Asp
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                                                          75
                 65
Ser Ala Phe His Pro Asp Thr Gly Glu Lys Met Asn Val Ile Gly
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                                      85
                                                          90
Arg Met Ser Phe Gln Leu Pro Gly Gly Met Ile Ile Thr Gly Phe
                                     100
                 95
Met Leu Gln Phe Tyr Arg Thr Met Pro Ala Val Ile Phe Trp Gln
                110
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Trp Val Asn Gln Ser Phe Asn Ala Leu Val Asn Tyr Thr Asn Arg
                125
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Asn Ala Ala Ser Pro Thr Ser Val Arg Gln Met Ala Leu Ser Tyr
                140
                                     145
                                                         150
Phe Thr Ala Thr Thr Ala Val Ala Thr Ala Val Gly Met Asn
                155
                                     160
Met Leu Thr Lys Lys Ala Pro Pro Leu Val Gly Arg Trp Val Pro
                                     175
                                                         180
                170
Phe Ala Ala Val Ala Ala Ala Asn Cys Val Asn Ile Pro Met Met
                185
                                     190
                                                         195
Arg Gln Gln Glu Leu Ile Lys Gly Ile Cys Val Lys Asp Arg Asn
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Glu Asn Glu Ile Gly His Ser Arg Arg Ala Ala Ala Ile Gly Ile
                215
                                     220
Thr Gln Val Val Ile Ser Arg Ile Thr Met Ser Ala Pro Gly Met
                230
                                     235
Ile Leu Leu Pro Val Ile Met Glu Arg Leu Glu Lys Leu His Phe
                245
                                     250
Met Gln Lys Val Lys Val Leu His Ala Pro Leu Gln Val Met Leu
                                                         270
                260
                                     265
Ser Gly Cys Phe Leu Ile Phe Met Val Pro Val Ala Cys Gly Leu
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Ile Leu Phe Val Thr Pro Leu Leu Leu Leu Pro Leu Val Ile Leu
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Met Pro Ala Lys Phe Val Arg Cys Ala Tyr Val Ile Ile Leu Met
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                                      40
Ala Ile Tyr Trp Cys Thr Glu Val Ile Pro Leu Ala Val Thr Ser
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Leu Met Pro Val Leu Leu Phe Pro Leu Phe Gln Ile Leu Asp Ser
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Arg Gln Val Cys Val Gln Tyr Met Lys Asp Thr Asn Met Leu Phe
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Leu Gly Gly Leu Ile Val Ala Val Ala Val Glu Arg Trp Asn Leu
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His	Lvs	Ara	Ile	95 Ala	Leu	Ara	Thr	Leu	100 Leu	Tro	Val	Glv	Ala	105 Lvs
				110					115					120
				125					130				Leu	135
Ser	Met	Trp	Ile	Ser 140	Asn	Thr	Ala	Thr	Thr 145	Ala	Met	Met	Val	Pro 150
Ile	Val	Glu	Ala		Leu	Gln	Gln	Met		Ala	Thr	Ser	Ala	Ala 165
Thr	Glu	Ala	Gly		Glu	Leu	Val	Asp		Gly	Lys	Ala	Lys	
Leu	Pro	Ala	Asn		Ala	Val	Pro	Thr		Gly	Ser	Gln	Val	
Phe	Glu	Gly	Pro		Leu	Gly	Gln	Gln		Asp	Gln	Glu	Arg	
Arg	Leu	Cys	Lys		Met	Thr	Leu	Cys		Cys	Tyr	Ala	Ala	
Ile	Gly	Gly	Thr		Thr	Leu	Thr	Gly		Gly	Pro	Asn	Va1	
Leu	Leu	Gly	Gln		Asn	Glu	Leu	Phe		Asp	Ser	Lys	Asp	
Val	Asn	Phe	Ala		Trp	Phe	Ala	Phe		Phe	Pro	Asn	Met	
Val	Met	Leu	Leu		Ala	Trp	Leu	Trp		Gln	Phe	Val	Tyr	
Arg	Phe	Asn	Phe		Lys	Ser	Trp	Gly		Gly	Leu	Glu	Ser	
Lys	Asn	Glu	Lys		Ala	Leu	Lys	Val		Gln	Glu	Glu	Tyr	
Lys	Leu	Gly	Pro		Ser	Phe	Ala	Glu		Asn	Val	Leu	Ile	
Phe	Phe	Leu	Leu		Ile	Leu	Trp	Phe	Ser	Arg	Asp	Pro	Gly	Phe 345
Met	Pro	Gly	Trp		Thr	Val	Ala	Trp	Va1 355	Glu	Glu	Arg	Lys	Thr 360
Pro	Phe	Tyr	Pro		Pro	Leu	Leu	qaA		Lys	Val	Thr	Gln	
Lys	Val	Pro	Trp		Ile	Val	Leu	Leu		Gly	Gly	Gly	Phe	
Leu	Ala	Lys	Gly		G1u	Ala	Ser	Gly		Ser	Va1	Trp	Met	
Lys	Gln	Met	Glu		Leu	His	Ala	Va1		Pro	Ala	Ala	Ile	
Leu	Ile	Leu	Ser		Leu	Val	Ala	Val		Thr	Glu	Сув	Thr	
Asn	Val	Ala	Thr		Thr	Leu	Phe	Leu		Ile	Phe	Ala	Ser	
Ser	Arg	Ser	Ile		Leu	Asn	Pro	Leu		Ile	Met	Leu	Pro	
Thr	Leu	Ser	Ala		Phe	Ala	Phe	Met		Pro	Val	Ala	Thr	
Pro	Asn	Ala	Ile		Phe	Thr	Tyr	Gly		Leu	Lys	Va1	Ala	
Met	Val	Lys	Thr		Val	Ile	Met	Asn		Ile	Gly	Val	Phe	
Val	Phe	Leu	Ala		Asn	Thr	Trp	Gly		Ala	Ile	Phe	Asp	
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Phe Phe Leu Ser Phe Cys Phe Tyr Phe Phe Tyr Asn Ile Thr Leu

.410

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Glu Gly Ile Ala Ile Phe Leu Leu Arg Pro Ser Asp Leu Gln Ser
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Ile Leu Ser Asp Ala Trp Phe His Phe Val Phe Phe Ile Gln Ala
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Val Leu Val Ile Leu Ser Val Phe Leu Tyr Leu Phe Ala Tyr Lys
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Glu Tyr Leu Ala Cys Leu Val Leu Ala Met Ala Leu Gly Trp Ala
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Asn Met Leu Tyr Tyr Thr Arg Gly Phe Gln Ser Met Gly Met Tyr
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Ser Val Met Ile Gln Lys Val Ile Leu His Asp Val Leu Lys Phe
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Leu Phe Val Tyr Ile Val Phe Leu Leu Gly Phe Gly Val Ala Leu
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Ala Ser Leu Ile Glu Lys Cys Pro Lys Asp Asn Lys Asp Cys Ser
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Ser Tyr Gly Ser Phe Ser Asp Ala Val Leu Glu Leu Phe Lys Leu
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Thr Ile Gly Leu Gly Asp Leu Asn Ile Gln Gln Asn Ser Lys Tyr
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                                    610
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Pro Ile Leu Phe Leu Phe Leu Leu Ile Thr Tyr Val Ile Leu Thr
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Phe Val Leu Leu Asn Met Leu Ile Ala Leu Met Gly Glu Thr
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Val Glu Asn Val Ser Lys Glu Ser Glu Arg Ile Trp Arg Leu Gln
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Arg Ala Arg Thr Ile Leu Glu Phe Glu Lys Met Leu Pro Glu Trp
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                                    670
Leu Arg Ser Arg Phe Arg Met Gly Glu Leu Cys Lys Val Ala Glu
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Asp Asp Phe Arg Leu Cys Leu Arg Ile Asn Glu Val Lys Trp Thr
                695
                                    700
Glu Trp Lys Thr His Val Ser Phe Leu Asn Glu Asp Pro Gly Pro
                710
                                    715
Val Arg Arg Thr Asp Phe Asn Lys Ile Gln Asp Ser Ser Arg Asn
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Arg Gly Leu Tyr Arg Gly Thr Ser Pro Ala Leu Leu Ala Tyr Val
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Thr Gln Gly Ser Val Leu Phe Met Cys Phe Gly Phe Cys Gln Gln
                                      85
                 80
Phe Val Arg Lys Val Ala Arg Val Glu Gln Asn Ala Glu Leu Asn
                 95
                                     100
Asp Leu Glu Thr Ala Thr Ala Gly Ser Leu Ala Ser Ala Phe Ala
                110
                                     115
Ala Leu Ala Leu Cys Pro Thr Glu Leu Val Lys Cys Arg Leu Gln
                125
                                     130
Thr Met Tyr Glu Met Lys Met Ser Gly Lys Ile Ala Gln Ser Tyr
                                     145
                140
Asn Thr Ile Trp Ser Met Val Lys Ser Ile Phe Met Lys Asp Gly
                                     160
                155
                                                         165
Pro Leu Gly Phe Tyr Arg Gly Leu Ser Thr Thr Leu Ala Gln Glu
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                                     175
                                                         180
Ile Pro Gly Tyr Phe Phe Tyr Phe Gly Gly Tyr Glu Ile Ser Arg
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                                     190
Ser Phe Phe Ala Ser Gly Gly Ser Lys Asp Glu Leu Gly Pro Val
                                     205
                200
Pro Leu Met Leu Ser Gly Gly Phe Ala Gly Ile Cys Leu Trp Leu
                                     220
                                                         225
Ile Ile Phe Pro Val Asp Cys Ile Lys Ser Arg Ile Gln Val Leu
                230
                                     235
Ser Met Phe Gly Lys Pro Ala Gly Leu Ile Glu Thr Phe Ile Ser
                                                         255
                245
                                     250
Val Val Arg Asn Glu Gly Ile Ser Ala Leu Tyr Ser Gly Leu Lys
                260
                                     265
                                                         270
Ala Thr Leu Ile Arg Ala Ile Pro Ser Asn Ala Ala Leu Phe Leu
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Val Tyr Glu Tyr Ser Arg Lys Met Met Met Asn Met Val Glu Glu
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Pro Gly Pro Gly Pro Ser Asp Ser Pro Glu Ala Ala Val Glu Lys
                                      40
                 35
Val Glu Val Glu Leu Ala Gly Pro Ala Thr Ala Glu Pro His Glu
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Pro Pro Glu Pro Pro Glu Gly Gly Trp Gly Trp Leu Val Met Leu
                                      70
                 65
Ala Ala Met Trp Cys Asn Gly Ser Val Phe Gly Ile Gln Asn Ala
                 80
                                      85
Cys Gly Val Leu Phe Val Ser Met Leu Glu Thr Phe Gly Ser Lys
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Asp Asp Asp Lys Met Val Phe Lys Thr Ala Trp Val Gly Ser Leu
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                                     115
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Thr Asp Leu Phe Gly Cys Arg Lys Thr Ala Val Val Gly Ala Ala
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Val Gly Phe Val Gly Leu Met Ser Ser Phe Val Ser Ser Ile
                155
                                     160
Glu Pro Leu Tyr Leu Thr Tyr Gly Ile Ile Phe Ala Cys Gly Cys
                                     175
                                                         180
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Ser Phe Ala Tyr Gln Pro Ser Leu Val Ile Leu Gly His Tyr Phe
                185
                                     190
Lys Lys Arg Leu Gly Leu Val Asn Gly Ile Val Thr Ala Gly Ser
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                                     205
Ser Val Phe Thr Ile Leu Leu Pro Leu Leu Leu Arg Val Leu Ile
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                                     220
Asp Ser Val Gly Leu Phe Tyr Thr Leu Arg Val Leu Cys Ile Phe
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                                     235
Met Phe Val Leu Phe Leu Ala Gly Phe Thr Tyr Arg Pro Leu Ala
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Thr Ser Thr Lys Asp Lys Glu Ser Gly Gly Ser Gly Ser Ser Leu
                260
                                     265
Phe Ser Arg Lys Lys Phe Ser Pro Pro Lys Lys Ile Phe Asn Phe
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                                     280
Ala Ile Phe Lys Val Thr Ala Tyr Ala Val Trp Ala Val Gly Ile
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                                     295
Pro Leu Ala Leu Phe Gly Tyr Phe Val Pro Tyr Val His Leu Met
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Lys His Val Asn Glu Arg Phe Gln Asp Glu Lys Asn Lys Glu Val
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                                     325
Val Leu Met Cys Ile Gly Val Thr Ser Gly Val Gly Arg Leu Leu
                                     340
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Phe Gly Arg Ile Ala Asp Tyr Val Pro Gly Val Lys Lys Val Tyr
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Leu Gln Val Leu Ser Phe Phe Phe Ile Gly Leu Met Ser Met Met
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Ile Pro Leu Cys Ser Ile Phe Gly Ala Leu Ile Ala Val Cys Leu
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Ile Met Gly Leu Phe Asp Gly Cys Phe Ile Ser Ile Met Ala Pro
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                                     400
Ile Ala Phe Glu Leu Val Gly Ala Gln Asp Val Ser Gln Ala Ile
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Gly Phe Leu Gly Phe Met Ser Ile Pro Met Thr Val Gly Pro
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Pro Ile Ala Gly Leu Leu Arg Asp Lys Leu Gly Ser Tyr Asp Val
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Ala Phe Tyr Leu Ala Gly Val Pro Pro Leu Ile Gly Gly Ala Val
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                                     460
Leu Cys Phe Ile Pro Trp Ile His Ser Lys Lys Gln Arg Glu Ile
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                                     475
Ser Lys Thr Thr Gly Lys Glu Lys Met Glu Lys Met Leu Glu Asn
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Gln Asn Ser Leu Leu Ser Ser Ser Ser Gly Met Phe Lys Lys Glu
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Arg Val Val Phe Pro Asn Asn Ser Ile Phe His Gln Asp Trp Glu
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Glu Val Ser Arg Arg Tyr Pro Gly Asn Arg Thr Cys Thr Thr Lys
                                      70
                                                           75
Tyr Thr Leu Phe Thr Phe Leu Pro Arg Asn Leu Phe Glu Gln Phe
                                      85
His Arg Trp Ala Asn Leu Tyr Phe Leu Phe Leu Val Ile Leu Ser
                 95
                                     100
Trp Met Pro Ser Met Glu Val Phe His Arg Glu Ile Thr Met Leu
                110
                                     115
Pro Leu Ala Ile Val Leu Phe Val Ile Met Ile Lys Asp Gly Met
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Glu Asp Phe Lys Arg His Arg Phe Asp Lys Ala Ile Asn Cys Ser
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Asn Ile Arg Ile Tyr Glu Arg Lys Glu Gln Thr Tyr Val Gln Lys
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Cys Trp Lys Asp Val Arg Val Gly Asp Phe Ile Gln Met Lys Cys
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Asn Glu Ile Val Pro Ala Asp Ile Leu Leu Leu Phe Ser Ser Asp
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                                     190
Pro Asn Gly Ile Cys His Leu Glu Thr Ala Ser Leu Asp Gly Glu
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                                     205
Thr Asn Leu Lys Gln Arg Arg Val Val Lys Gly Phe Ser Gln Gln
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                215
                                     220
Glu Val Gln Phe Glu Pro Glu Leu Phe His Asn Thr Ile Val Cys
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                                     235
Glu Lys Pro Asn Asn His Leu Asn Lys Phe Lys Gly Tyr Met Glu
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                                     250
His Pro Asp Gln Thr Arg Thr Gly Phe Gly Cys Glu Ser Leu Leu
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Leu Arg Gly Cys Thr Ile Arg Asn Thr Glu Met Ala Val Gly Ile
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                                     280
Val Ile Tyr Ala Gly His Glu Thr Lys Ala Met Leu Asn Asn Ser
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                                     295
Gly Pro Arg Tyr Lys Arg Ser Lys Ile Glu Arg Arg Met Asn Ile
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                                     310
Asp Ile Phe Phe Cys Ile Gly Ile Leu Ile Leu Met Cys Leu Ile
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                                     325
Gly Ala Val Gly His Ser Ile Trp Asn Gly Thr Phe Glu Glu His
                                     340
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Pro Pro Phe Asp Val Pro Asp Ala Asn Gly Ser Phe Leu Pro Ser
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Ala Leu Gly Gly Phe Tyr Met Phe Leu Thr Met Ile Ile Leu Leu
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                                     370
Gln Val Leu Ile Pro Ile Ser Leu Tyr Val Ser Ile Glu Leu Val
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Lys Leu Gly Gln Val Phe Phe Leu Ser Asn Asp Leu Asp Leu Tyr
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                                     400
                                                          405
Asp Glu Glu Thr Asp Leu Ser Ile Gln Cys Arg Ala Leu Asn Ile
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                                     415
                                                         420
Ala Glu Asp Leu Gly Gln Ile Gln Tyr Ile Phe Ser Asp Lys Thr
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                                     430
                                                         435
Gly Thr Leu Thr Glu Asn Lys Met Val Phe Arg Arg Cys Thr Ile
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                                     445
Met Gly Ser Glu Tyr Ser His Gln Glu Asn Ala Lys Arg Leu Glu
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Thr	Pro	Lys	Glu	Leu 470	Asp	Ser	Asp	Gly	Glu 475	Glu	Trp	Thr	Gln	Tyr 480
Gln	Cys	Leu	Ser		Ser	Ala	Arg	Trp		Gln	Asp	Pro	Ala	
Met	Arg	Ser	Gln		Gly	Ala	Gln	Pro		Arg	Arg	Ser	Gln	
Ala	Arg	Val	Pro		Gln	Gly	His	Tyr		Gln	Arg	Ser	Met	_
His	Arg	Glu	Ser		Gln	Pro	Pro	Val		Phe	Ser	Ser	Ser	
Glu	Lys	Asp	Val	Thr	Pro	Asp	Lys	Asn		Leu	Thr	Lys	Val	
Asp	Ala	Ala	Leu		Leu	Glu	Thr	Leu	Ser	Asp	Ser	Arg	Pro	
ГЛЗ	Ala	Ser	Leu		Thr	Thr	Ser	Ser		Ala	Asp	Phe	Phe	
Ala	Leu	Thr	Ile		Asn	Ser	Val	Met		Ser	Thr	Thr	Thr	
Pro	Arg	Gln	Arg		Asp	Asp	Gln	Lys		Val	Glu	Asn	Asp	
Cys	Gln	Cys	Бeu		Phe	Gln	Gly	Trp	610 Arg 625	Lys	Ile	Ser	Gly	
Thr	Tyr	Суз	Lys		Thr	Phe	Ile	Phe	Arg	Ile	Arg	Gln	Leu	
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His	Lys	Val	Thr	Ile	Lys	Pro	Ser	Ser		Ala	Leu	Gly	Thr	
Leu	Glu	Lys	Ile	665 Gln 680	Gln	Leu	Phe	Gln		Leu	Lys	Leu	Leu	
Leu	Ser	Gln	Ser		Ser	Ser	Thr	Ala		Ser	Asp	Thr	Asp	
Gly	Glu	Ser	Leu		Ala	Asn	V al	Ala		Thr	Asp	Ser	Asp	
Arg	Asp	Asp	Ala		Val	Сув	Ser	Gly		Asp	Ser	Thr	Asp	
Gly	Gly	Tyr	Arg		Ser	Met	Trp	Asp		Gly	Asp	Ile	Leu	
Ser	Gly	Ser	Gly		Ser	Leu	Glu	Glu		Leu	Glu	Ala	Pro	
Thr	Asp	Leu	Ala	Arg 770	Pro	Glu	Phe	Суѕ	Tyr 775	Glu	Ala	Glu	Ser	Pro 780
Asp	Glu	Ala	Ala	Leu 785	۷al	His	Ala	Ala	His 790	Ala	Tyr	Ser	Phe	Thr 795
Leu	Val	Ser	Arg	Thr 800	Pro	Glu	Gln	Val	Thr 805	Val	Arg	Leu	Pro	Gln 810
Gly	Thr	Сув	Leu	Thr 815	Phe	Ser	Leu	Leu	Cys 820	Thr	Leu	Gly	Phe	
Ser	Val	Arg	Lys	Arg 830	Met	Ser	Val	Val	Val 835	Arg	His	Pro	Leu	Thr 840
Gly	Glu	Ile	Val	Val 845	Tyr	Thr	Lys	Gly	Ala 850	Asp	Ser	Val	Ile	Met 855
Asp	Leu	Leu	Glu	Asp 860	Pro	Ala	Cys	Val	Pro 865	Asp	Ile	Asn	Met	Glu 870
Lys	Lys	Leu	Arg	Lys 875	Ile	Arg	Ala	Arg	Thr 880	Gln	Lys	His	Leu	Asp 885
Leu	Tyr	Ala	Arg		Gly	Leu	Arg	Thr		Суз	Ile	Ala	Lys	Lys 900
Val	Val	Ser	Glu		Asp	Phe	Arg	Arg	Trp 910	Ala	Ser	Phe	Arg	Arg 915
Glu	Ala	Glu	Ala		Leu	Asp	Asn	Arg	_	Glu	Leu	Leu	Met	
Thr	Ala	Gln	His		Glu	Asn	Gln	Leu		Leu	Leu	Gly	Ala	

Glv	T1a	Glu	Asp	935 Ara	T.eu	Gln	Glu	Glv	940 Val	Pro	Aso	Тhr	Tle	945 Ala
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				965					970					975
			Thr	980					985					990
Asn	Gln	Thr	Asp	Thr 995	Va1	Tyr	Thr		Asn L000	Thr	Glu	Asn		Glu LOO5
Thr	Cys	Glu	Ser	Ile L010	Leu	Asn	Суз		Leu 1015	Glu	Glu	Leu		Gln 1020
Phe	Arg	Glu	Leu		Lys	Pro	Asp		Lys 1030	Leu	Phe	Gly		Arg L035
Leu	Pro	Ser	Lys		Pro	Ser	Ile		Ser 1045	Glu	Ala	Val		Pro LO50
Glu	Ala	Gly	Leu		Ile	Asp	Gly	Lys		Leu	Asn	Ala	Ile	
Gln	Gly	Lys	Leu		Lys	Lys	Phe	Leu		Leu	Thr	Gln	Tyr	
Arg	Ser	Val	Leu		Суз	Arg	Ser	Thr		Leu	Gln	Lys	Ser	
Ile	Val	Lys	Leu		Arg	Asp	Lys	Leu		Val	Met	Thr	Leu	
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Ser	Ser	qaA	Phe		Ile	Thr	Arg	Phe		His	Leu	Lys	Lys	
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Val	Va1	Tyr	Tyr		Tyr	Lys	Asn	Val		Tyr	Val	Asn	Leu	
Phe	Trp	Tyr	Gln		Phe	Сув	Gly	Phe		Ser	Ser	Thr	Met	
Asp	Tyr	Trp	${\tt Gln}$		Ile	Phe	Phe	Asn		Phe	Phe	Thr	Ser	
Pro	Pro	Leu	Val		Gly	Val	Leu	Asp		Asp	Ile	Ser	Ala	
Thr	Leu	Leu	Ala		Pro	Glu	Leu	Tyr		Ser	Gly	Gln	Asn	
Glu	Cys	Тух	Asn		Ser	Thr	Phe	Trp		Ser	Met	Val	Asp	
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Lys	Gly	Ser	Asp	Ile 1280	Asp	Val	Phe	Thr		Gly	Thr	Pro	Ile	
Thr	Ile	Ser	Leu		Thr	Ile	Leu	Leu		Gln	Ala	Met	Glu	
Lys	Thr	Trp	Thr		Phe	His	Gly	Val		Leu	Leu	Gly	Ser	
Leu	Met	Tyr	Phe		Val	Ser	Leu	Leu		Asn	Ala	Thr	Cys	
Ile	Суз	Asn	Ser		Thr	Asn	Pro	Tyr		Val	Met	Glu	Gly	
Leu	Ser	Asn	Pro		Phe	Tyr	Leu	Val		Phe	Leu	Thr	Pro	
Val	Ala	Leu	Leu		Arg	Tyr	Phe	Phe	Leu	Ser	Leu	Gln	Gly	
Cys	Gly	Lys	Ser	Leu	Ile	Ser	Lys	Ala		Lys	Ile	Asp	Lys	
Pro	Pro	Asp	Lys		Asn	Leu	Glu	Ile		Ser	Trp	Arg	Ser	Arg
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WO 02/12340
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Pro Val Ser Ser Ile Thr Gly Gln Asp Phe Ser Ala Ser Thr Pro
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Lys Ser Ser Asn Pro Pro Lys Arg Lys His Val Glu Glu Ser Val
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Leu His Glu Gln Arg Cys Gly Thr Glu Cys Met Arg Asp Asp Ser
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                                    1465
Cys Ser Gly Asp Ser Ser Ala Gln Leu Ser Ser Gly Glu His Leu
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Leu Gly Pro Asn Arg Ile Met Ala Tyr Ser Gly Gly Gln Thr Asp
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